

Clinical and inflammatory characteristics of the European U-BIOPRED adult severe asthma cohort



Dominick E. Shaw^{1,44}, Ana R. Sousa^{2,44}, Stephen J. Fowler³, Louise J. Fleming⁴, Graham Roberts^{5,6,7}, Julie Corfield^{8,9}, Ioannis Pandis¹⁰, Aruna T. Bansal¹¹, Elisabeth H. Bel¹², Charles Auffray¹³, Chris H. Compton², Hans Bisgaard¹⁴, Enrica Bucchioni¹⁵, Massimo Caruso¹⁶, Pascal Chanez¹⁷, Barbro Dahlén¹⁸, Sven-Erik Dahlen¹⁹, Kerry Dyson²⁰, Urs Frey²¹, Thomas Geiser²², Maria Gerhardsson de Verdier⁸, David Gibeon⁴, Yi-ke Guo¹⁰, Simone Hashimoto¹², Gunilla Hedlin²³, Elizabeth Jeyasingham²⁴, Pieter-Paul W. Hekking¹², Tim Higenbottam²⁵, Ildikó Horváth²⁶, Alan J. Knox¹, Norbert Krug²⁷, Veit J. Erpenbeck²⁸, Lars X. Larsson⁸, Nikos Lazarinis¹⁸, John G. Matthews²⁹, Roelinde Middelveld²⁰, Paolo Montuschi³⁰, Jacek Musial³¹, David Myles³², Laurie Pahus³³, Thomas Sandström³⁴, Wolfgang Seibold³⁵, Florian Singer³⁶, Karin Strandberg¹⁸, Jorgen Vestbo³⁷, Nadja Vissing³⁸, Christophe von Garnier^{22,39}, Ian M. Adcock^{4,40}, Scott Wagers⁴¹, Anthony Rowe⁴², Peter Howarth⁴³, Ariane H. Wagener¹², Ratko Djukanovic⁴³, Peter J. Sterk^{12,45} and Kian Fan Chung^{4,40,45} on behalf of the U-BIOPRED Study Group⁴⁶

Affiliations: ¹Respiratory Research Unit, University of Nottingham, Nottingham, UK. ²Respiratory Therapeutic Unit, GSK, Stockley Park, UK. ³Respiratory and Altergy Research Group, University of Nothampton, UK. \$\text{Notality Park, UK. }\text{Notality Park }\text{Notality Park, UK. }\text{Notality Park,

Correspondence: Dominick Shaw, Clinical Sciences Building, Nottingham City Hospital, Edwards Lane, Nottingham, NG5 1PB, UK. E-mail: dominic.shaw@nottingham.ac.uk

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ABSTRACT U-BIOPRED is a European Union consortium of 20 academic institutions, 11 pharmaceutical companies and six patient organisations with the objective of improving the understanding of asthma disease mechanisms using a systems biology approach.

This cross-sectional assessment of adults with severe asthma, mild/moderate asthma and healthy controls from 11 European countries consisted of analyses of patient-reported outcomes, lung function, blood and airway inflammatory measurements.

Patients with severe asthma (nonsmokers, n=311; smokers/ex-smokers, n=110) had more symptoms and exacerbations compared to patients with mild/moderate disease (n=88) (2.5 exacerbations *versus* 0.4 in the preceding 12 months; p<0.001), with worse quality of life, and higher levels of anxiety and depression. They also had a higher incidence of nasal polyps and gastro-oesophageal reflux with lower lung function. Sputum eosinophil count was higher in severe asthma compared to mild/moderate asthma (median count 2.99% *versus* 1.05%; p=0.004) despite treatment with higher doses of inhaled and/or oral corticosteroids.

Consistent with other severe asthma cohorts, U-BIOPRED is characterised by poor symptom control, increased comorbidity and airway inflammation, despite high levels of treatment. It is well suited to identify asthma phenotypes using the array of "omic" datasets that are at the core of this systems medicine approach.



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Severe asthma results in more airway inflammation, worse symptoms and lower lung function, despite increased therapy http://ow.ly/QznR3

Introduction

A substantial number of patients with asthma require systemic corticosteroids to control symptoms and/or suffer from poor control and frequent severe exacerbations despite currently available treatment [1, 2]. Although recently developed biologic compounds targeting cytokines of the type 2 pathways show promise [3, 4], identification of new treatment targets and the selection of patients best suited to respond to individual biologics is still hampered by a poor understanding of the physiological, pathological and molecular heterogeneity of severe asthma [5, 6].

Severe asthma is a collection of disease entities with varying pathophysiological characteristics [7] that result in symptoms of cough, wheeze and breathlessness, with frequent exacerbations. To address the problem of phenotypic difference and heterogeneity, the Unbiased Biomarkers for the Prediction of Respiratory Disease Outcomes (U-BIOPRED) project was set up in 2009 as a public–private partnership within the framework of the Innovative Medicines Initiative, engaging academia, the pharmaceutical industry and patient groups. The aim of U-BIOPRED is to identify multi-dimensional phenotypes of severe asthma and new treatment targets using a combination of state of the art "omics" (transcriptomic, proteomic, lipidomic and metabolomic) technologies applying a systems biology approach [8], thereby driving unbiased discovery in both adult and paediatric severe asthma [9]. This novel approach is designed to make drug development more effective and efficient.

We present the baseline characteristics of the adult participants with severe asthma who form the majority of the U-BIOPRED cohort and compare these participants with those suffering from mild/moderate disease, in terms of their clinical, symptomatic, functional and biomarker features. In a parallel paper the characteristics of the paediatric cohort are reported. These first publications of U-BIOPRED will serve as the reference documents for all subsequent publications using the "omics" technologies that are at the core of this programme.

Methods

Participants

This was a multicentre prospective cohort study recruiting from 16 clinical centres in 11 European countries. Details of the participating centres, assessments and standard operating procedures are available

This article has been amended according to the correction published in the June 2017 issue of the European Respiratory Journal.

This study is registered on ClinicalTrials.gov (identifier: NCT01982162).

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Conflict of interest: Disclosures can be found alongside the online version of this article at erj.ersjournals.com

in the online supplement. Prior to enrolment participants with severe asthma were required to have been under follow-up by a respiratory physician for at least 6 months, during which time assessments had been undertaken to optimise asthma control and medication adherence [2]. The study was approved by the ethics committee for each participating clinical institution, and adhered to the standards set by International Conference on Harmonisation and Good Clinical Practice. It is registered on ClinicalTrials. gov (identifier: NCT01982162). All participants gave written and signed informed consent.

Adult groups

The definition of severe asthma used in this study was agreed at a U-BIOPRED consensus meeting [2]. Participants with asthma had either airflow reversibility (increase in forced expiratory volume in 1 s (FEV1) >12% predicted or 200 mL following inhalation of 400 μ g salbutamol), airway hyperresponsiveness (methacholine provocative concentration causing a 20% fall in FEV1 <8 mg·mL⁻¹, or diurnal peak expiratory flow amplitude >8% of mean), or a decrease in FEV1 of 12% predicted or 200 mL within 4 weeks after tapering maintenance treatment. Four groups were recruited, as follows.

