





Interleukin- 1α : a key player for epithelial-to-mesenchymal signalling in COPD?

Antoine Froidure^{1,2,3,4}, Maha Zohra Ladjemi^{3,4} and Charles Pilette^{1,2}

Affiliations: ¹Institut de Recherche Expérimentale et Clinique, Pôle de Pneumologie, ORL et Dermatologie, Université Catholique de Louvain, Brussels, Belgium. ²Cliniques Universitaires Saint-Luc, Service de Pneumologie, Brussels, Belgium. ³UMR Inserm U1152, Labex Inflammex, Université Paris 7, Paris, France. ⁴These authors contributed equally to this manuscript.

Correspondence: Charles Pilette, Université Catholique de Louvain, Avenue Hippocrate 54/B1-54.04, Brussels B-1200, Belgium. E-mail: charles.pilette@uclouvain.be



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Interleukin-1 alpha released by airway epithelial cells may instruct fibroblasts to become pro-inflammatory in COPD http://ow.ly/CLYK301tyOj

The lungs are continuously exposed to inhaled particles and irritants. Tobacco smoking is the leading cause of several airway diseases, including lung cancer, idiopathic pulmonary fibrosis and chronic obstructive pulmonary disease (COPD). In COPD, repeated exposure to cigarette smoke induces chronic inflammation and structural remodelling of the airways, with peribronchial fibrosis and epithelial-to-mesenchymal transition [1], resulting in (mostly irreversible) airway obstruction. Thus, abnormal airway responses to noxious gases probably results from aberrant signalling and crosstalk between epithelial, mesenchymal and immune cells.

In this issue of the European Respiratory Journal, Osei et al. [2] describe how epithelial cells signal to fibroblasts through interleukin (IL)- 1α , with relevance to COPD. Using co-cultures of human primary cells (bronchial epithelial cells (BECs) and lung fibroblasts) and human cell lines (MRC5 fetal fibroblasts and 16HBE epithelial cells), the authors show that co-culture of BECs and fibroblasts induced the production of heat shock protein (Hsp)70, IL-8/CXCL8 and IL- 1β by lung fibroblasts, while decreasing expression of α -smooth muscle actin, transforming growth factor (TGF)- β 1 and extracellular matrix proteins. These effects, which were not different between control- and COPD-derived cells, were recapitulated by using BEC (16HBE)-conditioned medium, suggesting the requirement for soluble factor(s). Using neutralising antibodies, the authors went on to demonstrate that release of CXCL8 and Hsp70 by fibroblasts in the co-culture system was independent of IL- 1β and prostaglandin E2 but dependent on IL- 1α . In addition, IL- 1α release by BECs was enhanced in COPD-derived cells by exposure to cigarette smoke extract. Although this *in vitro* model may not completely recapitulate the *in vivo* situation, it provides important new insights regarding the signals delivered by airway epithelial cells and regulating mesenchymal cells.

There is accumulating evidence that BECs are key cells in frontline defence that directly signal to immune cells, in particular antigen-presenting cells [3, 4]. Several previous works identified that BECs shape the immune response engaged by pathogens [5]. For instance, BECs exposed to *Klebsiella pneumoniae* regulate local immune responses notably by acting on the myeloid dendritic cell network [6]. Respiratory syncytial virus-infected BECs regulate CD8⁺ T-cell activation and antiviral activity, according to changes in epithelial expression of the "checkpoint" immune receptor PD-L1 [7]. BECs also regulate dendritic cell differentiation and maturation as well as responsiveness to lipopolysaccharide [8], and inhibit T-cell recall responses towards common aeroallergens in order to ensure mucosal homeostasis and dampen allergic responses [9]. It has also been shown in asthma that BECs may suppress constitutive and IgE-dependent histamine release by lung mast cells [10], further highlighting the central role of the epithelium in sensing and shaping mucosal danger signalling. We recently showed that BECs from COPD patients also imprint

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B-cells with signals promoting maturation into IgA-producing plasma cells, while cigarette smoke partly counteracts this IgA-promoting effect [11]. This effect was at least partly related to an intrinsic ability of COPD BECs to produce increased amounts of certain cytokines (IL-6 and BAFF; B-cell activating factor) and to induce B-cell expression of TACI (transmembrane activator and CAML interactor). Osei *et al.* [2] also addressed intrinsic epithelial changes, showing that an amplified IL-1 α response to cigarette smoke extract is seen in BECs from COPD patients as compared to cells from controls.

