

European Respiratory Society Statement on

Familial Pulmonary Fibrosis

Supplementary materials

Telomere length analysis methods

Quantitative polymerase chain reaction (Q-PCR)

The QPCR determination of telomere lengths was first described by Cawton et al, 2002 [1]. It consists of measuring telomere signal (T) normalized to a reference single-gene copy signal (S). The measure provides a T/S ratio [2].

Advantages: easily available, cheap, fast, from extracted DNA, low amount of DNA needed (50 ng), easy to apply to large populations

Disadvantages: amplification step (PCR), poor reproducibility, intra-assay variation >10%, no information on the “shortest telomere” [3].

Terminal restriction fragment analysis

It consists of measuring TL with a TTAGGG-labelled probe by a southern blot. It measures the intensity of a telomere smear and thus provides average TL.

Advantages: reproducible, applicable to several tissues, quantitative, No amplification step

Disadvantages: long, difficult to apply to large populations, large amount of DNA required (3 µg), difficulty to detect very short telomeres, results may vary based on the restriction enzymes used.

Single TEломere Length Analysis (STELA)

Single TEломere Length Analysis or STELA consists of the application of single molecule PCR to generate highly accurate telomere measures from a small amount of starting material. STELA takes advantage of the fact that telomeres end with a single stranded 3' G-rich overhang, used as a template that allows ligating an oligonucleotide linker to the 5' end of the telomere. A linker-specific primer is then used in with a primer specific for a unique subtelomeric sequence and generates a

single amplicon for each telomere. The main drawback is that not all chromosome ends have suitable sequence for the design of unique chromosome arm primers, and thus STELA is usually restricted to well characterized ends; XpYp, 2p, 11q, 12q and 17p. To minimize PCR artifacts, sub-visible amplification is conducted, and STELA products are resolved by agarose gel electrophoresis, Southern blotted and probed with the specific subtelomeric sequence.

Advantage: requires very few cells (>50), does not require specific material, powerful in research for relative and absolute TL measure, applicable to several tissues

Disadvantages: requires high expertise, not standardized at this point, not validated for clinical purposes.

(Q-FISH and) Flow-FISH

Interphase quantitative fluorescence in situ hybridization (Q-FISH) consists of determining telomere fluorescent intensity after hybridization with a fluorescent peptide nucleic acid telomeric repeat. The technique may be applied to metaphase, but only on proliferating cells.

Flow-FISH is a similar method where fluorescence is measured by fluorescent-accelerated cell signal (FACS) [4]. Flow FISH now offers an extensive quantitative reference data related to telomere length currently available and is the first of the telomere length methods to have been validated for clinical diagnostic purposes. Flow-FISH is feasible on nucleated cells, which means that the material (DNA) is readily available. Flow-FISH is currently the most reliable and easily applicable method to measure telomere length.

Advantages: reproducible, several samples handled at the same time, possible to implement in clinical practice

Disadvantages: requires expertise, necessity of a control population curve, not applicable in patients with blood involvement of telomere syndrome, requires living cells (blood) (<48h)

Perspectives in TL techniques

New techniques are currently developed.

- Luminex assay
- WGS for TL measurement

Comparison of TL measure

Although several reviews [5, 6] on TL measurement methods exist, only few teams directly compared those methods on a similar population [7–9]. Some data exist on direct comparison between TL measure by Q-PCR and Flow-FISH: all studies demonstrate that Flow-FISH is more reliable, reproducible and yields the best sensitivity and specificity.

Supplementary Figure PRISMA flow diagram of literature selection.

Methods for the Literature search:

Databases: PubMed; Embase.com

Limits: publication date: no

Language: no

Study types: no

Age: no

Exclude animal studies and conference abstracts

Search: 15 september 2020.

