



Biomarkers to identify *Mycobacterium tuberculosis* infection among borderline QuantiFERON results

Jonathan W. Uzorka¹, Jaap A. Bakker², Krista E. van Meijgaarden¹, Eliane M.S. Leyten³, Nathalie M. Delfos⁴, David J. Hetem⁵, Jos Kerremans⁶, Mieke Zwarts⁷, Sandra Cozijn⁸, Tom H.M. Ottenhoff¹, Simone A. Joosten¹ and Sandra M. Arend¹

¹Dept of Infectious Diseases, Leiden University Medical Centre, Leiden, The Netherlands. ²Dept of Clinical Chemistry and Laboratory Medicine, Leiden University Medical Centre, Leiden, The Netherlands. ³Dept of Internal Medicine, Haaglanden Medical Centre, Den Haag, The Netherlands. ⁴Dept of Internal Medicine, Alrijne Hospital, Leiderdorp, The Netherlands. ⁵Dept of Medical Microbiology, Haaglanden Medical Centre, Den Haag, The Netherlands. ⁶Dept of Medical Microbiology, Alrijne Hospital, Leiderdorp, The Netherlands. ⁷Dept of Clinical Chemistry and Laboratory Medicine, Haaglanden Medical Centre, Den Haag, The Netherlands. ⁸Dept of Medical Microbiology, Alrijne Hospital, Leiden, The Netherlands.

Corresponding author: Jonathan W. Uzorka (j.w.uzorka@lumc.nl)



Shareable abstract (@ERSpublications)

Additional laboratory testing of IP-10/CXCL10 and MIG/CXCL9 in individuals at increased risk of reactivation TB with a borderline QuantiFERON-TB Gold Plus result just below the formal cut-off helps to identify those with *M. tuberculosis* infection <https://bit.ly/3t8zR7r>

Cite this article as: Uzorka JW, Bakker JA, van Meijgaarden KE, et al. Biomarkers to identify *Mycobacterium tuberculosis* infection among borderline QuantiFERON results. *Eur Respir J* 2022; 60: 2102665 [DOI: 10.1183/13993003.02665-2021].

Copyright ©The authors 2022.

This version is distributed under the terms of the Creative Commons Attribution Non-Commercial Licence 4.0. For commercial reproduction rights and permissions contact permissions@ersnet.org

This article has an editorial commentary: <https://doi.org/10.1183/13993003.00697-2022>

Received: 8 Oct 2021
Accepted: 18 Dec 2021

Abstract

Background Screening for tuberculosis (TB) infection often includes QuantiFERON-TB Gold Plus (QFT) testing. Previous studies showed that two-thirds of patients with negative QFT results just below the cut-off, so-called borderline test results, nevertheless had other evidence of TB infection. This study aimed to identify a biomarker profile by which borderline QFT results due to TB infection can be distinguished from random test variation.

Methods QFT supernatants of patients with a borderline (≥ 0.15 and < 0.35 IU·mL⁻¹), low-negative (< 0.15 IU·mL⁻¹) or positive (≥ 0.35 IU·mL⁻¹) QFT result were collected in three hospitals. Bead-based multiplex assays were used to analyse 48 different cytokines, chemokines and growth factors. A prediction model was derived using LASSO regression and applied further to discriminate QFT-positive *Mycobacterium tuberculosis*-infected patients from borderline QFT patients and QFT-negative patients

Results QFT samples of 195 patients were collected and analysed. Global testing revealed that the levels of 10 kDa interferon (IFN)- γ -induced protein (IP-10/CXCL10), monokine induced by IFN- γ (MIG/CXCL9) and interleukin-1 receptor antagonist in the antigen-stimulated tubes were each significantly higher in patients with a positive QFT result compared with low-negative QFT individuals ($p < 0.001$). A prediction model based on IP-10 and MIG proved highly accurate in discriminating patients with a positive QFT (TB infection) from uninfected individuals with a low-negative QFT (sensitivity 1.00 (95% CI 0.79–1.00) and specificity 0.95 (95% CI 0.74–1.00)). This same model predicted TB infection in 68% of 87 patients with a borderline QFT result.

Conclusions This study was able to classify borderline QFT results as likely infection-related or random. These findings support additional laboratory testing for either IP-10 or MIG following a borderline QFT result for individuals at increased risk of reactivation TB.

Introduction

Screening for tuberculosis (TB) infection has been recommended in individuals at increased risk of developing reactivation TB [1, 2]. In a hospital setting, this predominantly includes patients eligible for monoclonal antibody therapy to treat immune-mediated inflammatory diseases or anti-rejection therapy following transplantation. Immunological tests are frequently employed to diagnose or exclude TB infection, sometimes combined with a chest radiograph [3]. There are two types of immunological tests



available: the century-old tuberculin skin test (TST) and the more recently introduced interferon- γ release assay (IGRA) [4, 5]. At present there are two commercially available IGRAs: QuantiFERON-TB Gold Plus (QFT; whole blood stimulation followed by ELISA) and T-SPOT.TB (ELISPOT assay on isolated cells). A recent meta-analysis of head-to-head diagnostic accuracy studies estimated that the sensitivity and specificity of T-SPOT.TB and QFT for active TB were 90.2% versus 91.4% (sensitivity) and 98.1% versus 97.8% (specificity), respectively [6], which was higher than previously reported and, in part, due to the respective study populations [7]. Of note, the accuracy of the QFT appeared to be similar (in head-to-head studies) or even slightly higher (QFT-Plus only) compared with its predecessor (QuantiFERON-TB Gold In-Tube) [6, 8].

The interpretation of both IGRA test results differs. Valid T-SPOT.TB test results are classified as negative, borderline or positive, while valid QFT test results are only interpreted dichotomously: either negative or positive [9]. Serial testing of healthcare workers showed that QFT results just above the cut-off value (≥ 0.35 IU·mL $^{-1}$) often reverted (became negative) and QFT results just below the cut-off value (< 0.35 IU·mL $^{-1}$) often converted (became positive) [10–12]. For this reason, the implementation of a certain “borderline zone” has been suggested, with the lower limit varying from 0.15 to 0.20 IU·mL $^{-1}$ and the upper limit varying from 0.70 to 0.80 IU·mL $^{-1}$ [12–15]. The proportion of individuals with such a result appears to be affected by the time since exposure to *Mycobacterium tuberculosis* and relevant comorbidities, such as immune-mediated inflammatory diseases, but interestingly not by HIV status [16, 17]. Our previous studies demonstrated that nearly two-thirds of patients with a result in the borderline zone below the cut-off had either a positive TST and/or a positive T-SPOT.TB result, suggesting a true infection with *M. tuberculosis* [18, 19]. Although a gold standard for diagnosing TB infection does not exist, we aimed to discriminate between infected and uninfected patients with borderline QFT results below the cut-off. This is particularly beneficial for patients who are soon to start biological or pharmacological immunosuppressive therapy, as withholding preventive therapy due to a false-negative QFT result may result in reactivation TB, while initiating preventive treatment in patients with true-negative QFT results may expose patients to potential harmful side-effects of preventive therapy and a delay of the immunosuppressive treatment.

Therefore, the aim of this study was to differentiate between borderline results due to specific T-cell responses to *M. tuberculosis* or random assay variability. A broad panel of immune markers were measured in QFT supernatant samples of individuals with low-negative, borderline and positive QFT results in order to find a biomarker (profile) that correlates well with the positive test results but is absent in individuals with low-negative (0.15 IU·mL $^{-1}$) test results.