Standing at the interface between the environment and host tissues, the innate immune system may be engaged through several pattern recognition receptors (PRRs) to pathogen-associated molecular patterns (PAMPs), namely Toll-like receptors (TLRs), NOD-like receptors, protein-activated receptors and RIG-like receptors. Ligation of these receptors triggers an immune response, initiated by the release of epithelial cytokines including IL-6, thymic stromal lymphopoietin (TSLP), IL-25 and IL-33 [12]. Upstream of these cytokines, the activation of NOD-like receptor family, pyrin domain containing 3 (NLRP3) inflammasome leads to the release of IL-1 α , IL-1 β and IL-18, acting as pyrogens and T-cell activators. Activated epithelial cells secrete pro-IL-1 β that is cleaved by caspase-1 to produce bioactive IL-1 β [13]. Despite recent evidence pointing to the involvement of NLRP3 inflammasome in asthma, COPD and idiopathic pulmonary fibrosis, and its close relationship with IL-1 β and the NLRP3 inflammasome, the role of IL-1 α in respiratory diseases has long been neglected. IL-1 α was described in 1985 when IL-1 was discovered to consist of two distinct proteins [14]. Studies of lung samples from COPD patients and from mice exposed to cigarette smoke as an experimental model of COPD showed increased expression of IL-1 α [15–17],

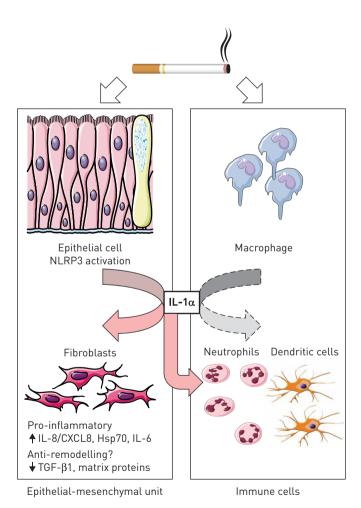


FIGURE 1 Interleukin (IL)- 1α as a key mediator of epithelial signalling to mesenchymal and immune cells. In chronic obstructive pulmonary disease, cigarette smoke probably triggers bronchial epithelial cells and alveolar macrophages to release IL- 1α following activation of NLRP3 (NOD-like receptor family, pyrin domain containing 3) inflammasome. IL- 1α mediates a shift in fibroblast phenotype towards pro-inflammatory fibroblasts releasing increased amounts of IL-8/CXCL8, which in turn attracts neutrophils in the airways. To what extent and how IL- 1α also regulates repair responses and matrix deposition [2, 20] remains to be studied, in order to clarify whether (and eventually under what circumstances) this cytokine is pro- or anti-fibrotic in the human lung.

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which mediates accumulation of neutrophils and dendritic cells in the murine model. IL- 1α expression was, however, mainly localised to haematopoietic cells such as alveolar macrophages [17, 18] and not to epithelial cells. In asthma, Willart *et al.* [19] demonstrated that epithelial activation by proteolytic allergens triggers the release of IL- 1α , which induces, through autocrine IL-1 receptor ligation, the release of pro-Th2 cytokines by BEC, ultimately promoting Th2-biased immune responses.