Group A: severe nonsmoking asthma (SAn)

Participants in this group were nonsmokers for at least the past 12 months, with a less than 5 pack-year smoking history, with asthma and uncontrolled symptoms defined according to Global Initiative for Asthma (GINA) guidelines [10] and/or frequent exacerbations (more than two per year) despite high-dose inhaled corticosteroids (\geqslant 1000 µg fluticasone propionate per day or equivalent dose).

Group B: smokers and ex-smokers with severe asthma (SAs/ex)

This group was defined as for the SAn group except that they were either current smokers or ex-smokers with a smoking history of at least 5 pack-years.

Group C: mild/moderate nonsmoking asthmatics (MMA)

Participants in this group were nonsmokers for at least the past 12 months, with a less than 5 pack-year smoking history and had controlled or partially controlled asthma symptoms, as defined by GINA, whilst receiving a dose of $<500 \,\mu g$ fluticasone propionate/day or equivalent.

Group D: healthy nonsmoking controls (HC)

These participants had no history of asthma or wheeze, had no other chronic respiratory disease, were nonsmokers for at least the past 12 months with a smoking history of ≤ 5 pack years and their pre-bronchodilator FEV1 was $\geq 80\%$ pred.

Protocol and assessments

Participants attended a screening visit to assess eligibility for the study (figure 1). They underwent a baseline visit (up to 28 days later) and were invited to attend for an optional bronchoscopy, high-resolution lung computed tomography and telemonitoring sessions. Spirometry, haematological profiles, and fraction of exhaled nitric oxide level ($F_{\rm eNO}$) at 50 mL·s⁻¹ were performed. Induced sputum was obtained [11] and differential sputum eosinophil and neutrophil counts measured following a standardised operating procedure. Sputum supernatants and cell pellets were collected. Allergic status was obtained by either skin prick testing or measurement of specific immunoglobulin (Ig)E to six common aeroallergens. Blood and urine samples were taken for lipidomic, proteomic and transcriptomic analyses for later assessment. An optional sample was taken for genetic analysis. Subsets of participants underwent plethysmographic measurements, high-resolution computed tomography and collection of exhaled breath for measurement of metabolites including volatile organic compounds, all for future analyses. All investigations were performed according to standardised operating procedures (online supplement).

Participants with severe asthma were reviewed at 12–18 months after enrolment and were also invited to attend if they experienced an exacerbation. At 12–24 months they were contacted by phone or by post to obtain information on asthma control.

Data were entered on an electronic case report form. The study was run and monitored by Cromsource (www.cromsource.com). Samples were sent to the Centre for Integrated Genomic Medical Research Biobank in Manchester, UK. Datasets were uploaded on to the tranSMART system, an open-source knowledge management platform for sharing research data [12] supported by the European Translational Information and Knowledge Management Services (eTRIKS) project.

The study aims are published on the U-BIOPRED home page (www.europeanlung.org/en/projects-and-research/projects/u-biopred/home).

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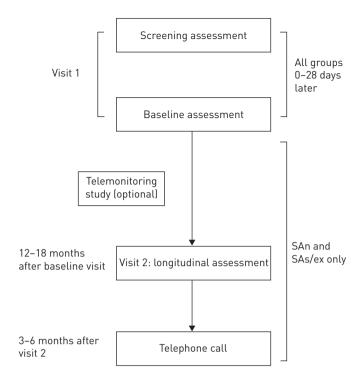


FIGURE 1 Visit schedule. SAn: severe nonsmoking asthma group; SAs/ex: smokers and ex-smokers with severe asthma.

Questionnaires

The following were administered at baseline: 1) the Asthma Control Questionnaire (ACQ5) [13] to assess current asthma control; 2) the Asthma Quality of Life Questionnaire (AQLQ) [14] to assess quality of life and psychological morbidity; 3) the Hospital Anxiety and Depression Scale (HADS) [15]; 4) Sino-Nasal Outcomes Test (SNOT20) [16] to measure upper airway symptoms; 5) the Epworth Sleepiness Scale (ESS) [17] to measure sleep and daytime drowsiness; and 6) the Medication Adherence Report Scale (MARS) [18] to measure adherence.

Statistical analysis

Continuously distributed data were either summarised using mean±sE if symmetrical, or median (interquartile range) values. Nonsymmetrical variables all exhibited positive skew and were log-transformed prior to association testing. Missing data were not imputed. p-values were calculated using a general linear model for continuous variables or a general logistic model for categorical variables. No adjustment for multiple testing was applied as the analyses were considered exploratory. Analyses were performed using R version 2.15.2 (R Core Team, 2012; www.r-project.org).

Results

A total of 610 adults were recruited over an 18-month period: 311, 110, 88 and 101 into the SAn, SAs/ex, MMA and HC groups, respectively (table 1 and figure 2).

There were more females in the SAn group (66%) compared to the other asthma groups (50%), with the age of onset of asthma 18 years later in the SAs/ex compared with SAn group. Participants with severe asthma had a higher body mass index (BMI) than those in the MMA and HC groups and were older (table 1). Both severe asthma groups experienced 2.5 exacerbations in the preceding 12 months as compared with 0.4 in the MMA group (p<0.001). There was a higher rate of intensive care unit admissions in the SAn participants compared to the SAs/ex group (p<0.05). Further split of the severe asthma groups based on current and ex-smoking is presented in the online supplement (table S5).

Spirometry

FEV1 (% predicted or actual) was lower in the three asthma groups compared to the HC group (p<0.001), with the severe asthma groups having the lowest FEV1. Forced vital capacity (FVC) (% predicted or actual) was also lower in both the SAn and SAs/ex groups when compared to the MMA (p<0.001) and HC groups (p<0.001). The mean FEV1/FVC ratio was lower in those with severe asthma (0.64 and 0.61, respectively) compared to the MMA (ratio 0.72; p<0.001) and HC groups (ratio 0.79; p<0.001), respectively (table 1).