Methods

The protocol of this study (G19.009) was approved by the local institutional review board (METC-LDD) and the requirement for informed consent was waived. The study aimed to include twice as many individuals with a borderline QFT result as those with a negative or positive QFT result. Considering the slightly varying lower margins of the borderline category below the cut-off (0.20 IU·mL $^{-1}$ being the most prevalent), the borderline limits for this study were set at ≥ 0.15 and < 0.35 IU·mL $^{-1}$ based on findings from two previous studies [19, 20].

Sample collection and data retrieval

Residual QFT samples (from the negative control and positive control and two antigen-specific tubes) obtained as part of routine care were collected at three distinct laboratory sites at three different hospitals in the Netherlands: Leiden University Medical Centre (LUMC; Leiden), Haaglanden Medical Centre Westeinde (HMC-W; The Hague) and Alrijne Hospital Leiderdorp (AL; Leiderdorp). At these three hospitals, QFTs were performed during screening for TB infection and sometimes diagnostic work-up for active TB of clinical patients as well as outpatients (often prior to biological treatment). In the defined study period, ~150 μ L blood plasma was isolated from all four QFT tubes in all individuals with a borderline (≥ 0.15 and < 0.35 IU·mL $^{-1}$) or positive (≥ 0.35 IU·mL $^{-1}$) QFT result, and in every third individual with a low-negative (< 0.15 IU·mL $^{-1}$) QFT result. Subsequently, all samples were stored at -80°C . At LUMC, samples were collected from July 2019 through November 2020. At AL and HMC-W, samples were collected between May 2020 and November 2020. Initially, all collected samples were included. After assessment of clinical data, which was retrieved between December 2020 and January 2021, we excluded all individuals < 18 years of age at the time of testing, as well as those who were diagnosed with active TB. Collected data included sociodemographics, region of origin, reason for testing, bacille Calmette–Guérin (BCG) vaccination status, TST results if available, immune status at the time of testing and relevant comorbidities, such as immune-mediated inflammatory diseases, malignancies and

chronic renal failure. Medical records were checked for evidence of *M. tuberculosis* infection (including active TB disease) in November 2021, 12–24 months after collection of the original sample.

Bead-based multiplex assay

All samples were analysed using the Bio-Plex Pro Human Cytokine Screening Panel 48-plex (Bio-Rad, Veenendaal, The Netherlands), which was selected for its broad coverage of relevant inflammatory pathways and included the following 48 cytokines, chemokines and growth factors: β -nerve growth factor (NGF), cutaneous T-cell attracting chemokine (CTACK), fibroblast growth factor (FGF)-basic, eotaxin (CCL11), granulocyte colony-stimulating factor (G-CSF), granulocyte–macrophage CSF (GM-CSF), GRO- α (CXCL1), hepatocyte growth factor (HGF), interferon (IFN)- γ , interleukin (IL)-1 β , IL-1 receptor antagonist (ra), IL-1 α , IL-2, IL-2ra, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IFN- α 2, IL-10, IL-12 (p40), IL-12 (p70), IL-13, IL-15, IL-16, IL-17A, IL-18, 10 kDa IFN- γ -induced protein (IP-10/CXCL10), leukaemia inhibitory factor (LIF), monocyte chemoattractant protein (MCP)-1, MCP-3, macrophage CSF (M-CSF), macrophage migration inhibitory factor (MIF), monokine induced by IFN- γ (MIG/CXCL9), macrophage inflammatory protein (MIP)-1 α , MIP-1 β , platelet-derived growth factor (PDGF)-BB, regulated on activation normal T-cell expressed and secreted (RANTES), stem cell factor (SCF), stem cell growth factor (SCGF)- β , stromal cell-derived factor (SDF)-1 α , tumour necrosis factor (TNF)- α , TNF- β , TNF-related apoptosis-inducing ligand (TRAIL), and vascular endothelial growth factor (VEGF). The multiplex magnetic bead assay was performed in accordance with the manufacturer's instructions. Subsequent processing was performed as described by VAN MEIJGAARDEN *et al.* [21]. The measured concentration of each biomarker in the unstimulated tube (NIL) was subtracted from that in the stimulated tubes (TB1 and TB2). Only the highest concentration of both stimulated tubes was used for analysis.

Statistical analyses

Statistical analyses were performed using SPSS Statistics version 25 (IBM, Armonk, NY, USA), R version 4.0.3 (R Foundation for Statistical Computing, Vienna, Austria) and Prism 8 (GraphPad, La Jolla, CA, USA). *p*-values <0.05 were considered statistically significant. Categorical data were analysed and displayed as numerator/denominator to account for missing data.

Linear discriminant analysis (LDA) was conducted in order to separate and classify all samples in low-negative, borderline and positive QFT categories, based on all biomarkers (except for IFN- γ). LDA is a technique which focuses predominantly on the maximalisation of the separability among known categories by maximising the distances (d^2) between the means to a central point while minimising the variation (s^2) within each category. The analyses were performed using R package “MASS”. For visualisation, R packages “caret” and “tidyverse” were used. Global testing was performed in order to test for pairwise differences in levels between patients with a low-negative, borderline and positive QFT result. All biomarkers were clustered using hierarchical clustering, based on absolute correlation distance and average linkage. Correction for multiple testing was applied using the “inheritance” procedure, which is an adaptation of the Meinshausen method [22]. All procedures were performed using R package “globaltest” version 5.44. LASSO (Least Absolute Shrinkage and Selection Operator) regression analysis was *a priori* chosen to construct a simple model enabling discrimination between individuals with a low-negative and positive QFT result based on relevant biomarkers derived from the global test analysis. In order to construct a biomarker signature, data were randomly split into a training set, containing two-thirds of all low-negatives and positives, and a test set, which consisted of the remaining one-third. If the model's predictive accuracy is sufficient it will be used to differentiate patients likely infected with *M. tuberculosis* from those without infection among those with a QFT borderline result. LASSO regression and subsequent receiver operating characteristic (ROC) curves were computed using R packages “glmnet” and “pROC”. A biomarker heatmap was visualised using R package “pheatmap”; correlations were performed using R packages “ggpubr”, “corrplot” and “ggcorplot”. Cut-offs were determined by calculating the Youden index.

Results

From 210 individuals, supernatants from two stimulated tubes, as well as corresponding positive (mitogen) and negative (NIL) control tubes were collected. Of those, samples from 15 individuals were excluded due to being underage at the time of testing (*n*=9), active concurrent TB (*n*=5) or if patient information could not be retrieved from the electronic patient files (*n*=1). The collected samples from the remaining 195 patients were included in this analysis. Of those, 57 had a low-negative, 87 a borderline and 51 a positive QFT result.