search in New PubMed

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((("Pulmonary Fibrosis"[Mesh] OR "Pulmonary Fibros*"[tiab] OR "Lung Fibros*"[tiab] OR "Hamman-Rich Syndrome"[tiab] OR "familial interstitial pneumoni*"[tiab] OR "Cryptogenic Organizing Pneumonia*"[tiab] OR "Lung Diseases, Interstitial"[Mesh>NoExp] OR "Alveolitis, Extrinsic Allergic"[Mesh>NoExp] OR "Idiopathic Interstitial Pneumonias"[Mesh>NoExp] OR "idiopathic Interstitial Pneumoni*"[tiab] OR ((("interstitial lung disease*"[tiab] OR "ILD"[ti] OR "ILDs"[ti]) NOT ("adult"[mesh] NOT ("infant"[mesh] OR "child"[mesh] OR "adolescent"[mesh]))) OR "hypersensitivity pneumoni*"[tiab])) AND ("Genes"[Mesh>NoExp] OR "Gene"[tiab] OR "Genes"[tiab] OR "Genetics"[Mesh>NoExp] OR "Human Genetics"[Mesh>NoExp] OR "Genetics, Medical"[Mesh] OR "Genetic*"[tiab] OR "Mutation"[Mesh>NoExp] OR "mutation*"[tiab] OR "Mucin-5B"[Mesh] OR "Mucin-5B"[tiab] OR "MUC5B"[tiab] OR "rs35705950"[tiab] OR "Telomerase"[Mesh] OR "telomerase*"[tiab] OR "STELA"[tiab] OR "Telomere*"[tiab] OR "familial*"[tiab] OR "hereditary"[tiab] OR "inheritance*"[tiab] OR "Anticipation, Genetic"[Mesh] OR "Penetrance"[Mesh] OR "penetrance*"[tiab] OR "SFTPA1"[tiab] OR "SFTPA2"[tiab] OR "Surfactant Metabolism Dysfunction, Pulmonary, 1" [Supplementary Concept] OR "SFTPB"[tiab] OR "Surfactant Metabolism Dysfunction, Pulmonary, 2" [Supplementary Concept] OR "SFTPC"[tiab] OR "Surfactant Metabolism Dysfunction, Pulmonary, 3" [Supplementary Concept] OR "ABCA3"[tiab] OR "AP3B1"[tiab] OR "NKX2-1"[tiab] OR "COPA"[tiab] OR "Surfactant Metabolism Dysfunction, Pulmonary, 4" [Supplementary Concept] OR "CSF2RA"[tiab] OR "CSF2RB"[tiab] OR "FARSB"[tiab] OR "FLNA"[tiab] OR "GATA2"[tiab] OR
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"SLC7A7"[tiab] OR "MARS"[tiab] OR "STAT3"[tiab] OR "STING"[tiab] OR "TBX4"[tiab] OR
"Hermanski-Pudlak Syndrome"[Mesh] OR "Hermanski-Pudlak"[tiab])) OR ((("Pulmonary Surfactant-
Associated Proteins/deficiency"[Mesh] OR ("surfactant"[tiab] AND "protein*"[tiab] AND
"deficienc*"[tiab])) AND ("Lung Diseases"[Mesh:NoExp] OR "Respiration Disorders"[Mesh:NoExp] OR
"Respiratory Distress Syndrome, Newborn"[Mesh] OR "respiratory failure"[tiab] OR "respiratory
distress"[tiab])) NOT ("animals"[Mesh] NOT "humans"[mesh])

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(('lung fibrosis'/de OR 'Pulmonary Fibros*':ti,ab,kw OR 'Lung Fibros*':ti,ab,kw OR 'Hamman-Rich
Syndrome':ti,ab,kw OR 'familial interstitial pneumoni*':ti,ab,kw OR 'Cryptogenic Organizing
Pneumonia*':ti,ab,kw OR 'interstitial lung disease'/de OR 'allergic pneumonitis'/de OR 'interstitial
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OR 'CSF2RA':ti,ab,kw OR 'CSF2RB':ti,ab,kw OR 'FARSB':ti,ab,kw OR 'FLNA':ti,ab,kw OR
'GATA2':ti,ab,kw OR 'SLC7A7':ti,ab,kw OR 'MARS':ti,ab,kw OR 'STAT3':ti,ab,kw OR 'STING':ti,ab,kw
OR 'TBX4':ti,ab,kw OR 'ocular albinism'/exp OR 'Hermanski-Pudlak':ti,ab,kw)) OR (((('surfactant
associated protein'/de OR 'surfactant':ti,ab,kw) AND 'protein*':ti,ab,kw AND 'deficienc*':ti,ab,kw)
AND ('lung disease'/de OR 'breathing disorder'/de OR 'neonatal respiratory distress syndrome'/de OR
'respiratory failure':ti,ab,kw OR 'respiratory distress':ti,ab,kw)) NOT 'conference abstract'/it NOT
(('animal experiment'/exp OR 'animal model'/exp OR 'nonhuman'/exp) NOT 'human'/exp)