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	Severe nonsmoking asthma	Smokers and ex-smokers with severe asthma #	Mild/moderate nonsmoking asthma	Healthy nonsmoking controls	p-value
Subjects n	311	110	88	101	
Age years	51.01±0.8 (n=311)	54.51±1.08 (n=110)	41.66±1.65 (n=88)	38.85±1.34 (n=101)	< 0.001
Age at diagnosis years	20 (7-38) (n=302)	38 (20-48) (n=109)	14 (6-32) (n=83)	NA	< 0.001
Females	205/311 (66%)	56/110 (51%)	44/88 (50%)	39/101 (39%)	< 0.001
BMI kg⋅m ⁻²	29.11±0.36 (n=311)	29.59±0.6 (n=110)	25.73±0.47 (n=88)	25.31±0.36 (n=101)	< 0.001
BMI >30 kg·m ⁻²	120/311 (38.6%)	44/110 (40%)	16/88 (18.18%)	12/101 (11.88%)	< 0.001
Serum IgE IU·mL ⁻¹	119.5 (45-342) (n=302)	126 (63-328) (n=104)	89.4 (49-244) (n=85)	23.45 (9-65) (n=98)	< 0.001
FEV ₁ % pred	67.5±1.26 (n=308)	67.2±1.84 (n=110)	89.5±1.86 (n=87)	101.76±1.29 (n=101)	< 0.001
FVC % pred	87.2±1.12 (n=308)	89.7±1.74 (n=110)	104.5±2.02 (n=87)	107.8±1.3 (n=101)	< 0.001
FEV ₁ /FVC ratio	0.64±0.01 (n=308)	0.61±0.01 (n=110)	0.72±0.01 (n=87)	0.79±0.01 (n=101)	< 0.001
Exacerbations in previous year n	2.48±0.13 (n=310)	2.55±0.26 (n=110)	0.38±0.08 (n=88)	NA	<0.001
Smoking history pack-years	2 (1-4) (n=47)	17.38 (10-26) (n=110)	4 (1-4) (n=13)	0.9 (0-3) (n=20)	<0.001
Intubation (ever)	35/307 (11%)	6/109 (6%)	0/87 (0%)	NA	0.083
ICU admission (ever)	80/307 (26%)	18/109 (17%)	1/86 (1%)	NA	< 0.001
Atopy test positive	213/272 (78.3%)	62/87 (71.3%)	72/78 (92.3%)	36/78 (46.2%)	<0.001

Data are presented as mean±sE, median (interquartile range) or n/N (%), unless otherwise stated. BMI: body mass index; FEV1: forced expiratory volume in 1 s; FVC: forced vital capacity; ICU: intensive care unit; NA: not applicable. #: 42 current smokers and 68 ex-smokers.

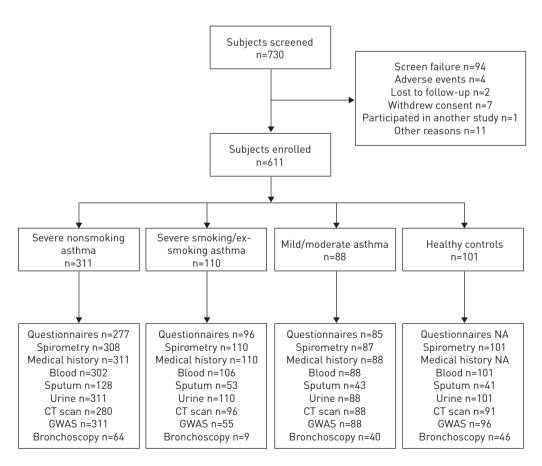


FIGURE 2 Consort diagram. CT: computed tomography; GWAS: genome-wide association study; NA: not applicable.

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Medications

Within the SAn and SAs/ex groups 46% and 45% respectively received daily oral corticosteroids, and 17% and 16%, respectively, received anti-IgE therapy. Use of nebulised β -agonist was higher in the SAn and SAs/ex groups. Other classes of therapy were also used (table 2).

Questionnaires

ACQ and AQLQ scores reflected worse asthma control and increased morbidity in both severe asthma groups with minimal impairment in the MMA group. A similar pattern was seen with anxiety and depression. There were more upper airway symptoms measured using the SNOT20 in both severe asthma groups compared with the MMA group. Similarly the ESS scores indicated that there was an increase in sleepiness in the severe asthma groups compared to only a very mild impairment in the MMA group (table 3).

The MARS questionnaire scores for adherence to treatment recorded by the three asthma groups were in the range of 21 to 22, with the severe asthma groups recording higher scores (p<0.005), indicating better adherence. The AQLQ score was correlated to several variables, including FEV1 (95% CI 0.5 to 0.7; p<0.001), FEV1/FVC (95% CI 1.14 to 2.8; p<0.001), exacerbations in the previous year (95% CI -0.8 to -0.2; p<0.001), BMI (95% CI -0.006 to -0.002; p<0.001) and pack-years smoked (95% CI -0.003 to -0.001; p<0.001) (figure 3).

Atopy and comorbidities

There was a high incidence of atopy in the four groups, at 70% in the asthma groups and 46% in the HC group. The incidence of allergic rhinitis, hay fever and nonallergic rhinitis were highest in the asthma groups. Participants in the HC group were much less allergic with only a third reporting hay fever and only a sixth, rhinitis or eczema.

The presence of nasal polyps was associated with severe asthma, regardless of smoking status (four-fold increased incidence in SAn and SAs/ex groups *versus* MMA group; p<0.001) (table 4). No such association was seen with allergic or nonallergic rhinitis, hay fever or reported eczema. Gastro-oesophageal reflux disease was more common in severe asthma (46% SAn, 63% SAs/ex) than in MMA (21%) and HC (11%), with a greater incidence reported in the SAs/ex group *versus* the SAns group (p=0.004).

Blood and sputum biomarkers

Blood eosinophil counts were similar in all three asthma groups. Each group had a significantly higher blood eosinophil count than the HC group (SAn *versus* HC, p=0.002; SAs/ex *versus* HC, p=0.005; MMA

TABLE 2 Medications			
	Severe nonsmoking asthma	Smokers and ex-smokers with severe asthma	Mild/moderate nonsmoking asthma
Subjects n	311	110	88
Oral corticosteroid	135/295 (45.8%)	46/103 (44.7%)	0/88 (0%)
Prednisolone (equ.) mg#	13.2±0.85 (n=122)	14.8±1.81 (n=36)	NA (NA)
Inhaled corticosteroids	310/311 (99.7%)	110/110 (100%)	87/88 (98.9%)
Long-acting β-agonist	305/309(98.7%)	109/110 (99.1%)	2/88 (2.3%)
Short-acting β-agonist	260/301 (86.3%)	82/105 (78.1%)	68/88 (77.3%)
Injected corticosteroids	19/284 (6.7%)	1/97 (1.0%)	0/88 (0%)
Mucolytic	31/286 (10.8%)	18/100 (18.0%)	0/88 (0%)
Anti-histamine	75/311 (24.1%)	16/110 (14.6%)	4/88 (4.5%)
Antibiotic (excluding macrolide)	11/288 (3.8%)	4/98 (4.1%)	0/88 (0%)
Macrolide	32/311 (10.3%)	13/110 (11.8%)	0/88 (0%)
Long-acting muscarinic antagonist	65/284 (22.9%)	27/97 (27.9%)	0/88 (0%)
Short-acting muscarinic antagonist	127/292 (43.5%)	48/104 (46.2%)	0/88 (0%)
Omalizumab	50/287 (17.4%)	16/98 (16.3%)	0/88 (0%)
Immunosuppressant	9/311 (2.9%)	4/110 (3.6%)	0/88 (0%)
Leukotriene modifier	139/298 (46.6%)	45/106 (42.5%)	0/88 (0%)
Cromones	10/284 (3.5%)	2/97 (2.1%)	0/88 (0%)
Anti-fungal agent	5/311 (1.6%)	1/110 (1.0%)	0/88 (0%)
Xanthine	59/289 (20.4%)	21/100 (21.0%)	0/88 (0%)
Nebulised β-agonist	82/284 (28.9%)	24/97 (24.7%)	2/88 (2.3%)

Data are presented as n/N (%) or mean±SE, unless otherwise stated. NA: not applicable. #: hydrocortisone and triamcinolone doses were converted to equivalent prednisolone dose (four healthy control participants took as required antihistamines).