The majority of samples were from LUMC (61.5%). Sociodemographics did not differ between the three hospitals (table 1). In almost two-thirds of our population a QFT was performed during screenings for TB infection, most of which were performed prior to immunosuppressive therapy or transplantation, while in

TABLE 1 Characteristics of all included patients

	Site			All (n=195)	p-value
	LUMC (n=120)	HMC-W (n=60)	AL (n=15)		
Age (years)	49.5±18.7	51.6±18.1	53.2±20.2	50.4±18.6	0.649
Male	61/120 (50.8)	28/60 (46.7)	9/15 (60.0)	98/195 (50.3)	0.639
Originating from TB endemic region [#]	35/117 (29.9)	23/60 (38.3)	3/15 (20.0)	61/192 (31.8)	0.343
Reason for testing					0.018
Suspect for active TB	36/120 (30.0)	29/60 (48.3)	8/15 (53.3)	73/195 (37.4)	
Screening for TB infection	84/120 (70.0)	31/60 (51.7)	7/15 (46.7)	122/195 (62.6)	
Immunosuppression/pre-transplantation	40/84 (47.6)	22/31 (71.0)	5/7 (71.4)	67/122 (54.9)	
Screening donor	7/84 (8.3)	0/31 (0)	0/7 (0)	7/122 (5.7)	
Contact investigation	19/84 (22.6)	3/31 (9.7)	0/7 (0)	22/122 (18.0)	
Screening healthcare worker	9/84 (10.7)	1/31 (3.2)	0/7 (0)	10/122 (8.2)	
Other	9/84 (10.7)	5/31 (16.1)	2/7 (28.6)	16/122 (13.1)	
Immunosuppressive therapy[¶]	22/89 (24.7)	11/56 (19.6)	0/14 (0)	33/126 (20.8)	0.129
Biologicals	1/22 (4.5)	0/11 (0)	0/14 (0)	1/33 (3.0)	1.000
Calcineurin inhibitors	2/22 (9.1)	0/11 (0)	0/14 (0)	2/33 (6.1)	0.542
Anti-metabolites	5/22 (22.7)	1/11 (9.1)	0/14 (0)	6/33 (18.2)	0.637
Glucocorticoids	13/22 (65.0)	7/11 (63.6)	0/14 (0)	20/33 (60.6)	1.000
DMARDs	5/22 (22.7)	4/11 (36.4)	0/14 (0)	9/33 (27.3)	0.438
QFT					<0.001
<0.15 IU·mL ⁻¹	29/120 (24.2)	18/60 (30.0)	10/15 (66.7)	57/210 (29.2)	
≥0.15 and <0.35 IU·mL ⁻¹	65/120 (54.2)	21/60 (35.0)	1/15 (6.7)	87/210 (44.6)	
≥0.35 IU·mL ⁻¹	26/120 (21.7)	21/60 (35.0)	4/15 (26.7)	51/210 (26.2)	

Data are presented as mean±SD or n/N (%), unless otherwise stated. LUMC: Leiden University Medical Centre; HMC: Haaglanden Medical Centre Westeinde; AL: Alrijne Hospital Leiderdorp; TB: tuberculosis; DMARD: disease-modifying anti-rheumatic drug; QFT: QuantiFERON-TB Gold Plus. [#]: defined as a country with a TB incidence ≥40 cases of active TB per 100 000 inhabitants; [¶]: only includes individuals from whom type and dose of immunosuppressive therapy could be retrieved.

over one-third a QFT was performed due to suspicion of active TB. None of the included individuals had developed reactivation TB after a 1- to 2-year follow-up, despite immune-modulating therapy in many.

Discriminating groups using LDA

LDA was performed using data for all biomarkers (with omission of IFN- γ) measured in the *M. tuberculosis* antigen-stimulated tubes corrected for background (NIL tubes) to investigate whether biomarkers other than IFN- γ were able to discriminate samples of patients with low-negative, borderline and positive QFT results. The plot in figure 1 shows obvious grouping according to these 47 biomarkers. Patients with a positive QFT result were clearly separated from those with a low-negative QFT result. However, the plot also shows that patients with a borderline QFT result were often classified as either low-negative or positive, highlighting the diversity of this specific group. As a result, the prediction accuracy of the model was only 66.7%.

Identification of biomarkers for TB infection

A heatmap with an overview of all biomarker levels is displayed in supplementary figure S1. All TB1 biomarkers correlated strongly with their counterparts measured in the TB2 tubes, except for IL-15 in the low-negative category and IL-3 among those with a positive QFT result (supplementary figure S2a–d). Global test analysis was performed on all biomarkers (with omission of IFN- γ) measured in the TB-specific tubes (corrected for NIL) in order to construct a more precise profile associated with TB infection. After correction for multiple testing, this analysis revealed three biomarkers that were significantly higher in patients with TB infection (based on a positive QFT result) compared with individuals with a low-negative QFT result (IP-10, MIG and IL-1ra; $p<0.001$) (figure 2). Among those with either a positive or low-negative QFT result, ROC analysis revealed that both MIG (area under the curve (AUC) 0.98, 95% CI 0.96–1.00; cut-off 1176 pg·mL⁻¹) and IP-10 (AUC 0.95, 95% CI 0.91–0.99; cut-off 861 pg·mL⁻¹) (supplementary figures S3a and b) were highly accurate for TB infection (based on a positive QFT result), while the accuracy of IL-1ra was moderate (AUC 0.78, 95% CI 0.70–0.87; cut-off 3444 pg·mL⁻¹) (supplementary figure S3c). The relationship between these three biomarkers using Spearman's correlation coefficient revealed that IP-10 and MIG were strongly correlated ($R=0.88$, $p<0.001$) (figure 3a). As expected, the correlations between IL-1ra and both IP-10 and MIG were significant but moderate ($R=0.41$, $p<0.001$ and $R=0.43$, $p<0.001$, respectively) (figure 3b and c).

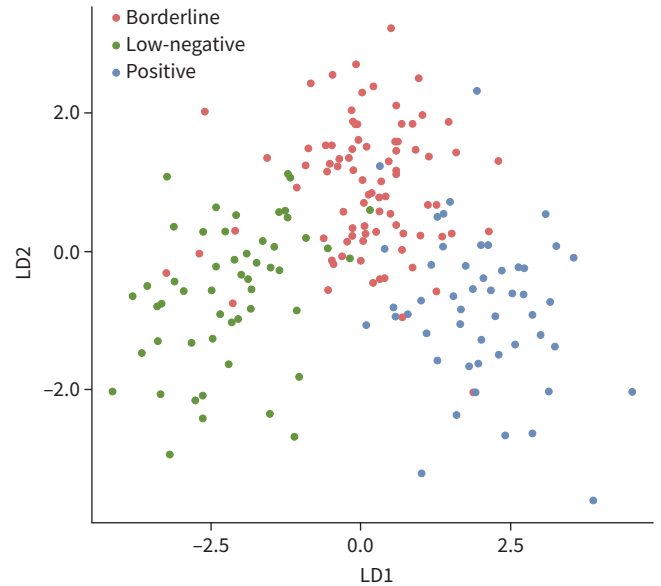


FIGURE 1 Classification of all samples using linear discriminant analysis (LDA). QuantiFERON-TB Gold Plus results distinguished by LDA, including all cytokines (except for interferon- γ), chemokines and growth factors. All data were \log_2 -transformed prior to analysis. Includes all data.

