



Measurement of hypoxia in the lung in idiopathic pulmonary fibrosis: a matter of control

Copyright ©The authors 2022.

This version is distributed under the terms of the Creative Commons Attribution Non-Commercial Licence 4.0. For commercial reproduction rights and permissions contact permissions@ersnet.org

Received: 13 Oct 2021
Accepted: 18 Nov 2021

To the Editor:

We read with great interest the paper by PORTER *et al.* [1] published in the October 2021 issue of the *European Respiratory Journal*. The authors' aim was to explore the potential benefit of the hypoxia tracer [¹⁸F]fluoromisonidazole ([¹⁸F]F-MISO) in idiopathic pulmonary fibrosis (IPF). Given the lack of non-invasive imaging tools for the diagnosis and/or the follow-up of patients with IPF, this study appears to be an essential first step towards the personalised management of IPF patients through imaging biomarkers for early/active fibrosis. *In vivo* molecular imaging, in particular positron emission tomography (PET), has become a crucial tool in preclinical research, clinical trials and medical practice, especially in the field of oncology. In lung fibrosis, recent advances have been made with the aim of developing molecular imaging tools in preclinical models, a necessary step toward clinical certification [2]. Among tracers validated at the preclinical level, imaging probes targeting collagen (⁶⁸Ga-CBP8 [3]), integrins ([¹⁸F]FB-A20FMDV2 [4]) and glucose metabolism ([¹⁸F]FDG [5]) have been successfully evaluated in clinical trials and may ultimately improve IPF management.

While chronic hypoxia of the lung is a significant clinical feature in patients with IPF, the study by PORTER *et al.* [1] is the first to explore the potential role of the hypoxia tracer [¹⁸F]F-MISO in these patients. However, the results of this study were disappointingly far from our expectations considering that high levels of hypoxia biomarkers have been found in IPF patients, suggesting a hypoxic microenvironment in the IPF lung [6]. In addition, our group previously suggested that [¹⁸F]F-MISO imaging could be a promising tool for early detection and monitoring in a preclinical model of lung fibrosis [7]. Although we are aware that our preclinical results may not be entirely relevant for human IPF, we believe that the study from PORTER *et al.* [1] may suffer from flaws that could explain, at least in part, their underwhelming results. In our opinion, the main issue resides in the use of lung areas with a "normal" appearance as controls for fibrotic areas. When they used this control, PORTER *et al.* [1] assumed that the regions of IPF lungs that appear to be normal are *de facto* not hypoxic. We believe that this assumption may be incorrect since we demonstrated in our preclinical results that there was also an increase in [¹⁸F]F-MISO lung uptake in areas that seemed "normal" on computed tomography (figure 1). These data are in line with other studies demonstrating that hypoxia inducible transcription factor (HIF)-1 α and CA-IX are upregulated, not only in areas of active fibrosis, but also within areas of IPF lungs that appear histologically normal [8]. These findings suggest that the activation of hypoxia signalling is an early event that drives the remodelling of areas in the IPF lung that are not yet fibrotic, thus promoting disease progression. As an alternative, considering that hypoxic volumes are more localised in lung cancer than in IPF, seemingly "normal" zones distant from tumours in lung cancer patients would have been much more reliable controls but would require the inclusion of more than two patients to be statistically relevant. Further, PORTER *et al.* [1] do not specify whether the IPF patients included in the work were undertaking anti-fibrotic treatment. This question may be crucial considering that we demonstrated that [¹⁸F]F-MISO uptake was dramatically decreased by both nintedanib and pirfenidone in preclinical models [7], and the same effect has been reported in cancer [9].

In addition, while we understand that average pulmonary uptake (SUV_{mean}) values may have been more useful in this study than SUV_{max} (classically used for [¹⁸F]F-MISO in oncology) considering that IPF is a diffuse disease, no comparison between SUV_{mean} from IPF and lung tumours is provided. These data



Shareable abstract (@ERSpublications)

Despite the discouraging results provided by Porter and co-workers, we believe that there is room for improvements, mainly by using better controls, which may ultimately lead to more promising outcomes for the use of hypoxia-focused imaging in IPF patients <https://bit.ly/30Ku2AV>

Cite this article as: Bellaye P-S, Beltramo G, Burgy O, *et al.* Measurement of hypoxia in the lung in idiopathic pulmonary fibrosis: a matter of control. *Eur Respir J* 2022; 59: 2102711 [DOI: 10.1183/13993003.02711-2021].