TABLE 3 Questionnaires

	Severe nonsmoking asthma	Smokers and ex-smokers with severe asthma	Mild/moderate nonsmoking asthma	Unadjusted p-value#
Subjects n	311	110	88	
ACQ	2 20 . 0 07 (~ 277)	2 22 : 0 12 (= 0/)	0.0/+0.07 (= 05)	.0.001
Mean ACQ5	2.28±0.07 (n=277)	2.23±0.12 (n=96)	0.86±0.07 (n=85)	<0.001
Mean ACQ7	2.67±0.08 (n=277)	2.62±0.12 (n=96)	1.01±0.07 (n=85)	<0.001
AQLQ				
Total	4.48±0.07 (n=276)	4.44±0.13 (n=92)	5.84±0.1 (n=84)	< 0.001
Symptoms	4.46±0.08 (n=276)	4.36±0.14 (n=92)	5.87±0.1 (n=84)	< 0.001
Émotional	4.63±0.1 (n=276)	4.52±0.16 (n=92)	5.98±0.13 (n=84)	< 0.001
Environmental stimuli	4.69±0.09 (n=276)	4.57±0.16 (n=92)	5.63±0.14 (n=84)	< 0.001
Activity limitation	4.35±0.07 (n=276)	4.45±0.13 (n=92)	5.81±0.11 (n=84)	< 0.001
HADS				
Total	12.33±0.54 (n=223)	13.64±1.01 (n=72)	7.01±0.7 (n=70)	< 0.001
Anxiety	6.94±0.3 (n=223)	7.71±0.54 (n=72)	4.24±0.41 (n=70)	< 0.001
Depression	5.39±0.28 (n=223)	5.93±0.56 (n=72)	2.77±0.39 (n=70)	< 0.001
SNOT20 (total)	31.76±1.01 (n=283)	32.12±1.92 (n=97)	15.42±1.42 (n=83)	< 0.001
ESS (total)	7.48±0.26 (n=277)	7.95±0.47 (n=95)	5.49±0.41 (n=85)	< 0.001
MARS (total)	22.44±0.14 (n=278)	22.17±0.29 (n=94)	21.35±0.4 (n=84)	0.002

Data are presented as mean±sE, unless otherwise stated. ACQ: Asthma Control Questionnaire; AQLQ: Asthma Quality of Life Questionnaire; HADS: Hospital Anxiety and Depression Score; SNOT20: Sino-nasal Outcome Test 20; ESS: Epworth Sleepiness Scale; MARS: Medication Adherence Report Scale. **: severe asthma (nonsmoking asthma group and smokers and ex-smokers with severe asthma group combined) versus mild/moderate nonsmoking asthma group.

versus HC, p<0.001). Blood neutrophil counts were significantly higher in the severe asthma groups compared to the MMA group.

Sputum samples were provided and met criteria for analysis in 44.2% of the asthma participants and 40.6% of the HC group. Median sputum eosinophil counts for the SAn, SAs/ex, MMA, and HC groups were 2.75%, 4.13%, 1.05% and 0%, respectively (table 5). The sputum eosinophil count was higher in the two severe asthma groups combined compared to the mild/moderate asthma group (table 5; figure 4).

There were no significant differences in differential sputum neutrophil counts between the two severe asthma groups, which when combined were significantly higher compared to the MMA group (table 5).

There was a significant negative association between log sputum eosinophils (absolute or %) and FEV1 (% predicted or actual value) when all cohorts were considered and an adjustment for age, sex and smoking was applied. There were significant negative associations between log blood eosinophils (%) and FEV1/FVC ratio (p=0.002) and between blood neutrophils (%) and actual FEV1 (p=0.002) and FEV1/FVC ratio (p=0.026).

Exhaled nitric oxide

 $F_{\rm eNO}$ levels in all asthma groups were higher than those in the HC group, but the $F_{\rm eNO}$ levels in the severe asthma groups were not different from the levels in the MMA group. The presence of nasal polyps was associated with a higher $F_{\rm eNO}$ (mean increase 2.1 ppb, 95% CI 1.5 to 2.9 ppb; p<0.001) (table 5).

Discussion

In this large European cohort, patients with severe asthma experienced more symptoms, more exacerbations, higher levels of anxiety and depression, and a higher incidence of nasal polyps, gastro-oesophageal reflux symptoms and airflow obstruction than patients with milder disease. The clinical characteristics of asthma were present despite higher doses of treatment that included doses of inhaled corticosteroids equal or more than 1000 μ g of fluticasone (or equivalent), with 45% of the combined severe asthma group receiving a daily dose of prednisolone. The characteristic features of the severe asthma U-BIOPRED cohort are similar to those reported in previous cohort studies [6, 19–21]. While the entry criteria for severe asthma were comparable for most of these cohort studies, the ENFUMOSA study required a lower threshold with an inhaled corticosteroid dose of \geqslant 1200 μ g of budesonide or beclomethasone with at least one exacerbation in the past year. Of these five cohorts, the current U-BIOPRED severe asthma cohort appears to be the most severe with a higher reported exacerbation rate of 2.5 per year, a reduced mean FEV1 of 67.5% of predicted and a higher proportion of patients on oral corticosteroid therapy taking a mean dose of 14 mg per day.

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One of the novel features of the U-BIOPRED cohort is the inclusion of a smoking and ex-smoking severe asthma group. Patients with asthma who smoke have been reported to have poorer disease control and a reduced therapeutic response to inhaled corticosteroids [22], possibly through the induction of corticosteroid insensitivity [23, 24]. However, our analyses of the nonsmoking and the smoking/ex-smoking severe asthma groups identified few differences in demographics, airway physiology, inflammatory markers and asthma symptoms between these groups. In both groups, a similar percentage received oral corticosteroid therapy; they also had similar degrees of airflow obstruction. The slightly lower level of Feno in the smoking/ex-smoking group might be explained by an effect of current smoking [25]. One notable difference is that asthma onset occurred on average 18 years later in the smokers and ex-smokers than in the nonsmokers, and

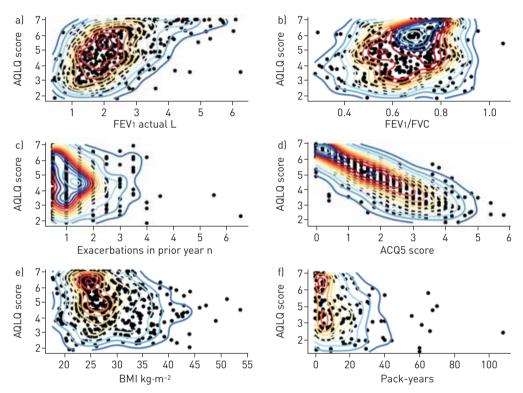


FIGURE 3 Contour plots of the Asthma Quality of Life Questionnaire (AQLQ) related to baseline demographics. Figures represent scatter plots describing the relationship between each factor and the asthma quality of life z-score. The contour lines are coloured blue to red, to indicate increasing density of points in the graph. The density was modelled using two-dimensional kernel density estimation. The contour plots show weak inverse relationships and particularly the scatter between quality of life and c) exacerbations, e) body mass index (BMI) and f) smoking history (pack-years), a strong inverse relationship between quality of life and d) asthma control (ACQ5) and weak positive relationships between quality of life and a, b) measures of lung function (forced expiratory volume in 1 s (FEV1) and FEV1 to forced vital capacity (FVC) ratio).