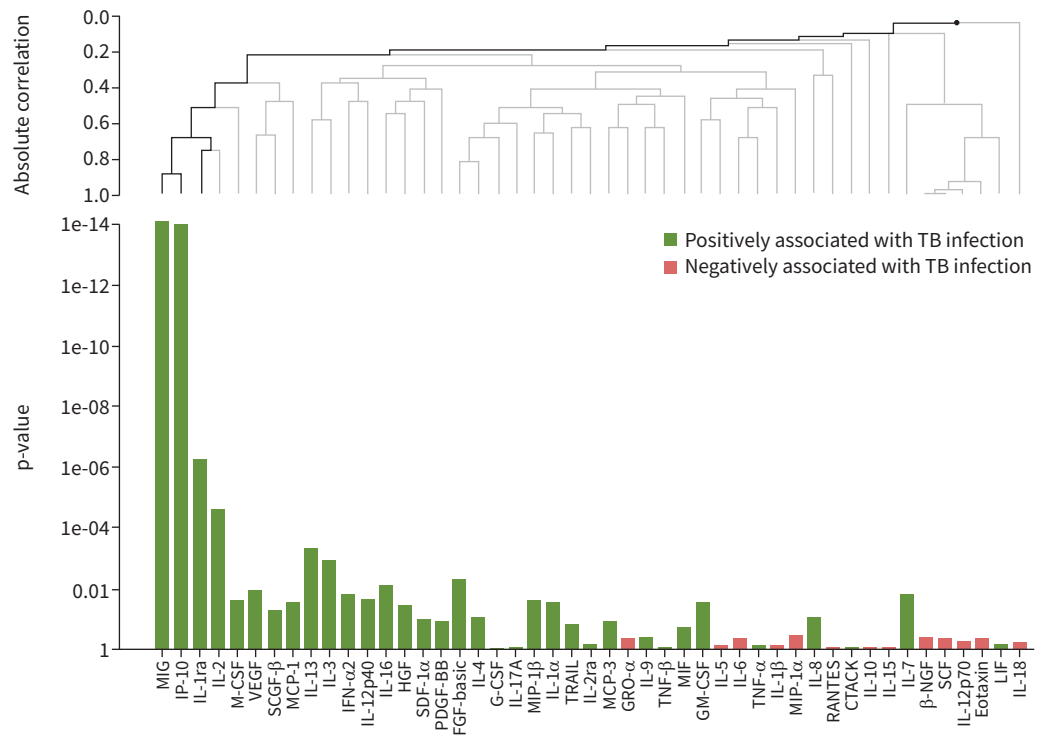


FIGURE 2 Comparison of biomarker levels between positive *versus* low-negative QuantiFERON-TB Gold Plus (QFT) samples. A pairwise comparison of all cytokines, chemokines and growth factors between positive and low-negative QFT samples was conducted using global test analysis. The absolute correlation between biomarkers is depicted using hierarchal clustering. Uncorrected p-values are depicted on the left of the bar chart. Black lines on the clustering graph indicate significant multiplicity corrected p-values, while nonsignificant p-values are depicted with grey lines. Includes all data. See the bead-based multiplex assay section of the Methods for details of biomarkers included in the Bio-Plex Pro Human Cytokine Screening Panel.

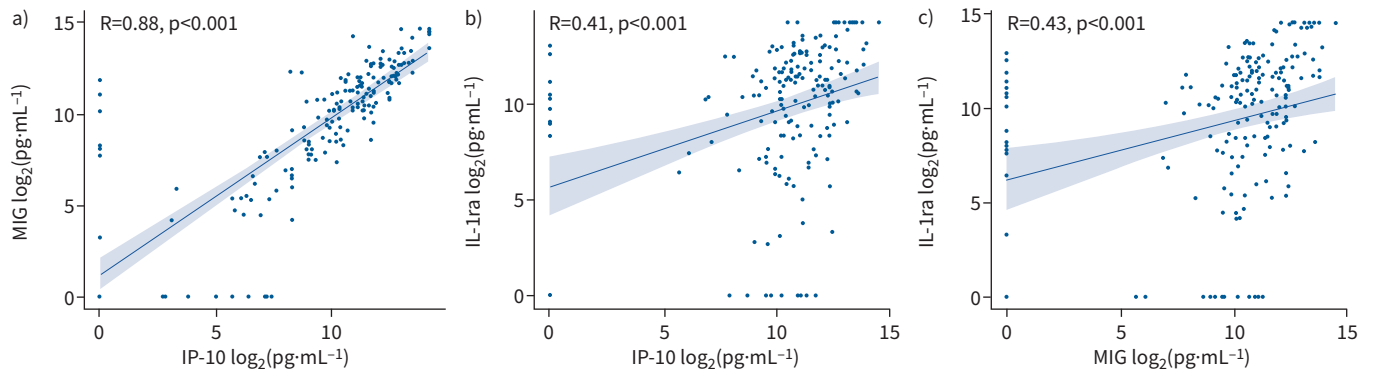


FIGURE 3 Correlation between 10 kDa interferon (IFN)- γ -induced protein (IP-10), monokine induced by IFN- γ (MIG) and interleukin-1 receptor antagonist (IL-1ra). A positive correlation between a) IP-10 and MIG, b) IP-10 and IL-1ra, and c) MIG and IL-1ra was measured by Spearman's correlation coefficient and two-tailed p-value analysis. All data were \log_2 -transformed prior to analysis. Includes all data.

Development of a prediction model for TB infection

The discriminatory biomarkers IP-10, MIG and IL-1ra were subsequently used in a LASSO regression model to obtain a biomarker signature that was able to distinguish patients with TB infection, based on a positive QFT result, from uninfected individuals, based on a low-negative QFT result (training data) (figure 4a). The classifying capacity of this biomarker profile was validated and displayed as a ROC curve (test data) (figure 4b), which yielded an AUC of 0.97 (95% CI 0.92–1.00). Corresponding sensitivity and specificity were high: 1.00 (95% CI 0.79–1.00) and 0.95 (95% CI 0.74–1.00), respectively. Adding biomarkers that were significant without correction did not improve the classifying capability of the biomarker signature. Of note, the model assigned “TB infection” to all patients who were excluded due to microbiologically proven active TB ($n=5$) (supplementary table S1).

Prediction model applied to borderline results

Subsequently, the same regression model, which successfully discriminated between patients with TB infection and uninfected individuals, was used to further characterise individuals with a borderline QFT result. Among patients with a borderline QFT result, 59 out of 87 (67.8%) were positive and thus likely to have TB infection (figure 4c). Of the three patients with a low-negative QFT result who were classified as TB-infected in the model, one had a QFT result of $0.14 \text{ IU}\cdot\text{mL}^{-1}$ in association with a TST induration of 10 mm, one received glucocorticoids during testing and one was not tested with the TST.

The characteristics of those 59 patients with a borderline QFT result of whom the model predicted to have TB infection (TB infection profile+) were compared with the 28 patients with a borderline QFT result in whom the model predicted to be uninfected (TB infection profile-) (table 2). Borderline TB infection profile+ individuals did not differ significantly from TB infection profile- individuals regarding clinical characteristics. Although mean TST values were higher among TB infection profile+ individuals, median TST and QFT values did not differ significantly from TB infection profile- individuals. Findings on chest radiography possibly suggestive of prior TB infection were scarce and did not significantly differ among groups (table 2). After Bonferroni adjustment for multiple testing, no significant differences were noted between biomarker values with respect to TB endemicity, immunosuppressive therapy and autoimmune diseases (supplementary tables S2–S4).

Classification of borderline results based on individual biomarker cut-offs

Median levels of IP-10 and MIG differed significantly between individuals with a low-negative, borderline or positive QFT result ($p<0.001$) (figure 5), while levels of IL-1ra were similar among those with a low-negative and borderline QFT result. Of note, specific focus on the low-positive QFT results ($n=12$; ≥ 0.35 and $<0.70 \text{ IU}\cdot\text{mL}^{-1}$) revealed a similar quantitative relation between QFT results and IP-10/MIG values (supplementary figure S4). An additional analysis of borderline QFT results, independent of the prediction model, was performed based on the individual cut-offs of IP-10 and MIG combined; cut-offs were determined based on the optimal sensitivity and specificity combination in ROC curves as described for supplementary figure S3 (figure 6). The analysis revealed that 54 out of 87 (62%) of all individuals with a borderline result had both IP-10 and MIG levels above the cut-offs designated for TB infection. In addition, all 54 individuals were predicted to have TB infection by our previous model.