Table S1 Other genes associated with ILD

Disease	Genes	Mode of inheritance	Age of presentation pulmonary symptoms	Non ILD pulmonary and Extra pulmonary phenotype	Fre-quency	Most frequent pattern	Implication for management/ Therapy for pulmonary disease	References
Hermansky–Pudlak syndrome	<i>HPS1</i>	AR	30-40yo 2 – 15 y	-Albinism - Spontaneous bleeding	rare	Unclassifiable pulmonary fibrosis	Unknown: Antifibrotic drugs? Lung transplantation may be considered	[10, 11]
	<i>AP3B1 / HSP2</i>	AR						
	<i>HPS4</i>	AR						
Interferonopathy	<i>STING1/TMEM173</i>	AD	Infancy- young adult	Vasculopathy with onset in infancy, Auto-inflammatory features (SAVI)	rare	Unclassifiable pulmonary fibrosis , Alveolar hemorrhage	Unknown: Antifibrotic drugs ? Jak inhibitor?	[12, 13] [14]
	<i>COPA</i>	AD		Arthralgia, kidney disease				
	<i>OAS1</i>	AD		Fever, dermatitis, inflammatory bowel disease,	Ultrarare	Alveolar proteinosis	Allogeneic stem cell transplantation?	[15–17]
Interferonopathy	<i>ZNFX1</i>	AR	Infancy, children	Viral infections, Inflammatory episodes, early-onset seizures, renal disease	Ultrarare	Unclassifiable pulmonary fibrosis	Jak inhibitor?	[18]

Aminoacyl-tRNA synthetases	<i>MARS</i>	AR	Infancy- young adult (Founder effect Reunion Island)	Anemia, hepatomegaly, feeding difficulties, failure to thrive and hypoalbuminemia	Rare	Pulmonary alveolar proteinosis, Pulmonary fibrosis	Methionine supplementation? Whole lung lavage?	[19, 20]
	<i>FARS1</i>	AR	Infancy, childhood	Neurological findings, liver dysfunction, and connective tissue, muscular and vascular abnormalities.	Ultra rare	Interstitial lung disease with cholesterol pneumonitis	Unknown	[21]
GM-CSF receptor	<i>CSF2RA</i>	AR	Infancy, childhood, adults		Ultra rare	Alveolar proteinosis	Whole lung lavage? Autologous transplantation of genetically corrected macrophages? GM-CSF? Stem cell transplantation?	[22]
	<i>CSF2RB</i>	AR	Infancy, childhood, adults		Ultra rare	Alveolar proteinosis	Whole lung lavage? Stem cell transplantation?	[23]
Fibrosis, neurodegeneration, and cerebral angiomatosis (FINCA)	<i>NHLRC2</i>	AR	Infancy- young adult	Neurodegeneration and cerebral angiomatosis	Ultra rare	desquamative interstitial pneumonia; nonspecific interstitial pneumonia	Unknown	[24]

Acid Sphingomyelinase Deficiency (ASMD, Niemann–Pick disease)	<i>SMPD1</i>	AR	Type A / 3yo Type B : later onset	Hepatomegaly Splenomegaly Thrombocytopenia	rare	Not Fibrosing Interstitial lung disease with ground glass opacities	Enzyme replacement therapy to be confirmed	[25, 26]
Niemann–Pick disease, Type C	<i>NPC1, NPC2</i>	AR	Type C	ILD, PAP, Hepato-splenomegaly	rare	Unclassifiable pulmonary fibrosis, Alveolar Proteinosis	Unknown	[27]
GATA2 deficiency	<i>GATA2</i>	AR	Adult	monoMac syndrome: monocytopenia, Mycobacterial infection Myelodysplastic syndrome	Rare	Unclassifiable pulmonary fibrosis, Alveolar Proteinosis	Stem cell transplantation	[28, 29]
Pulmonary alveolar microlithiasis	<i>SLC34A2</i>	AR	5-41 yo		Ultra rare	Not Fibrosing sandstorm-like	Lung transplantation may be considered	[30]
Poikiloderma lung fibrosis	<i>FAM111B</i>	AD	young	Hereditary Fibrosing Poikiloderma with Tendon Contractures, Myopathy, Exocrine pancreatic dysfunction,	Ultra rare	Unclassifiable pulmonary fibrosis	Unknown	[31, 32]