TABLE 4 Comorbidities							
	Severe nonsmoking asthma	Smokers and ex-smokers with severe asthma	Mild/moderate nonsmoking asthma	Healthy nonsmoking controls	p-value	p-value#	
Subjects n	311	110	88	101			
Allergic rhinitis diagnosed	164/277 (59.2%)	44/101 (43.6%)	42/70 (60%)	5/30 (16.7%)	< 0.001	0.442	
Hayfever diagnosed	135/284 (47.5%)	51/100 (51%)	46/73 (63.0%)	10/33 (30.3%)	0.019	0.024	
Nonallergic rhinitis diagnosed	42/284 (14.8%)	17/101 (16.8%)	8/72 (11.1%)	1/34 (2.9%)	0.090	0.356	
Nasal polyps diagnosed	103/291 (35.4%)	34/101 (33.7%)	7/76 (9.2%)	3/34 (8.8%)	< 0.001	< 0.001	
Eczema diagnosed	107/294 (36.4%)	31/101 (30.7%)	25/75 (33.3%)	5/35 (14.3%)	0.013	0.789	
GORD diagnosed	135/289 (46.7%)	63/99 (63.6%)	16/75 (21.3%)	4/35 (11.4%)	<0.001	<0.001	

GORD: gastro-oesophageal reflux disease. #: severe asthma (nonsmoking asthma group and smokers and ex-smokers with severe asthma group combined) versus mild/moderate nonsmoking asthma group.

TABLE 5 Biomarkers in blood, sputum and exhaled air

	Severe nonsmoking asthma	Smokers and ex-smokers with severe asthma	Mild/moderate nonsmoking asthma	Healthy nonsmoking controls	p-value	p-value#
Subjects n	311	110	88	101		
Exhaled NO ppb	26.5 (16-47) (n=290)	23.5 (12-42) (n=104)	25 (18-54) (n=87)	19.25 (13-29) (n=96)	< 0.001	0.438
Sputum						
Sputum eosinophils %	2.75 (0-19) (n=128)	4.13 (1-14) (n=53)	1.05 (0-3) (n=43)	0 (0-0) (n=41)	< 0.001	0.004
Sputum neutrophils %	53.69 (34-75) (n=128)	55.15 (35-65) (n=53)	44.5 (26-62) (n=43)	39.56 (21-56) (n=41)	0.002	0.042
Sputum differential eosinophil count >1.9%	74 (57.81%) (n=128)	32 (60.38%) (n=53)	17 (39.53%) (n=43)	1 (2.44%) (n=41)	<0.001	0.026
Blood						
Blood eosinophils %	2.94 (1-6) (n=302)	2.88 (1-5) (n=106)	3.00 (2-5) (n=88)	2.10 (1-3) (n=101)	0.001	0.295
Blood eosinophils absolute	0.2 (0.3) (n=302)	0.22 (0.29) (n=106)	0.23 (0.2) (n=88)	0.1 (0.11) (n=101)	0.001	0.295
Blood neutrophils %	62 (55-70) (n=302)	61.75 (55-69) (n=106)	56.83 (52-63) (n=88)	57.34 (51-64) (n=101)	< 0.001	< 0.001
Blood neutrophils absolute	4.73 (3.1) (n=302)	4.97 (2.87) (n=106)	3.64 (1.75) (n=88)	3.03 (1.6) (n=101)	<0.001	<0.001

Data are presented as median (interquartile range), unless otherwise stated. #: severe asthma (nonsmoking asthma group and smokers and ex-smokers with severe asthma group combined) versus mild/moderate nonsmoking asthma group.

yet the degree of airflow obstruction measured was similar. One interpretation is that there may be a more rapid rate of loss of lung function in the patients with asthma who smoke. The significant correlation between AQLQ scores and the number of pack-years of smoking exposure would also support a contribution of cigarette smoke to impaired quality of life in this group. We also split the demographic data of the groups by smoking status rather than severity (table S5 of the online supplement). This revealed that current smokers had a lower BMI compared with ex- and never-smokers.

In agreement with the SARP study [20], patients with severe asthma (especially smokers) were less frequently atopic than those with mild/moderate disease. There was also a clear association of both nasal polyps and gastro-oesophageal reflux disease with disease severity, with approximately one-third and one-half reporting polyps and reflux respectively, a finding that is in keeping with previous reports [5]. Nasal polyps are commonly found in severe asthma, and are associated with a particularly severe phenotype. There is evidence that treating nasal polyps with anti-IgE therapy results in better asthma outcomes [26]; however, whether this is due to an effect on the underlying asthma or the polyps is unknown. The link with higher F_{eNO} levels is in keeping with work showing that nasal polypectomy leads to a fall in F_{eNO} [25].

Our findings are also similar to other studies published from severe asthma registries. In agreement with both the British Thoracic Society's [27] and Belgium's [28] severe asthma registries our patients are predominantly female, with a high BMI and evidence of fixed airflow obstruction. Moreover there are similarly high levels of reflux, nasal polyps and exacerbations despite greater levels of medication.

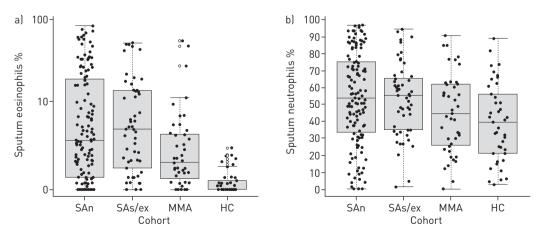


FIGURE 4 a) Sputum eosinophil count and b) sputum neutrophil count, by cohort. The boxes represent median and interquartile range values; whiskers extend to the extreme point, no more than 1.5 times the interquartile range from the box, with outliers denoted by open circles and raw data by black circles overlaid. SAn: severe nonsmoking asthma; SAs/ex: smokers and ex-smokers with severe asthma; MMA: mild/moderate nonsmoking asthma; HC: healthy nonsmoking controls.

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We found a greater degree of sputum eosinophilia in the two severe asthma groups compared to the mild/moderate asthma group. Up to 60% of patients in the two severe asthma groups had a differential sputum eosinophil of >1.9% (the established upper limit of normal for differential sputum eosinophil counts [29]). This percentage is similar to previous reports in severe asthma [21]. The level of sputum eosinophilia observed in the mild/moderate asthma group are also similar to those reported previously [30].