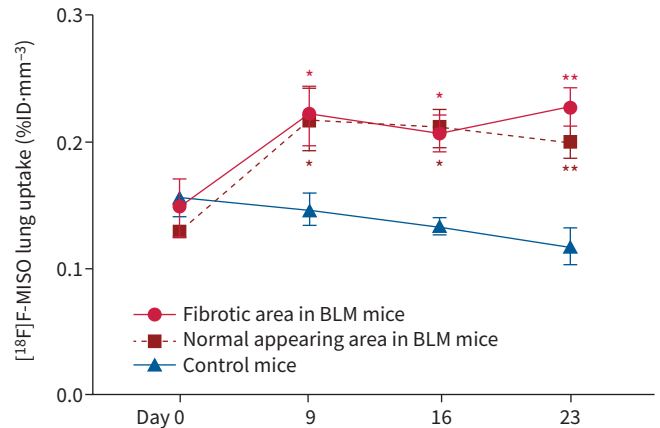


FIGURE 1 Fluorine-18-labelled fluoromisonidazole ($[^{18}\text{F}]\text{F-MISO}$) lung uptake is upregulated in seemingly normal and fibrotic lung areas in bleomycin (BLM)-induced lung fibrosis. Graph represents the evolution of $[^{18}\text{F}]\text{F-MISO}$ lung uptake (% injected dose (ID) per mm^3) in BLM-receiving mice at day 0 (baseline before BLM), and days 9, 16 and 23 in normal appearing and fibrotic lung areas (segmented on computed tomography images). $[^{18}\text{F}]\text{F-MISO}$ lung uptake in mice receiving NaCl serves as control. Results are presented as mean \pm SEM, n=5 per group. *: $p<0.05$; **: $p<0.01$, for statistical comparison between BLM and control mice. Data from TANGUY *et al.* [7].

could be used to compare the level of hypoxia in tumours and in IPF lungs. Even in hypoxic tumours, $[^{18}\text{F}]\text{F-MISO}$ uptake can be relatively low (*e.g.* SUV_{mean} between 1.5 and 2 [10]), and one could imagine that the SUV_{mean} presented here (1.6 and 1.55 for control and fibrotic areas, respectively) could mean that both normal appearing and fibrotic lung areas are hypoxic in IPF patients. Therefore, considering the diffuse nature of IPF and the relatively low uptake of $[^{18}\text{F}]\text{F-MISO}$, an imaging protocol including a PET scan at 120 min post-injection, which is a common schedule for cancer trials, may have improved SUV values and would have been easier to compare with the existing data in cancer.

While we understand the difficulty of including patients in this type of clinical trial, the heterogeneity of lung function parameters in the IPF cohort may be an additional drawback of the current study. Heterogeneity may be beneficial in a large clinical trial, but it may also hide potentially interesting results in a particular subset of patients (*e.g.* mild *versus* severe fibrosis) in trials with a small number of patients. A correlation between $[^{18}\text{F}]\text{F-MISO}$ SUV_{mean} and forced vital capacity and/or transfer factor of the lung for carbon monoxide would provide a better idea of whether hypoxia is related to disease stage or severity, as is the case in preclinical models of lung fibrosis [7] and in oncology.

Despite the discouraging results reported by PORTER *et al.* [1], we strongly believe that there is room for improvement, which may ultimately lead to more promising outcomes for the use of hypoxia-focused imaging in IPF patients.