				pancreatic cancer				
Polidase deficiency	<i>PEPD</i>	AR	young	Mental retardation, facial dysmorphism, dermatologic manifestations including ulcerations	Ultra rare	Unclassifiable pulmonary fibrosis	Unknown	[33, 34]
Lysinuric protein intolerance	<i>SLC7A7</i>	AR	Infancy- young adult	metabolic S: vomiting, diarrhea, failure to thrive, hepatomegaly, diffuse cirrhosis, low blood urea, hyperammonemia, and leukopenia	Ultra rare	Alveolar proteinosis	Specific diet	[35, 36]
Mitochondrial respiratory chain complex deficiency : Fanconi renotubular syndrome 5	<i>NDUFAF6</i>	AR	2-40 yo	renotubular syndrome, interstitial renal fibrosis; pulmonary microlithiasis	Ultra rare	Unclassifiable pulmonary fibrosis	Unknown	[37]
Werner	<i>WRN</i>	AR	>10-yo 55 yo	scleroderma-like skin changes, cataract, subcutaneous calcification, premature arteriosclerosis,	Ultra rare	Unclassifiable pulmonary fibrosis	Unknown	[38, 39]

				diabetes mellitus, and premature aged facies				
Calcium pathway?	<i>S100A13/</i> <i>S100A3</i>	AR (Founder effect Saudi Arabia)	Young : 12- 15yo		Ultra rare	Unclassifiable pulmonary fibrosis	Unknown	[40]

Table S2 Result of the survey for Familial Pulmonary Fibrosis (FPFFPF) definition

How should we call it? *	n=13 3 (21.3%) 3 (32.3%) 10 (76.9%)
What does the definition of FPF require?*	n=13 13 (100%) 0 (0%) 0 (0%) 9 (69.2%)
What degree of relationship still define FPF?*	n=13 11 (84.6%) 12 (92.3%) 3 (23.1%) 1 (7.7%)
What kind of ILD should be included in the definition of FPF? *	n=13 3 (23.1%) 4 (30.8%) 2 (15.4%) 1 (7.7%) 7 (53.8%)
For instance, do you include the following? *	n=12 7 (58.3%) 10 (83.3%) 6 (50%) 11 (91.7%) 5 (41.7%) 8 (66.7%) 8 (66.7%) 1 (8.3%) 1 (8.3%) 3 (25%) 3 (25%) 1 (8.3%)
Do you support the following definition of familial ILD : occurrence of 2 or more first or second degree related family members with fibrotic ILD	N=13 13 (100%) 0 (0%)
How should we call it?*	N=14 5 (35.7%) 4 (28.6%) 4 (28.6%) 2 (14.2%) 1 (7.1%)

Do you include any polymorphisms in your diagnostic workup?	N=13
Yes	1 (7.7%)
No	12 (92.3%)

Table S3 Result of the survey for the screening of asymptomatic relatives

Should we offer a screening for asymptomatic relatives?	N=14 Yes 10 (71.4%) No 4 (28.6%)
Who should be evaluated?*	N=14 All relatives of FPF 9 (64.3%) Only if a mutation is evidenced in the proband 3 (21.4%) Only relatives carriers of the know mutation 2 (14.3%)
Should we do a Pulmonary CT scan?	N=14 Yes 14 (100%) No 0
When should we do a Pulmonary CT scan?*	N=14 In case of symptoms 10 (71.4%) In case of auscultation abnormalities 10 (71.4%) IN every relatives >18 3 (21.4%) >40 7 (50%) >50 10 (71.4%) >60 0 (0%) 10 years before the proband ILD onset 6 (42.9%)
Should we offer other pulmonary evaluation?*	N=13 Pulmonary function test 12 (92.3%) Pulmonary echocardiography 1 (7.7%)
Should we offer other test in TRG relatives?*	N=14 Complete blood count 13 (92.9%) Hepatic enzyme 12 (85.7%) Hepatic echocardiography 4 (28.6%) Telomere length analysis 4 (28.6%)
How often should we offer this screening?	N=13 Only once 1 (7.7%) Every Year 3 (23.1%) Every 2 years 2 (15.4%) Every 3 years 1 (7.7%) Every 4 years 1 (7.7%) Every 5 years 5 (38.5%)

For marked questions (*) respondents could choose more than one choice.