The higher blood neutrophil count in participants with severe asthma may represent the effect of systemic corticosteroids which can increase blood neutrophil numbers. Sputum neutrophil counts were similar in the three asthma groups and were significantly higher than in the healthy control group. This similarly could represent the effect of corticosteroids although severe asthma has been linked to a higher level of sputum neutrophils [31, 32].

The impact and burden on our participants' health was noticeable with measures of symptoms and quality of life being far worse in severe asthma as compared to mild/moderate asthma, despite the use of higher doses and more classes of asthma treatment. Levels of anxiety and depression were also higher with severe asthma. There were significant relationships between quality of life measures and airflow obstruction, smoking history and BMI, supporting the contribution of these factors to an impairment of quality of life; however, the scatter of data reveals that these parameters are not closely related. The number of exacerbations experienced was greater than 2.5 exacerbations per participant in both severe asthma groups in the preceding year. These findings highlight the need for an integrative assessment of clinical and physiological disease markers and, additionally, biological markers of disease in the assessment of severe asthma. For example, the finding that bariatric surgery has an effect on measures of airway hyperresponsiveness [33] and is associated with a lower all-cause mortality at 5 years particularly in younger, predominantly female populations [34] may point towards the need for specific and targeted intervention in people with severe asthma and obesity.

There are several limitations to our study. First, there is no perfect way to assess treatment adherence; however, we only approached patients managed in a specialist respiratory clinic and only those who had been assessed to be adherent were eligible for the study. Furthermore, MARS scores were high, indicating good levels of self-reported adherence. Secondly, subjective or historical data were assessed by questionnaire which may be prone to recall bias. Thirdly, the success rate in obtaining adequate quality sputum for analysis was in the 42–50% range and the number of bronchoscopies was relatively lower in the SA and SAs/ex groups. Finally, due to the numerous formulations and inhaler devices used across Europe it was not possible to calculate the precise daily equivalent inhaled corticosteroid dose for each participant and therefore these data are not shown; however, high (>1000 μ g FP) or low (<500 μ g fluticasone propionate per day) dose was a study entry requirement for the severe and moderate groups, respectively.

We have been successful in recruiting a substantial cohort of patients with the most severe asthma that has similar characteristics to previously reported cohorts. This gives confidence that the U-BIOPRED consortium will define distinct phenotypes and endotypes of severe asthma. Matching these data to the "omics" information with future unsupervised analyses will help identify new treatments for patients with severe asthma who currently have limited treatment options, and will improve our understanding of this important chronic disease.

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The members of the U-BIOPRED Study Group are as follows: H. Ahmed, European Institute for Systems Biology and Medicine, University of Lyon, France; D. Allen, North West Severe Asthma Network; Pennine Acute Hospital NHS Trust; P. Badorrek, Fraunhofer İTEM; S. Ballereau, European Institute for Systems Biology and Medicine, University of Lyon, France; F. Baribaud, Janssen R&D, USA; M.K. Batuwitage, Imperial College, London, UK; A. Bedding, Roche Diagnostics GmbH, Mannheim, Germany; A.F. Behndig, Umeå University; A. Berglind, Karolinska University Hospital and Karolinska Institutet; A. Berton, Boehringer Ingelheim Pharma GmbH & Co. KG; J. Bigler, Amgen Inc; M.J. Boedigheimer, Amgen Inc; K. Bønnelykke, University of Copenhagen and Danish Pediatric Asthma Center, Gentofte Hospital, University of Copenhagen, Denmark; P. Brinkman, Academic Medical Centre, University of Amsterdam; A. Bush, Department of Paediatrics and National Heart and Lung Institute, Imperial College, London; Department of Respiratory Paediatrics, Royal Brompton Hospital, London, UK; D. Campagna, University of Catania; C. Casaulta, University Children's Hospital Bern, Switzerland; A. Chaiboonchoe, European Institute for Systems Biology and Medicine, University of Lyon, France; T. Davison, Janssen R&D, USA; B. De Meulder, European Institute for Systems Biology and Medicine, University of Lyon, France; I. Delin, Institute of Environmental Medicine, Karolinska Instituet, Stockholm, Sweden; P. Dennison, NIHR Southampton Respiratory Biomedical Research Unit and University of Southampton; P. Dodson, AstraZeneca, Mölndal, Sweden; L. El Hadjam, European Institute for Systems Biology and Medicine, University of Lyon, France; D. Erzen, Boehringer Ingelheim Pharma GmbH & Co. KG; C. Faulenbach, Fraunhofer ITEM; K. Fichtner, Boehringer Ingelheim Pharma GmbH & Co. KG; N. Fitch, BioSci Consulting, Belgium; E. Formaggio, PhD, Project manager, Verona Italy; M. Gahlemann, Boehringer Ingelheim (Schweiz) GmbH; G. Galffy, Semmelweis University, Budapest, Hungary; D. Garissi, Global Head Clinical Research Division, CROMSOURCE, Italy; T. Garret, BioSci Consulting, Belgium; J. Gent, Royal