Pierre-Simon Bellaye^{1,2}, Guillaume Beltramo^{2,3}, Olivier Burgy^{2,3}, Bertrand Collin^{1,4}, Alexandre Cochet^{1,5} and Philippe Bonniaud^{2,3}

¹Centre George-François Leclerc, Service de médecine nucléaire, Plateforme d'imagerie et de radiothérapie précliniques, Dijon, France. ²Centre de Référence Constitutif des Maladies Pulmonaires Rares de l'Adultes de Dijon, réseau OrphaLung, Filère RespiFil, Centre Hospitalier Universitaire de Bourgogne, Dijon, France. ³INSERM U1231, Equipe HSP-pathies, Dijon, France. ⁴Institut de Chimie Moléculaire de l'Université de Bourgogne, UMR CNRS 6302, Université de Bourgogne Franche-Comté, Dijon, France. ⁵ImVIA, EA 7535, Université de Bourgogne, Dijon, France.

Corresponding author: Pierre-Simon Bellaye (psbellaye@cgfl.fr)

Acknowledgements: The authors thank Suzanne Rankin for reviewing the English.

Conflict of interest: P-S. Bellaye reports a research grant paid to his institution (Centre George François Leclerc, Dijon) from the ANR (HYMAGE-IPF – ANR-20-CE17-0005). G. Beltramo reports a research grant paid to his institution (CHU Dijon) from GIRCI Est (FIPOXY). O. Burgy has nothing to disclose. B. Collin has nothing to disclose. A. Cochet has nothing to disclose. P. Bonniaud reports receiving support for attending medical and research conferences and personal fees for advisory board work from Roche and Boehringer.

References

- 1 Porter JC, Win T, Erlandsson K, *et al.* Measurement of hypoxia in the lung in idiopathic pulmonary fibrosis: an F-MISO PET/CT study. *Eur Respir J* 2021; 58: 2004584.
- 2 Désogère P, Montesi SB, Caravan P. Molecular probes for imaging fibrosis and fibrogenesis. *Chemistry* 2019; 25: 1128–1141.
- 3 Montesi SB, Izquierdo-Garcia D, Désogère P, *et al.* Type I collagen-targeted positron emission tomography imaging in idiopathic pulmonary fibrosis: first-in-human studies. *Am J Respir Crit Care Med* 2019; 200: 258–261.
- 4 Lukey PT, Coello C, Gunn R, *et al.* Clinical quantification of the integrin $\alpha v\beta 6$ by [18F]FB-A20FMDV2 positron emission tomography in healthy and fibrotic human lung (PETAL Study). *Eur J Nucl Med Mol Imaging* 2020; 47: 967–979.
- 5 Win T, Sreaton NJ, Porter JC, *et al.* Pulmonary 18F-FDG uptake helps refine current risk stratification in idiopathic pulmonary fibrosis (IPF). *Eur J Nucl Med Mol Imaging* 2018; 45: 806–815.
- 6 Kottmann RM, Kulkarni AA, Smolnycki KA, *et al.* Lactic acid is elevated in idiopathic pulmonary fibrosis and induces myofibroblast differentiation via pH-dependent activation of transforming growth factor- β . *Am J Respir Crit Care Med* 2012; 186: 740–751.
- 7 Tanguy J, Goirand F, Bouchard A, *et al.* [18F]FMISO PET/CT imaging of hypoxia as a non-invasive biomarker of disease progression and therapy efficacy in a preclinical model of pulmonary fibrosis: comparison with the [18F]FDG PET/CT approach. *Eur J Nucl Med Mol Imaging* 2021; 48: 3058–3074.
- 8 Bodempudi V, Hergert P, Smith K, *et al.* miR-210 promotes IPF fibroblast proliferation in response to hypoxia. *Am J Physiol Lung Cell Mol Physiol* 2014; 307: L283–L294.
- 9 Quintela-Fandino M, Lluch A, Manso L, *et al.* 18F-fluoromisonidazole PET and activity of neoadjuvant nintedanib in early HER2-negative breast cancer: a window-of-opportunity randomized trial. *Clin Cancer Res* 2017; 23: 1432–1441.
- 10 Fleming IN, Manavaki R, Blower PJ, *et al.* Imaging tumour hypoxia with positron emission tomography. *Br J Cancer* 2015; 112: 238–250.