Bibliography

1. Cawthon RM. Telomere measurement by quantitative PCR. *Nucleic Acids Res.* 2002; 30: e47.
2. O'Callaghan NJ, Fenech M. A quantitative PCR method for measuring absolute telomere length. *Biol. Proced. Online* 2011; 13: 3.
3. Wang Y, Savage SA, Alsaggaf R, Aubert G, Dagnall CL, Spellman SR, Lee SJ, Hicks B, Jones K, Katki HA, Gadalla SM. Telomere Length Calibration from qPCR Measurement: Limitations of Current Method. *Cells* 2018; 7: E183.
4. Baerlocher GM, Vulto I, de Jong G, Lansdorp PM. Flow cytometry and FISH to measure the average length of telomeres (flow FISH). *Nat. Protoc.* 2006; 1: 2365–2376.
5. Aubert G, Hills M, Lansdorp PM. Telomere length measurement-caveats and a critical assessment of the available technologies and tools. *Mutat. Res.* 2012; 730: 59–67.
6. Derradji H, Bekaert S, Van Oostveldt P, Baatout S. Comparison of different protocols for telomere length estimation by combination of quantitative fluorescence *in situ* hybridization (Q-FISH) and flow cytometry in human cancer cell lines. *Anticancer Res.* 2005; 25: 1039–1050.
7. Gutierrez-Rodrigues F, Santana-Lemos BA, Scheucher PS, Alves-Paiva RM, Calado RT. Direct comparison of flow-FISH and qPCR as diagnostic tests for telomere length measurement in humans. *PLoS One* 2014; 9: e113747.
8. Ferreira MSV, Kirschner M, Halfmeyer I, Estrada N, Xicoy B, Isfort S, Vieri M, Zamora L, Abels A, Bouillon A-S, Begemann M, Schemionek M, Maurer A, Koschmieder S, Wilop S, Panse J, Brümmendorf TH, Beier F. Comparison of flow-FISH and MM-qPCR telomere length assessment techniques for the screening of telomeropathies. *Ann. N. Y. Acad. Sci.* 2020; 1466: 93–103.
9. Behrens YL, Thomay K, Hagedorn M, Ebersold J, Henrich L, Nustedt R, Schlegelberger B, Göhring G. Comparison of different methods for telomere length measurement in whole blood and blood cell subsets: Recommendations for telomere length measurement in hematological diseases. *Genes. Chromosomes Cancer* 2017; 56: 700–708.
10. El-Chemaly S, Young LR. Hermansky-Pudlak Syndrome. *Clin. Chest Med.* 2016; 37: 505–511.
11. Hengst M, Naehrlich L, Mahavadi P, Grosse-Onnebrink J, Terheggen-Lagro S, Skanke LH, Schuch LA, Brasch F, Guenther A, Reu S, Ley-Zaporozhan J, Gries M. Hermansky-Pudlak syndrome type 2 manifests with fibrosing lung disease early in childhood. *Orphanet J Rare Dis* 2018/03/28 ed. Ludwig-Maximilians University, Dr von Haunersches Kinderspital, German Center for Lung Research (DZL), Lindwurmstr. 4, 80337, Munich, Germany. University Hospital Gießen and Marburg, German Center for Lung Research, Feulgenstr. 12, 35385, Gießen, Germany.; 2018; 13: 42.
12. Picard C, Thouvenin G, Kannengiesser C, Dubus JC, Jeremiah N, Rieux-Lauca F, Crestani B, Belot A, Thivolet-Bejui F, Secq V, Menard C, Reynaud-Gaubert M, Reix P. Severe Pulmonary Fibrosis as the First Manifestation of Interferonopathy (TMEM173 Mutation). *Chest* 2016; 150: e65-71.
13. Liu Y, Jesus AA, Marrero B, Yang D, Ramsey SE, Sanchez GAM, Tenbrock K, Wittkowski H, Jones OY, Kuehn HS, Lee C-CR, DiMattia MA, Cowen EW, Gonzalez B, Palmer I, DiGiovanna JJ, Biancotto A, Kim H, Tsai WL, Trier AM, Huang Y, Stone DL, Hill S, Kim HJ, St Hilaire C, Gurprasad