Brompton and Harefield NHS Foundation Trust; E. Guillmant-Farry, Royal Brompton Hospital, London, UK; E. Henriksson, Karolinska Institutet; U. Hoda, Imperial College; J.M. Hohlfeld, Fraunhofer ITEM; X. Hu, Amgen Inc; A. James, Karolinska Institutet; K. Johnson, Centre for respiratory medicine and allergy, Institute of Inflammation and repair, University Hospital of South Manchester, NHS Foundation Trust, Manchester, UK; N. Jullian, European Institute for Systems Biology and Medicine, University of Lyon, France; G. Kerry, Centre for respiratory medicine and allergy, Institute of Inflammation and repair, University Hospital of South Manchester, NHS Foundation Trust, Manchester, UK; M. Klüglich, Boehringer Ingelheim Pharma GmbH & Co. KG; R. Knowles, Arachos Pharma, Stevenge, UK; J.R. Konradsen, Karolinska University Hospital and Karolinska Institutet; K. Kretsos, UCB, Slough, UK; L. Krueger, University Children's Hospital Bern, Switzerland; A-S. Lantz, Karolinska University Hospital and Karolinska Institutet; C. Larminie, GSK, London, UK; P. Latzin, University Children's Hospital Bern, 3010 Bern, Switzerland; D. Lefaudeux, European Institute for Systems Biology and Medicine, University of Lyon, France; N. Lemonnier, European Institute for Systems Biology and Medicine, University of Lyon, France; L.A. Lowe, Centre for respiratory medicine and allergy, Institute of Inflammation and repair, University Hospital of South Manchester, NHS Foundation Trust, Manchester, UK; R. Lutter, Academic Medical Centre, University of Amsterdam; A. Manta, Roche Diagnostics GmbH, Mannheim, Germany; A. Mazein, European Institute for Systems Biology and Medicine, University of Lyon, France; L. McEvoy, University Hospital, Department of Pulmonary Medicine, Bern, Switzerland; A. Menzies-Gow, Royal Brompton and Harefield NHS Foundation Trust; N. Mores, Università Cattolica del Sacro Cuore; C.S. Murray, Centre for Respiratory Medicine and Allergy, The University of Manchester, Manchester Academic Health Science Centre, University Hospital of South Manchester NHS Foundation Trust, Manchester, UK; K. Nething, Boehringer Ingelheim Pharma GmbH & Co. KG; U. Nihlén, Department of Respiratory Medicine and Allergology, Skåne University Hospital, Lund, Sweden; AstraZeneca R&D, Mölndal, Sweden; R. Niven, North West Severe Asthma Network, University Hospital South Manchester NHS Trust; B. Nordlund, Astrid Lindgren Children's Hospital, Karolinska University Hospital, Stockholm, Sweden; Department of Women's and Children's Health, Karolinska Institutet, Stockholm, Sweden; S. Nsubuga, Royal Brompton Hospital, London, UK; J. Pellet, European Institute for Systems Biology and Medicine, University of Lyon, France; C. Pison, European Institute for Systems Biology and Medicine, University of Lyon, France; G. Praticò, CROMSOURCE, Verona, Italy; M. Puig Valls, CROMSOURCE, Barcelona, Spain; K. Riemann, Boehringer Ingelheim Pharma GmbH & Co. KG; J.P. Rocha, Royal Brompton and Harefield NHS Foundation Trust; C. Rossios, Imperial College; G. Santini, Università Cattolica del Sacro Cuore; M. Saqi, European Institute for Systems Biology and Medicine, University of Lyon, France; S. Scott, North West Severe Asthma Network; Countess of Chester NHS Trust; N. Sehgal, North West Severe Asthma Network; Pennine Acute Hospital NHS Trust; A. Selby, NIHR Southampton Respiratory Biomedical Research Unit, Clinical and Experimental Sciences and Human Development and Health, Southampton, UK; P. Söderman, Astrid Lindgren Children's Hospital, Karolinska University Hospital, Stockholm, Sweden; Department of Women's and Children's Health, Stockholm, Sweden; A. Sogbesan, Royal Brompton and Harefield NHS Foundation Trust; F. Spycher, University Hospital, Department of Pulmonary Medicine, Bern, Switzerland; S. Stephan, Centre for respiratory medicine and allergy, Institute of Inflammation and repair, University Hospital of South Manchester, NHS Foundation Trust, Manchester, UK; J. Stokholm, University of Copenhagen and Danish Pediatric Asthma Center, Gentofte Hospital, University of Copenhagen, Denmark; M. Sunther, Centre for respiratory medicine and allergy, Institute of Inflammation and repair, University Hospital of South Manchester, NHS Foundation Trust, Manchester, UK; M. Szentkereszty, Semmelweis University, Budapest, Hungary; L. Tamasi, Semmelweis University, Budapest, Hungary, K. Tariq, NIHR Southampton Respiratory Biomedical Research Unit and University of Southampton; S. Valente, Università Cattolica del Sacro Cuore; W.M. van Aalderen, Academic Medical Centre, University of Amsterdam; C.M. van Drunen, Academic Medical Centre, University of Amsterdam; J. Van Eyll, UCB, Slough, UK; A. Vyas, North West Severe Asthma Network; Lancashire Teaching Hospitals NHS Trust; W. Yu, Amgen Inc; W. Zetterquist, Department of Woman and Child Health, Karolinska Institutet, Department of Woman and Child Health, Karolinska Institutet, Stockholm, Sweden; Z. Zolkipli, NIHR Southampton Respiratory Biomedical Research Unit, University Hospital Southampton NHS Foundation Trust, Southampton, UK; Clinical and Experimental Sciences and Human Development in Health Academic Unit, University of Southampton Faculty of Medicine, Southampton, UK; The David Hide Asthma and Allergy Research Centre, St Mary's Hospital, Isle of Wight, UK; A.H. Zwinderman, Academic Medical Centre, University of

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Investigators and contributors: Nora Adriaens, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands; Antonios Aliprantis, Merck Research Laboratories, Boston, USA; Kjell Alving, Dept Women's and Children's Health, Uppsala University, Uppsala, Sweden; Per Bakke, Department of Clinical Science, University of Bergen, Borgen, Norway; David Balgoma, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Clair Barber, NIHR Southampton Respiratory Biomedical Research Unit and Clinical and Experimental Sciences, Southampton, UK; Frédéric Baribaud, Janssen R&D, USA; Stewart Bates, Respiratory Therapeutic Unit, GSK, London, UK; An Bautmans, MSD, Brussels, Belgium; Jorge Beleta, Almirall S.A., Barcelona, Spain; Grazyna Bochenek, II Department of Internal Medicine, Jagiellonian University Medical College, Krakow, Poland; Joost Brandsma, University of Southampton, Southampton, UK; Armin Braun, Fraunhofer Institute for Toxicology and Experimental Medicine, Hannover, Germany; Dominic Burg, Centre for Proteomic Research, Institute for Life Sciences, University of Southampton, Southampton, UK; Leon Carayannopoulos, previously at: MSD, USA; João Pedro Carvalho da Purificação Rocha, Royal Brompton and Harefield NHS Foundation Trust, London, UK; Romanas Chaleckis, Centre of Allergy Research, Karolinska Institutet, Stockholm, Sweden; Arnaldo D'Amico, University of Rome "Tor Vergata', Rome Italy; Jorge De Alba, Almirall S.A., Barcelona, Spain; Inge De Lepeleire, MSD, Brussels, Belgium; Tamara Dekker, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands; Annemiek Dijkhuis, Academic Medical Centre, University of Amsterdam, The Netherlands; Aleksandra Draper, BioSci Consulting, Maasmechelen, Belgium; Jessica Edwards, Asthma UK, London, UK; Rosalia Emma, Department of Clinical and Experimental Medicine, University of Catania, Catania, Italy; Magnus Ericsson, Karolinska University Hospital, Stockholm, Śweden; Breda Flood, European Federation of Allergy and Airways Diseases Patient's Associations, Brussels, Belgium; Hector Gallart, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Cristina Gomez, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Kerry Gove, NIHR Southampton Respiratory Biomedical Research Unit and Clinical and Experimental Sciences, Southampton, UK; Neil Gozzard, UCB, Slough, UK; John Haughney, International Primary Care