Osei et al. [2] went one step further by providing evidence of a key role for IL-1 α in mediating epithelial signalling to fibroblasts. They showed that IL-1 α switches on a differentiation programme in fibroblasts towards an inflammatory phenotype (with increased hsp70 and IL-8/CXCL8, and to some extent IL-6), while abrogating their production of matrix proteins. Suwara et al. [20] recently showed that human lung fibrobasts also switch towards inflammatory fibroblasts upon incubation with conditioned medium from damaged BECs, increasing their release of IL-8/CXCL8, MCP-1 and IL-6 that was abrogated by anti-IL-1 α or anti-IL-1Ra, but not anti-IL-1 β , antibodies. The inflammatory phenotype of fibrobasts was accentuated by concomitant activation of TLR3, through nuclear factor κ B signalling. Furthermore, a reduced collagen deposition was noticed in IL-1 α - $^{-1}$ - and IL-1R1- $^{-1}$ - mice following bleomycine exposure. The finding of Osei et al. [2], i.e. a reduced synthesis of matrix proteins upon IL-1 α activation, challenges this view of IL-1 α as a profibrotic mediator. Thus, whether IL-1 α is involved in the loss of small conducting airways or in peribronchial fibrosis observed in COPD [21] remains unclear. In cancer, BAE et al. [22] showed that IL-1 α may enhance oral squamous carcinoma-associated fibroblast proliferation through the secretion of CCL7, CXCL1 and IL-8/CXCL8 chemokines.

Interactions between epithelial and mesenchymal cells are critical during branching morphogenesis and lung development, and include in the adult lung several aspects including dynamic transition states (epithelial-to-mesenchymal and mesenchymal-to-epithelial transition) and paracrine regulation. In asthma, IL-4/IL-13 activate BECs to release TGF- β 2 which, in turn, stimulates myofibroblast transformation and secretion of matrix proteins and vascular mitogens (e.g. vascular endothelial growth factor, endothelin-1) [23], leading to the concept of "epithelial-mesenchymal trophic unit". This epithelial-mesenchymal signalling pathway should be further explored in chronic airway diseases such as COPD [24], notably by using multicellular culture models with primary cells as carried out by OSEI et al. [2].

Altogether, this work illustrates that repeated broncho-epithelial injury by cigarette smoke, in individuals with putative susceptibility factors (which remain largely unknown), may trigger an altered programming of airway epithelial cells resulting in chronic inflammasome activation which includes the release of IL- 1α (figure 1). Whereas this cytokine probably contributes to persistent accumulation of neutrophils by instructing a pro-inflammatory programming of fibroblasts, it remains to be determined to what extent it also contributes to altered repair responses and matrix deposition in conducting airways of the human COPD lung. An additional difficulty in COPD is the association in the same disease of opposed features of matrix turnover at different airway levels, namely matrix deposition in some conducting airways (while some other small airways may disappear) and matrix destruction in respiratory parts of the lung, or even between adjacent alveolar zones in the combined emphysema fibrosis phenotype [25]. Nevertheless, manipulation of the epithelial-fibroblast pathway should help the development of new therapeutic strategies, and the recent attempt to target the inflammasome in COPD [26] might open the way to new biotherapies in severe chronic obstructive lung disease.

References

- 1 Gohy ST, Hupin C, Fregimilicka C, et al. Imprinting of the COPD airway epithelium for dedifferentiation and mesenchymal transition. Eur Respir J 2015; 45: 1258–1272.
- Osei ET, Noordhoek JA, Hackett TL, et al. Interleukin-1α drives the dysfunctional cross-talk of the airway epithelium and lung fibroblasts in COPD. Eur Respir J 2016; 48: 359–369.
- Froidure A, Shen Č, Pilette C. Dendritic cells revisited in human allergic rhinitis and asthma. *Allergy* 2016; 71: 137–148.
- Gohy ST, Hupin C, Pilette C, et al. Chronic inflammatory airway diseases: the central role of the epithelium revisited. Clin Exp Allergy 2016; 46: 529–542.
- Hiemstra PS, McCray PB Jr, Bals R. The innate immune function of airway epithelial cells in inflammatory lung disease. *Eur Respir J* 2015; 45: 1150–1162.
- 6 Pichavant M, Taront S, Jeannin P, et al. Impact of bronchial epithelium on dendritic cell migration and function: modulation by the bacterial motif KpOmpA. J Immunol 2006; 177: 5912–5919.
- 7 Telcian AG, Laza-Stanca V, Edwards MR, et al. RSV-induced bronchial epithelial cell PD-L1 expression inhibits CD8+ T cell nonspecific antiviral activity. J Infect Dis 2011; 203: 85–94.
- 8 Rate A, Upham JW, Bosco A, et al. Airway epithelial cells regulate the functional phenotype of locally differentiating dendritic cells: implications for the pathogenesis of infectious and allergic airway disease. I Immunol 2009: 182: 72–83.
- Papazian D, Wagtmann VR, Hansen S, et al. Direct contact between dendritic cells and bronchial epithelial cells inhibits T cell recall responses towards mite and pollen allergen extracts in vitro. Clin Exp Immunol 2015; 181: 207–218.
- Martin N, Ruddick A, Arthur GK, et al. Primary human airway epithelial cell-dependent inhibition of human lung mast cell degranulation. PloS One 2012; 7: e43545.