- S, Plass N, Chapelle D, Horkayne-Szakaly I, Foell D, et al. Activated STING in a vascular and pulmonary syndrome. *N. Engl. J. Med.* 2014; 371: 507–518.
14. Watkin LB, Jessen B, Wiszniewski W, Vece TJ, Jan M, Sha Y, Thamsen M, Santos-Cortez RL, Lee K, Gambin T, Forbes LR, Law CS, Stray-Pedersen A, Cheng MH, Mace EM, Anderson MS, Liu D, Tang LF, Nicholas SK, Nahmod K, Makedonas G, Canter DL, Kwok PY, Hicks J, Jones KD, Penney S, Jhangiani SN, Rosenblum MD, Dell SD, Waterfield MR, et al. COPA mutations impair ER-Golgi transport and cause hereditary autoimmune-mediated lung disease and arthritis. *Nat Genet* 2015; 47: 654–660.
 15. Cho K, Yamada M, Agematsu K, Kanegane H, Miyake N, Ueki M, Akimoto T, Kobayashi N, Ikemoto S, Tanino M, Fujita A, Hayasaka I, Miyamoto S, Tanaka-Kubota M, Nakata K, Shiina M, Ogata K, Minakami H, Matsumoto N, Ariga T. Heterozygous Mutations in OAS1 Cause Infantile-Onset Pulmonary Alveolar Proteinosis with Hypogammaglobulinemia. *Am. J. Hum. Genet.* 2018; 102: 480–486.
 16. Magg T, Okano T, Koenig LM, Boehmer DFR, Schwartz SL, Inoue K, Heimall J, Licciardi F, Ley-Zaporozhan J, Ferdman RM, Caballero-Oteyza A, Park EN, Calderon BM, Dey D, Kanegane H, Cho K, Montin D, Reiter K, Griese M, Albert MH, Rohlfs M, Gray P, Walz C, Conn GL, Sullivan KE, Klein C, Morio T, Hauck F. Heterozygous OAS1 gain-of-function variants cause an autoinflammatory immunodeficiency. *Sci. Immunol.* 2021; 6: eabf9564.
 17. Seidl E, Schramm D, Schön C, Reiter K, Pawlita I, Kappler M, Reu-Hofer S, Hauck F, Albert M, Griese M. Pulmonary alveolar proteinosis due to heterozygous mutation in OAS1: Whole lung lavages for long-term bridging to hematopoietic stem cell transplantation. *Pediatr. Pulmonol.* 2022; 57: 273–277.
 18. Vavassori S, Chou J, Faletti LE, Haunerdinger V, Opitz L, Joset P, Fraser CJ, Prader S, Gao X, Schuch LA, Wagner M, Hoefele J, Maccari ME, Zhu Y, Elakis G, Gabbett MT, Forstner M, Omran H, Kaiser T, Kessler C, Olbrich H, Frosk P, Almutairi A, Platt CD, Elkins M, Weeks S, Rubin T, Planas R, Marchetti T, Koovely D, et al. Multisystem inflammation and susceptibility to viral infections in human ZNFX1 deficiency. *J. Allergy Clin. Immunol.* 2021; 148: 381–393.
 19. Hadchouel A, Drummond D, Pontoizeau C, Aoust L, Hurtado Nedelec M-M, El Benna J, Gachelin E, Perisson C, Vigier C, Schiff M, Lacaille F, Molina TJ, Berteloot L, Renolleau S, Ottolenghi C, Tréluyer J-M, de Blic J, Delacourt C. Methionine supplementation for multi-organ dysfunction in MetRS-related pulmonary alveolar proteinosis. *Eur. Respir. J.* 2021; : 2101554.
 20. Lenz D, Stahl M, Seidl E, Schöndorf D, Brennenstuhl H, Gesenhues F, Heinemann T, Longerich T, Mendes MI, Prokisch H, Salomons GS, Schön C, Smith DEC, Sommerburg O, Wagner M, Westhoff JH, Reiter K, Staufen C, Griese M. Rescue of respiratory failure in pulmonary alveolar proteinosis due to pathogenic MARS1 variants. *Pediatr. Pulmonol.* 2020; 55: 3057–3066.
 21. Schuch LA, Forstner M, Rapp CK, Li Y, Smith DEC, Mendes MI, Delhommel F, Sattler M, Emiralioglu N, Taskiran EZ, Orhan D, Kiper N, Rohlfs M, Jeske T, Hastreiter M, Gerstlauer M, Torrent-Vernetta A, Moreno-Galdó A, Kammer B, Brasch F, Reu-Hofer S, Griese M. FARS1-related disorders caused by bi-allelic mutations in cytosolic phenylalanyl-tRNA synthetase genes: Look beyond the lungs! *Clin. Genet.* John Wiley & Sons, Ltd; 2021; 99: 789–801.
 22. Hildebrandt J, Yalcin E, Bresser H-G, Cinel G, Gappa M, Haghghi A, Kiper N, Khalilzadeh S, Reiter K, Sayer J, Schwerk N, Sibbersen A, Van Daele S, Nübling G, Lohse P, Griese M. Characterization of CSF2RA mutation related juvenile pulmonary alveolar proteinosis. *Orphanet J. Rare Dis.* 2014; 9: 171.