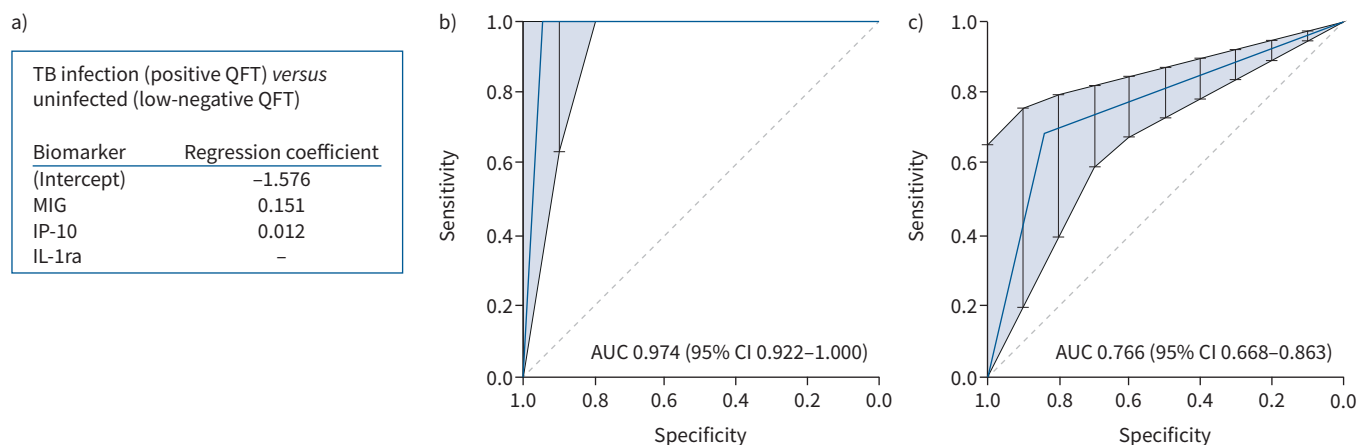


FIGURE 4 Accuracy of the biomarker signature. **a)** Biomarker signature derived from LASSO regression analysis based on patients with a positive QuantiFERON-TB Gold Plus (QFT) versus low-negative QFT result. Includes only training data. **b, c)** Receiver operating characteristic curve plots showing the prediction accuracy of the model allowing discrimination **b)** between patients with tuberculosis (TB) infection (based on a positive QFT result) and without TB infection (based on a low-negative QFT result) and **c)** between patients with a borderline QFT result and subjects without TB infection (based on a low-negative QFT result). Includes only test data. MIG: monokine induced by interferon (IFN)- γ ; IP-10: 10 kDa IFN- γ -induced protein; IL-1ra: interleukin-1 receptor antagonist; AUC: area under the curve.

TABLE 2 Characteristics by QuantiFERON-TB Gold Plus (QFT) result and biomarker profile

	Low-negative (n=57)	Borderline [#]		Positive (n=51)	All (n=195)	p-value
		Profile- (n=28)	Profile+ (n=59)			
Age (years)	47.4±16.6	48.5±19.8	48.9±18.8	56.7±18.8	50.4±18.6	0.042
Male	26/57 (45.6)	16/28 (57.1)	26/59 (44.1)	30/51 (58.8)	98/195 (50.3)	0.332
Originating from a TB endemic region [¶]	8/57 (14.0)	8/27 (29.6)	16/57 (28.1)	29/51 (56.9)	61/192 (31.8)	<0.001
Reason for testing						0.033
Suspect for active TB	20/57 (35.1)	6/28 (21.4)	20/59 (33.9)	27/51 (52.9)	73/195 (37.4)	
Screening for TB infection	37/57 (64.9)	22/28 (78.6)	39/59 (66.1)	24/51 (47.1)	122/195 (62.6)	
Comorbidity	38/57 (66.7)	20/28 (71.4)	47/59 (79.7)	32/51 (62.7)	137/195 (70.3)	0.236
Autoimmune disease	28/57 (49.1)	7/28 (25.0)	18/59 (30.5)	15/51 (29.4)	68/195 (34.9)	0.059
Chronic renal failure	1/57 (1.8)	3/28 (10.7)	5/59 (8.5)	3/51 (5.9)	12/195 (6.2)	0.261
Immunosuppressive therapy [*]	11/51 (21.6)	8/23 (34.8)	9/44 (20.5)	5/41 (12.2)	33/159 (20.8)	0.199
Biologicals	0/11 (0)	0/8 (0)	0/9 (0)	1/5 (20.0)	1/33 (3.0)	0.157
Calcineurin inhibitors	0/11 (0)	1/8 (12.5)	0/9 (0)	1/5 (20.0)	2/33 (6.1)	0.215
Anti-metabolites	1/11 (9.1)	2/8 (25.0)	2/9 (22.2)	1/5 (20.0)	6/33 (18.2)	0.810
Glucocorticoids	6/11 (54.5)	6/8 (75.0)	5/9 (55.6)	3/5 (60.0)	20/33 (60.6)	0.893
DMARDs	6/11 (54.5)	0/8 (0)	3/9 (33.3)	0/5 (0)	9/33 (27.3)	0.026
HIV	1/52 (1.9)	1/23 (4.3)	1/46 (2.2)	1/41 (2.4)	4/161 (2.5)	0.168
Inflammation [§]	26/52 (50.0)	13/24 (54.2)	25/47 (53.2)	27/44 (61.4)	91/165 (55.2)	0.731
ESR	25.5±31.6	41.1±30.9	35.8±35.5	34.0±34.6	32.4±33.3	0.607
CRP	35.0±76.1	22.4±28.3	17.9±36.0	13.7±16.3	23.1±50.2	0.520
Chest radiography lesions ^f						
Apical fibrotic scarring	0/40 (0)	1/17 (5.9)	1/40 (2.5)	1/36 (2.8)	3/133 (2.3)	0.340
Calcified nodules	3/37 (7.5)	2/17 (11.8)	2/40 (5.0)	4/36 (11.1)	11/133 (8.3)	0.682
Pleural thickening	0/40 (0)	1/17 (5.9)	1/40 (2.5)	2/36 (5.6)	4/133 (3.0)	0.377
BCG vaccinated	12/46 (26.1)	12/22 (54.5)	12/49 (24.5)	24/42 (57.1)	60/159 (37.7)	0.003
TST (mm)	2.9±5.2	4.1±8.5	6.2±11.0	18.9±11.5	7.6±10.9	<0.001
QFT result (IU·mL ⁻¹)	0.02±0.05	0.21±0.05	0.25±0.06	3.27±3.13	0.97±2.10	<0.001

Data are presented as mean±SD or n/N (%), unless otherwise stated. TB: tuberculosis; DMARD: disease-modifying anti-rheumatic drug; ESR: erythrocyte sedimentation rate; CRP: C-reactive protein; BCG: bacille Calmette-Guérin; TST: tuberculin skin test. [#]: individuals with “Profile+” and “Profile-” are those predicted to have TB infection or not, respectively, by regression analysis; [¶]: defined as a country with a TB incidence ≥ 40 cases of active TB per 100 000 inhabitants; ^{*}: only includes individuals from whom type and dose of immunosuppressive therapy could be retrieved; [§]: defined as either CRP ≥ 10 mg·L⁻¹ and/or ESR ≥ 20 mm·h⁻¹; ^f: lesions on chest radiography possible suggestive of prior TB infection.