- Ladjemi MZ, Lecocq M, Weynand B, et al. Increased IgA production by B-cells in COPD via lung epithelial 11 interleukin-6 and TACI pathways. Eur Respir J 2015; 45: 980-993
- Gregory LG, Jones CP, Walker SA, et al. IL-25 drives remodelling in allergic airways disease induced by house dust mite. Thorax 2013; 68: 82-90.
- De Nardo D, De Nardo CM, Latz E. New insights into mechanisms controlling the NLRP3 inflammasome and its role in lung disease. Am J Pathol 2014; 184: 42-54.
- March CJ, Mosley B, Larsen A, et al. Cloning, sequence and expression of two distinct human interleukin-1 14 complementary DNAs. Nature 1985; 315: 641-647.
- Doz E, Noulin N, Boichot E, et al. Cigarette smoke-induced pulmonary inflammation is TLR4/MyD88 and 15 IL-1R1/MyD88 signaling dependent. J Immunol 2008; 180: 1169-1178.
- Pauwels NS, Bracke KR, Dupont LL, et al. Role of IL-1alpha and the Nlrp3/caspase-1/IL-1beta axis in cigarette smoke-induced pulmonary inflammation and COPD. Eur Respir J 2011; 38: 1019-1028.
- Botelho FM, Bauer CM, Finch D, et al. IL-1alpha/IL-1R1 expression in chronic obstructive pulmonary disease and mechanistic relevance to smoke-induced neutrophilia in mice. PloS One 2011; 6: e28457.
- Nikota JK, Shen P, Morissette MC, et al. Cigarette smoke primes the pulmonary environment to IL-1alpha/ CXCR-2-dependent nontypeable Haemophilus influenzae-exacerbated neutrophilia in mice. J Immunol 2014; 193: 3134-3145
- Willart MA, Deswarte K, Pouliot P, et al. Interleukin-1alpha controls allergic sensitization to inhaled house dust mite via the epithelial release of GM-CSF and IL-33. J Exp Med 2012; 209: 1505-1517.
- Suwara MI, Green NJ, Borthwick LA, et al. IL-1alpha released from damaged epithelial cells is sufficient and 20
- essential to trigger inflammatory responses in human lung fibroblasts. *Mucosal Immunol* 2014; 7: 684–693. Verleden SE, Vasilescu DM, Willems S, *et al.* The site and nature of airway obstruction after lung transplantation. Am J Respir Crit Care Med 2014; 189: 292-300.
- Bae JY, Kim EK, Yang DH, et al. Reciprocal interaction between carcinoma-associated fibroblasts and squamous carcinoma cells through interleukin-1alpha induces cancer progression. Neoplasia 2014; 16: 928-938
- Richter A, Puddicombe SM, Lordan JL, et al. The contribution of interleukin (IL)-4 and IL-13 to the 23 epithelial-mesenchymal trophic unit in asthma. Am J Respir Cell Mol Biol 2001; 25: 385-391.
- Boxall C, Holgate ST, Davies DE. The contribution of transforming growth factor-beta and epidermal growth factor signalling to airway remodelling in chronic asthma. Eur Respir J 2006; 27: 208-229.
- Epaud R, Delestrain C, Louha M, et al. Combined pulmonary fibrosis and emphysema syndrome associated with ABCA3 mutations. Eur Respir J 2014; 43: 638-641.
- Rogliani P, Calzetta L, Ora J, et al. Canakinumab for the treatment of chronic obstructive pulmonary disease. Pulm Pharmacol Ther 2015; 31: 15-27.

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