23. Suzuki T, Maranda B, Sakagami T, Catellier P, Couture CY, Carey BC, Chalk C, Trapnell BC. Hereditary pulmonary alveolar proteinosis caused by recessive CSF2RB mutations. *Eur Respir J* 2011; 37: 201–204.
24. Rapp CK, Van Dijck I, Laugwitz L, Boon M, Briassoulis G, Ilia S, Kammer B, Reu S, Hornung S, Buchert R, Sofan L, Froukh T, Witters P, Rymen D, Haack TB, Proesmans M, Gries M. Expanding the phenotypic spectrum of FINCA (fibrosis, neurodegeneration, and cerebral angiogenesis) syndrome beyond infancy. *Clin. Genet.* 2021; 100: 453–461.
25. Levran O, Desnick RJ, Schuchman EH. Niemann-Pick disease: a frequent missense mutation in the acid sphingomyelinase gene of Ashkenazi Jewish type A and B patients. *Proc. Natl. Acad. Sci. U. S. A.* 1991; 88: 3748–3752.
26. Schuchman EH. The pathogenesis and treatment of acid sphingomyelinase-deficient Niemann-Pick disease. *J. Inherit. Metab. Dis.* 2007; 30: 654–663.
27. Gries M, Brasch F, Aldana VR, Cabrera MM, Goelnitz U, Ikonen E, Karam BJ, Liebisch G, Linder MD, Lohse P, Meyer W, Schmitz G, Pamir A, Ripper J, Rolfs A, Schams A, Lezana FJ. Respiratory disease in Niemann-Pick type C2 is caused by pulmonary alveolar proteinosis. *Clin. Genet.* 2010; 77: 119–130.
28. Ballerie A, Nimubona S, Meunier C, Gutierrez FL, Desrues B, Delaval P, Jouneau S. Association of pulmonary alveolar proteinosis and fibrosis: patient with GATA2 deficiency. *Eur Respir J* 2016; 48: 1510–1514.
29. Jouneau S, Ballerie A, Kerjouan M, Demant X, Blanchard E, Lederlin M. Haemodynamically proven pulmonary hypertension in a patient with GATA2 deficiency-associated pulmonary alveolar proteinosis and fibrosis. *Eur. Respir. J.* 2017; 49.
30. Corut A, Senyigit A, Ugur SA, Altin S, Ozcelik U, Calisir H, Yildirim Z, Gocmen A, Tolun A. Mutations in SLC34A2 cause pulmonary alveolar microlithiasis and are possibly associated with testicular microlithiasis. *Am. J. Hum. Genet.* 2006; 79: 650–656.
31. Mercier S, Kury S, Shaboodien G, Houniet DT, Khumalo NP, Bou-Hanna C, Bodak N, Cormier-Daire V, David A, Faivre L, Figarella-Branger D, Gherardi RK, Glen E, Hamel A, Laboisse C, Le Caignec C, Lindenbaum P, Magot A, Munnich A, Mussini JM, Pillay K, Rahman T, Redon R, Salort-Campana E, Santibanez-Koref M, Thauvin C, Barbarot S, Keavney B, Bezieau S, Mayosi BM. Mutations in FAM111B cause hereditary fibrosing poikiloderma with tendon contracture, myopathy, and pulmonary fibrosis. *Am J Hum Genet* 2013; 93: 1100–1107.
32. Mercier S, Küry S, Salort-Campana E, Magot A, Agbim U, Besnard T, Bodak N, Bou-Hanna C, Bréhéret F, Brunelle P, Caillou F, Chabrol B, Cormier-Daire V, David A, Eymard B, Faivre L, Figarella-Branger D, Fleurence E, Ganapathi M, Gherardi R, Goldenberg A, Hamel A, Igual J, Irvine AD, Israël-Biet D, Kannengiesser C, Laboisse C, Caignec CL, Mahé J-Y, Mallet S, et al. Expanding the clinical spectrum of hereditary fibrosing poikiloderma with tendon contractures, myopathy and pulmonary fibrosis due to FAM111B mutations. *Orphanet J. Rare Dis.* 2015; 10: 135.
33. Nir V, Ilivitky A, Hakim F, Yoseph RB, Gur M, Mandel H, Bentur L. Pulmonary manifestations of prolidase deficiency. *Pediatr. Pulmonol.* 2016; 51: 1229–1233.