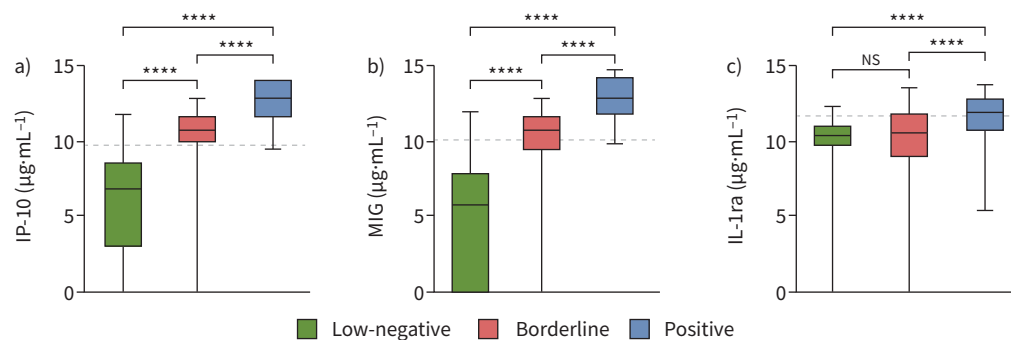


FIGURE 5 Biomarker values by QuantiFERON-TB Gold Plus categories: **a)** 10 kDa interferon (IFN)- γ -induced protein (IP-10), **b)** monokine induced by IFN- γ (MIG) and **c)** interleukin-1 receptor antagonist (IL-1ra). Biomarker values (medians and quartiles) are displayed as boxes, with whiskers representing 5–95% percentiles. Significant differences between groups were calculated using the Mann-Whitney U-test. Dashed lines represent the optimal cut-off (Youden index) derived from receiver operating characteristic curves (supplementary figure S3). All data were \log_2 -transformed prior to analysis. Includes all data. ****: $p < 0.0001$; NS: nonsignificant.

Discussion

After analysing a broad set of cytokines, chemokines and growth factors, we identified three biomarkers (IP-10, MIG and IL-1ra) in *M. tuberculosis*-specific antigen-stimulated QFT tubes that could discriminate patients with TB infection from likely uninfected individuals, in addition to the universally used biomarker IFN- γ . Subsequent LASSO regression analysis created a simple prediction model, including only MIG and IP-10. Application of this model on patients with low-negative and positive QFT results revealed that it was highly accurate in both detecting and “excluding” TB infection. Among patients with a borderline QFT result, this model predicted that nearly two-thirds of all patients with a borderline QFT result were likely *M. tuberculosis* infected, corroborated by combined MIG/IP-10 positivity independent of the model, which is in accordance with the previously reported proportion of infected individuals among those with a borderline QFT result [18, 19].

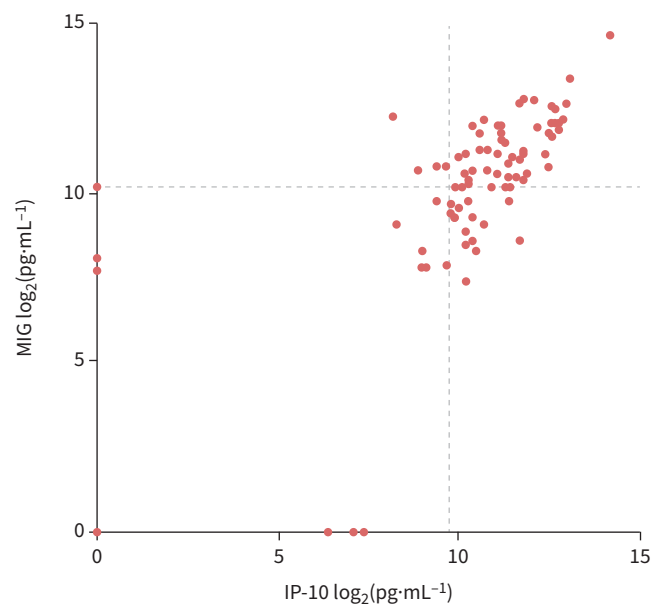


FIGURE 6 10 kDa interferon (IFN)- γ -induced protein (IP-10) and monokine induced by IFN- γ (MIG) levels among borderline QuantiFERON-TB Gold Plus (QFT) samples. IP-10 and MIG levels of all 87 patients with a borderline QFT result. Dashed lines represent the optimal cut-offs (Youden index) derived from receiver operating characteristic curves (supplementary figure S3). All data were \log_2 -transformed prior to analysis. Includes all data.

All of the biomarkers in our model are involved in the host's defence against an infection with *M. tuberculosis*. During infection, IP-10 attracts T-helper type 1 cells to the site of inflammation where it binds a receptor shared with MIG: CXCR3 [23, 24]. CXCR3 is an important receptor in mediating both innate and adaptive immune responses [24]. CXCR3-expressing T-cells were found in the lungs of infected mice, as well as in necrotic granulomas in nonhuman primate models [25, 26]. Absence of IP-10 increases *M. tuberculosis* bacillary growth *in vitro* [27]. The role of MIG during infection is similar to that of IP-10 [24]. However, the secretion of IP-10 is stimulated by more cytokines than just IFN- γ , whereas the secretion of MIG almost exclusively depends on IFN- γ [28]. Thus, IP-10 and MIG appear to have a fundamental role in the formation of granulomas and subsequent host defence against *M. tuberculosis*. Of note, IL-1ra inhibits the pro-inflammatory cytokine IL-1 and has been hypothesised to be an integral, self-limiting feature of alveolar macrophages [28, 29]. Unlike IP-10 and MIG, the secretion of IL-1ra is not directly stimulated by IFN- γ .

Both IP-10 and MIG have been shown to be accurate alternatives for IFN- γ in diagnosing or excluding TB infection in several previous studies [30, 31]. A recent systematic review and meta-analysis found a good sensitivity (85%) and moderate specificity (63%) of IP-10 in regular blood collection tubes for culture-positive active TB [32]. Among individuals with TB infection (based on a positive TST or QFT result), pooled sensitivity and specificity were 85% and 89%, respectively [33]. MIG levels are also high in patients with TB. A systematic review and meta-analysis on the predictive accuracy of MIG in antigen-stimulated and unstimulated supernatants among patients with culture-positive TB found that pooled sensitivity and specificity were both 84% [34]. A prior study investigated the diagnostic accuracy of IP-10/MIG values for TB infection in antigen-stimulated QFT tubes derived from healthcare workers classified as those without TB infection (defined by a QFT result $<0.1 \text{ IU}\cdot\text{mL}^{-1}$ and TST result $<10 \text{ mm}$), undetermined TB infection (defined by a QFT result ≥ 0.1 and $<0.35 \text{ IU}\cdot\text{mL}^{-1}$ and/or a TST $>10 \text{ mm}$) and with TB infection (QFT result $\geq 0.35 \text{ IU}\cdot\text{mL}^{-1}$) [35]. They found that a combined two-by-two cut-off (derived from individual ROC curves) of both IP-10 and MIG resulted in an increased specificity of 88%, while sensitivity for TB infection remained 100% in the group with undetermined TB infection, which is similar to our findings. Recently, a study in pregnant women with a borderline QFT result in Ethiopia found that 41% of these women had a high likelihood of TB infection based on a *k*-nearest neighbour analysis [36]. However, this percentage included predominantly patients with a borderline QFT result above the cut-off (up to $0.70 \text{ IU}\cdot\text{mL}^{-1}$), while our study included only borderline QFT results below the cut-off. The diagnostic accuracy of IL-1ra for *M. tuberculosis* infection has not yet been studied.

The predicted percentage (68%) of *M. tuberculosis*-infected patients despite a QFT result just below the cut-off is corroborated by estimations in our previous studies. We first found that borderline QFT results occurred in three-fold excess over random test variability [19]; the second study corroborated the latter and found a positive TST and/or T-SPOT.TB result in two-thirds of all patients with a borderline QFT result [18].

A key limitation in this study is the lack of a gold diagnostic standard for TB infection, which is inherent to the subject of study. Our reference group for TB infection consisted of individuals with a positive QFT result, while individuals with a low-negative QFT result were assumed uninfected. During validation, the model predicted three out of 19 patients with a low-negative QFT result to be likely infected. Interestingly, one of the three had been exposed to someone with active TB and had a positive TST result, highlighting the strength of the model in identifying possible false-negative QFT results. A second key limitation encompasses the lack of an independent validation cohort with low TB risk individuals with a low-negative QFT result for the most ideal assessment of the model's (already high) specificity. However, only eight subjects with a low-negative QFT result originated from a TB endemic area and none developed reactivation TB during follow-up. However, more data are needed to evaluate the use of these biomarkers in individuals with a low-negative QFT result. A strength of our study is that it included patient samples from three distinct laboratory sites at three different hospitals, which resulted in a heterogeneous cohort. Our population itself was another strength of this study because it included patients with autoimmune diseases and immunosuppressive therapy, making our findings more likely to be clinically useful, as our targeted population (those at increased risk of reactivation) often includes patients with such conditions. Also, the accuracy of IP-10 and MIG as individual biomarkers for TB infection was high, likely increasing the concomitant accuracy of our model, including for borderline QFT positives. Our prediction model attributes a relatively small coefficient to IP-10 compared with that of MIG. However, this is most likely due to the intrinsic aspects of the LASSO regression method. In case of collinearity, LASSO tends to assign a relatively low coefficient to one variable, in this case IP-10. However, our study clearly showed that IP-10 was strongly correlated with MIG and thus, according to our model, likely a good predictor for responses as well (figures 3a and 4a and b). Our results show that European Union *in vitro* diagnostic medical device regulation IVDR 2017/746-compliant IP-10 and/or MIG assays are urgently needed.

Currently, there are two *in vitro* tests under development using IP-10 as a readout, one based on ELISA and one based on a lateral flow assay (IP-10 IGRA LF), both of which are developed by R-Biopharm (Darmstadt, Germany) [37, 38]. In light of the several aforementioned studies that proved high diagnostic accuracy of IP-10 for TB infection and our findings, such an IP-10-based assay could be a useful tool to increase the diagnostic sensitivity for TB infection, in particular among those with a borderline QFT result. These findings, if corroborated by different studies (*e.g.* conducted in TB endemic countries), may also encourage the QFT's manufacturer to include a second biomarker, IP-10/MIG, as a readout for a future QFT test.

The clinical relevance of these findings for an individual depends on the assumed risk of progression to active TB. For example, during occupational screening of healthcare workers in low TB endemic areas, the benefits of preventive treatment for those with a borderline QFT result do not outweigh the harm of potential side-effects. In contrast, targeted preventive treatment of patients eligible for treatment with immunosuppressive agents who have a borderline QFT result and IP-10 or MIG response will reduce the risk of reactivation TB significantly without exposing uninfected patients to drug toxicity or interactions and delayed start of their immunosuppressive treatment. If our results can be corroborated in a larger cohort, additional measurement of IP-10 and/or MIG in QFT supernatants may well be able to refine clinical decision making by increasing diagnostic accuracy in the borderline QFT-zone.

Conflict of interest: T.H.M. Ottenhoff reports grants from NWO-TTW (PI: T.H.M. Ottenhoff), Dutch Government, Technical Sciences; ZonMw (PI: T.H.M. Ottenhoff), Dutch Government (ZonMw); IMI2 HOR2020 VSV EBOPLUS (PI: C.A. Siegrist), European Commission HOR2020 IMI2 Program; NWO-TTW (PI: J. Bouwstra), Dutch Government, Technical Sciences; NWO-TTW (PI: T.H.M. Ottenhoff), Dutch Government, Technical Sciences, NACTAR Program; NWO-Chemical Sciences (PI: A. Minnaard), Dutch Government, Technical Sciences; EC HOR2020 TRANSVAC2 (PI: European Vaccine Initiative (EVI)), European Commission HOR2020 Program; IMI2 EC HOR2020 Respiri-TB (PI: M. Lamers), European Commission HOR2020 IMI2 Program; IMI2 EC HOR2020 Respiri-NTM (PI: M. Lamers), European Commission HOR2020 IMI2 Program; NIH (PI: T.H.M. Ottenhoff); NIH, NIAID, grant: 1R01AI141315-01A1; EC HOR2020 SMA-TB (PI: C. Vilaplana); European Commission HOR2020 Program; leadership at the Tuberculosis Vaccine Initiative (TBVI; www.tbvi.eu); outside the submitted work. S.A. Joosten reports grants from NIH (PI: T.H.M. Ottenhoff; co-PI: S.A. Joosten); NIH, NIAID, grant: 1R01AI141315-01A1; outside the submitted work. S.M. Arend reports travel support from Oxford Immunotec, outside the submitted work. All other authors have nothing to disclose.

Support statement: This work was supported by the European Commission Horizon 2020 research project TBVAC2020 consortium (number 643381 to T.H.M. Ottenhoff). The text represents the authors' views and does not necessarily represent a position of the Commission who will not be liable for the use made of such information. Funding information for this article has been deposited with the Crossref Funder Registry.

References

- 1 Solovic I, Sester M, Gomez-Reino JJ, *et al.* The risk of tuberculosis related to tumour necrosis factor antagonist therapies: a TBNET consensus statement. *Eur Respir J* 2010; 36: 1185–1206.
- 2 Leung CC, Rieder HL, Lange C, *et al.* Treatment of latent infection with *Mycobacterium tuberculosis*: update 2010. *Eur Respir J* 2011; 37: 690–711.
- 3 Hasan T, Au E, Chen S, *et al.* Screening and prevention for latent tuberculosis in immunosuppressed patients at risk for tuberculosis: a systematic review of clinical practice guidelines. *BMJ Open* 2018; 8: e022445.
- 4 American Thoracic Society. Targeted tuberculin testing and treatment of latent tuberculosis infection. *Am J Respir Crit Care Med* 2000; 161: S221–S247.
- 5 Mazurek GH, Jereb J, Vernon A, *et al.* Updated guidelines for using interferon gamma release assays to detect *Mycobacterium tuberculosis* infection – United States, 2010. *MMWR Recomm Rep* 2010; 59: 1–25.
- 6 Oh CE, Ortiz-Brizuela E, Bastos ML, *et al.* Comparing the diagnostic performance of QuantiFERON-TB Gold Plus to other tests of latent tuberculosis infection: a systematic review and meta-analysis. *Clin Infect Dis* 2021; 73: E1116–E1125.
- 7 Pai M, Zwerling A, Menzies D. Systematic review: T-cell-based assays for the diagnosis of latent tuberculosis infection: an update. *Ann Intern Med* 2008; 149: 177–184.
- 8 Sotgiu G, Saderi L, Petruccioli E, *et al.* QuantiFERON TB Gold Plus for the diagnosis of tuberculosis: a systematic review and meta-analysis. *J Infect* 2019; 79: 444–453.
- 9 Herrera V, Perry S, Parsonnet J, *et al.* Clinical application and limitations of interferon-gamma release assays for the diagnosis of latent tuberculosis infection. *Clin Infect Dis* 2011; 52: 1031–1037.
- 10 Metcalfe JZ, Cattamanchi A, McCulloch CE, *et al.* Test variability of the QuantiFERON-TB Gold In-Tube assay in clinical practice. *Am J Respir Crit Care Med* 2013; 187: 206–211.