34. Cottin V, Nasser M, Traclet J, Chalabreysse L, Lèbre AS, Si-Mohamed S, Philit F, Thivolet-Béjui F. Prolidase deficiency: a new genetic cause of combined pulmonary fibrosis and emphysema syndrome in the adult. *Eur Respir J* 2020; 55.
35. McManus DT, Moore R, Hill CM, Rodgers C, Carson DJ, Love AH. Necropsy findings in lysinuric protein intolerance. *J. Clin. Pathol.* 1996; 49: 345–347.
36. Mauhin W, Habarou F, Gobin S, Servais A, Brassier A, Grisel C, Roda C, Pinto G, Moshous D, Ghalim F, Krug P, Deltour N, Pontoizeau C, Dubois S, Assoun M, Galmiche L, Bonnefont J-P, Ottolenghi C, de Blic J, Arnoux J-B, de Lonlay P. Update on Lysinuric Protein Intolerance, a Multi-faceted Disease Retrospective cohort analysis from birth to adulthood. *Orphanet J. Rare Dis.* [Internet] 2017 [cited 2019 Jul 2]; 12Available from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5217205/>.
37. Hartmannova H, Piherova L, Tauchmannova K, Kidd K, Acott PD, Crocker JF, Oussedik Y, Mallet M, Hodanova K, Stranecky V, Pristoupilova A, Baresova V, Jedlickova I, Zivna M, Sovova J, Hulkova H, Robins V, Vrbacky M, Pecina P, Kaplanova V, Houstek J, Mracek T, Thibeault Y, Bleyer AJ, Kmoch S. Acadian variant of Fanconi syndrome is caused by mitochondrial respiratory chain complex I deficiency due to a non-coding mutation in complex I assembly factor NDUFAF6. *Hum Mol Genet* 2016; 25: 4062–4079.
38. Goto M. Werner's syndrome: from clinics to genetics. *Clin. Exp. Rheumatol.* 2000; 18: 760–766.
39. Goletto T, Crockett F, Aractingi S, Toper C, Senet P, Cadranel J, Naccache J-M. Interstitial Lung Disease in Werner Syndrome: A Case Report of a 55-Year-Old Male Patient. *Case Rep. Pulmonol.* 2015; 2015: 361694.
40. Al-Mutairy EA, Imtiaz FA, Khalid M, Al Qattan S, Saleh S, Mahmoud LM, Al-Saif MM, Al-Haj L, Al-Enazi A, AlJebreen AM, Mohammed SF, Mobeireek AF, Alkattan K, Chisti MA, Luzina IG, Al-Owain M, Weheba I, Abdelsayed AM, Ramzan K, Janssen LJ, Conca W, Alaiya A, Collison KS, Meyer BF, Atamas SP, Khabar KS, Hasday JD, Al-Mohanna F. An atypical pulmonary fibrosis is associated with co-inheritance of mutations in the calcium binding protein genes S100A3 and S100A13. *Eur Respir J* 2019; 54.