- 11 Nienhaus A, Ringshausen FC, Costa JT, *et al.* IFN-gamma release assay versus tuberculin skin test for monitoring TB infection in healthcare workers. *Expert Rev Anti Infect Ther* 2013; 11: 37–48.
- 12 Schablon A, Nienhaus A, Ringshausen FC, *et al.* Occupational screening for tuberculosis and the use of a borderline zone for interpretation of the IGR in German healthcare workers. *PLoS One* 2014; 9: e115322.
- 13 Tagmouti S, Slater M, Benedetti A, *et al.* Reproducibility of interferon gamma (IFN- γ) release assays. A systematic review. *Ann Am Thorac Soc* 2014; 11: 1267–1276.
- 14 Nemes E, Rozot V, Geldenhuys H, *et al.* Optimization and interpretation of serial QuantiFERON testing to measure acquisition of *Mycobacterium tuberculosis* infection. *Am J Respir Crit Care Med* 2017; 196: 638–648.
- 15 Banaei N, Pai M. Detecting new *Mycobacterium tuberculosis* infection. Time for a more nuanced interpretation of QuantiFERON conversions. *Am J Respir Crit Care Med* 2017; 196: 546–547.
- 16 Chiacchio T, Petruccili E, Vanini V, *et al.* Characterization of QuantiFERON-TB-Plus results in latent tuberculosis infected patients with or without immune-mediated inflammatory diseases. *J Infect* 2019; 79: 15–23.
- 17 Petruccioli E, Chiacchio T, Navarra A, *et al.* Effect of HIV infection on QuantiFERON-plus accuracy in patients with active tuberculosis and latent infection. *J Infect* 2020; 80: 536–546.
- 18 Uzorka JW, Bossink AWJ, Franken WPJ, *et al.* Borderline QuantiFERON results and the distinction between specific responses and test variability. *Tuberculosis* 2018; 111: 102–108.
- 19 Uzorka JW, Kroft LJM, Bakker JA, *et al.* Proof of concept that most borderline Quantiferon results are true antigen-specific responses. *Eur Respir J* 2017; 50: 1701630.
- 20 Uzorka JW, Delfos NM, Witte AMC, *et al.* Tuberculosis after a borderline QuantiFERON result during screening before infliximab. *Eur Respir J* 2018; 52: 1800913.
- 21 van Meijgaarden KE, Khatri B, Smith SG, *et al.* Cross-laboratory evaluation of multiplex bead assays including independent common reference standards for immunological monitoring of observational and interventional human studies. *PLoS One* 2018; 13: e0201205.
- 22 Goeman JJ, Finos L. The inheritance procedure: multiple testing of tree-structured hypotheses. *Stat Appl Genet Mol* 2012; 11: 11.
- 23 Whittaker E, Gordon A, Kampmann B. Is IP-10 a better biomarker for active and latent tuberculosis in children than IFNgamma? *PLoS One* 2008; 3: e3901.
- 24 Khader SA, Bell GK, Pearl JE, *et al.* IL-23 and IL-17 in the establishment of protective pulmonary CD4⁺ T cell responses after vaccination and during *Mycobacterium tuberculosis* challenge. *Nat Immunol* 2007; 8: 369–377.
- 25 Lim J, Derrick SC, Kolibab K, *et al.* Early pulmonary cytokine and chemokine responses in mice immunized with three different vaccines against *Mycobacterium tuberculosis* determined by PCR array. *Clin Vaccine Immunol* 2009; 16: 122–126.
- 26 Lin PL, Pawar S, Myers A, *et al.* Early events in *Mycobacterium tuberculosis* infection in cynomolgus macaques. *Infect Immun* 2006; 74: 3790–3803.
- 27 Agrawal N, Bhattacharyya C, Mukherjee A, *et al.* Dissecting host factors that regulate the early stages of tuberculosis infection. *Tuberculosis* 2016; 100: 102–113.
- 28 Chegou NN, Heyckendorf J, Walz G, *et al.* Beyond the IFN-gamma horizon: biomarkers for immunodiagnosis of infection with *Mycobacterium tuberculosis*. *Eur Respir J* 2014; 43: 1472–1486.
- 29 Lee JH, Chang JH. Changes of plasma interleukin-1 receptor antagonist, interleukin-8 and other serologic markers during chemotherapy in patients with active pulmonary tuberculosis. *Korean J Intern Med* 2003; 18: 138–145.
- 30 Ruhwald M, Aabye MG, Ravn P. IP-10 release assays in the diagnosis of tuberculosis infection: current status and future directions. *Expert Rev Mol Diagn* 2012; 12: 175–187.
- 31 Ruhwald M, Bjerregaard-Andersen M, Rabna P, *et al.* IP-10, MCP-1, MCP-2, MCP-3, and IL-1RA hold promise as biomarkers for infection with *M. tuberculosis* in a whole blood based T-cell assay. *BMC Res Notes* 2009; 2: 19.
- 32 Santos VS, Goletti D, Kontogianni K, *et al.* Acute phase proteins and IP-10 as triage tests for the diagnosis of tuberculosis: systematic review and meta-analysis. *Clin Microbiol Infect* 2019; 25: 169–177.
- 33 Qiu X, Tang Y, Yue Y, *et al.* Accuracy of interferon-gamma-induced protein 10 for diagnosing latent tuberculosis infection: a systematic review and meta-analysis. *Clin Microbiol Infect* 2019; 25: 667–672.
- 34 Li Y, He D, Che Y, *et al.* Monokine induced by gamma interferon for detecting pulmonary tuberculosis: a diagnostic meta-analysis. *Medicine* 2020; 99: e23302.
- 35 Rubbo PA, Nagot N, Le Moing V, *et al.* Multicytokine detection improves latent tuberculosis diagnosis in health care workers. *J Clin Microbiol* 2012; 50: 1711–1717.
- 36 Tesfaye F, Sturegard E, Walles J, *et al.* Alternative biomarkers for classification of latent tuberculosis infection status in pregnant women with borderline Quantiferon plus results. *Tuberculosis* 2020; 124: 101984.
- 37 Hamada Y, Cirillo DM, Matteelli A, *et al.* Tests for tuberculosis infection: landscape analysis. *Eur Respir J* 2021; 58: 2100167.
- 38 Migliori GB, Ong CWM, Petrone L, *et al.* The definition of tuberculosis infection based on the spectrum of tuberculosis disease. *Breathe* 2021; 17: 210079.