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Respiratory Group, Aberdeen, Scotland; Lorraine Hewitt, NIHR Southampton Respiratory Biomedical Research Unit, Southampton, UK; Jens Hohlfeld, Fraunhofer Institute for Toxicology and Experimental Medicine, Hannover, Germany; Cecile Holweg, Respiratory and Allergy Diseases, Genentech, San Francisco, USA; Richard Hu, Amgen Inc. Thousand Oaks, USA; Sile Hu, National Heart and Lung Institute, Imperial College, London, UK; Juliette Kamphuis, Longfonds, Amersfoort, The Netherlands; Erika J. Kennington, Asthma UK, London, UK; Dyson Kerry, CromSource, Stirling, UK; Hugo Knobel, Philips Research Laboratories, Eindhoven, The Netherlands; Johan Kolmert, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Maxim Kots, Chiesi Pharmaceuticals, SPA, Parma, Italy; Scott Kuo, National Heart and Lung Institute, Imperial College, London, UK; Maciei Kupczyk, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Bart Lambrecht, University of Gent, Gent, Belgium; Saeeda Lone-Latif, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands; Matthew J. Loza, Janssen R&D, USA; Lisa Marouzet, NIHR Southampton Respiratory Biomedical Research Unit, Southampton, UK; Jane Martin, NIHR Southampton Respiratory Biomedical Research Unit, Southampton, UK; Sarah Masefield, European Lung Foundation, Sheffield, UK; Caroline Mathon, Centre of Allergy Research, Karolinska Institutet, Stockholm, Sweden; Sally Meah, National Heart and Lung Institute, Imperial College, London, UK; Andrea Meiser, Data Science Institute, Imperial College, London, UK; Leanne Metcalf, previously at: Asthma UK, London, UK; Maria Mikus, Science for Life Laboratory and The Royal Institute of Technology, Stockholm, Sweden; Montse Miralpeix, Almirall, Barcelona, Spain; Philip Monk, Synairgen Research Ltd, Southampton, UK; Shama Naz, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Ben Nicholas, University of Southampton, Southampton, UK; Peter Nilsson, Science for Life Laboratory and The Royal Institute of Technology, Stockholm, Sweden; Jörgen Östling, AstraZeneca, Mölndal, Sweden; Antonio Pacino, Lega Italiano Anti Fumo, Catania, Italy; Susanna Palkonen, European Federation of Allergy and Airways Diseases Patient's Associations, Brussels, Belgium; Stelios Pavlidis, National Heart and Lung Institute, Imperial College, London, UK; Giorgio Pennazza, University of Rome 'Tor Vergata', Rome Italy; Anne Petrén, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Sandy Pink, NIHR Southampton Respiratory Biomedical Research Unit, Southampton, UK; Anthony Postle, University of Southampton, UK; Pippa Powell, European Lung Foundation, Sheffield, UK; Malayka Rahman-Amin, Previously at: Asthma UK, London, UK; Navin Rao, Janssen R&D, USA; Lara Ravanetti, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands; Emma Ray, NIHR Southampton Respiratory Biomedical Research Unit, Southampton, UK; Stacey Reinke, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Leanne Reynolds, previously at: Asthma UK, London, UK; John Riley, Respiratory Therapeutic Unit, GSK, London, UK; Martine Robberechts, MSD, Brussels, Belgium; Amanda Roberts, Asthma UK, London, UK; Kirsty Russell, National Heart and Lung Institute, Imperial College, London, UK; Michael Rutgers, Longfonds, Amersfoort, The Netherlands; Marco Santoninco, University of Rome 'Tor Vergata', Rome Italy; Corinna Schoelch, Boehringer Ingelheim Pharma GmbH & Co. KG, Biberach, Germany; James P.R. Schofield, Centre for Proteomic Research, Institute for Life Sciences, University of Southampton, Southampton, UK; Marcus Sjödin, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Paul J. Skipp, Centre for Proteomic Research, Institute for Life Sciences, University of Southampton, Southampton, UK; Barbara Smids, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands; Caroline Smith, NIHR Southampton Respiratory Biomedical Research Unit, Southampton, UK; Jessica Smith, Asthma UK, London, UK; Katherine M. Smith, University of Nottingham, UK; Doroteya Staykova, University of Southampton, Southampton, UK; Kai Sun, Data Science Institute, Imperial College, London, UK; John-Olof Thörngren, Karolinska University Hospital, Stockholm, Sweden; Bob Thornton, MSD, USA; Jonathan Thorsen, COPSAC, Copenhagen Prospective Studies on Asthma in Childhood, Herlev and Gentofte Hospital, University of Copenhagen, Copenhagen, Denmark; Marianne van de Pol, Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands; Marleen van Geest, AstraZeneca, Mölndal, Sweden; Jenny Versnel, previously at: Asthma UK, London, UK; Anton Vink, Philips Research Laboratories, Eindhoven, The Netherlands; Frans Wald, Boehringer Ingelheim Pharma GmbH & Co. KG, Biberach, Germany; Samantha Walker, Asthma UK, London, UK; Jonathan Ward, Histochemistry Research Unit, Faculty of Medicine, University of Southampton, Southampton, UK; Zsoka Weiszhart, Semmelweis University, Budapest, Hungary; Kristiane Wetzel, Boehringer Ingelheim Pharma GmbH, Biberach, Germany; Craig E. Wheelock, Centre for Allergy Research, Karolinska Institutet, Stockholm, Sweden; Coen Wiegman, National Heart and Lung Institute, Imperial College, London, UK; Siân Williams, International Primary Care Respiratory Group, Aberdeen, Scotland; Susan J. Wilson, Histochemistry Research Unit, Faculty of Medicine, University of Southampton, Southampton, UK; Ashley Woodcock, Centre for Respiratory Medicine and Allergy, Institute of Inflammation and Repair, University of Manchester and University Hospital of South Manchester, Manchester Academic Health Sciences Centre, Manchester, UK; Xian Yang, Data Science Institute, Imperial College, London, UK; Elizabeth Yeyasingham, UK Clinical Operations, GSK, Stockley Park, UK.

Partner organisations: Novartis Pharma AG; University of Southampton, Southampton, UK; Academic Medical Centre, University of Amsterdam, Amsterdam, The Netherlands; Imperial College London, London, UK; University of Catania, Catania, Italy; University of Rome 'Tor Vergata', Rome, Italy; Hvidore Hospital, Hvidore, Denmark; Jagiellonian Univ. Medi.College, Krakow, Poland; University Hospital, Inselspital, Bern, Switzerland; Semmelweis University, Budapest, Hungary; University of Manchester, Manchester, UK; Université d'Aix-Marseille, Marseille, France; Fraunhofer Institute, Hannover, Germany; University Hospital, Umea, Sweden; Ghent University, Ghent, Belgium; Ctr. Nat. Recherche Scientifique, Villejuif, France; Università Cattolica del Sacro Cuore, Rome, Italy; University Hospital, Copenhagen, Denmark; Karolinska Institutet, Stockholm, Sweden; Nottingham University Hospital, Nottingham, UK; University of Bergen, Bergen, Norway; Netherlands Asthma Foundation, Leusden, NL; European Lung Foundation, Sheffield, UK; Asthma UK, London, UK; European Fed. of Allergy and Airways Diseases Patients' Associations, Brussels, Belgium; Lega Italiano Anti Fumo, Catania, Italy; International Primary Care Respiratory Group, Aberdeen, Scotland; Philips Research Laboratories, Eindhoven, NL; Synairgen Research Ltd, Southampton, UK; Aerocrine AB, Stockholm, Sweden; BioSci Consulting, Maasmechelen, Belgium; Almirall; AstraZeneca; Boehringer Ingelheim; Chiesi; GlaxoSmithKline; Roche; UCB; Janssen Biologics BV; Amgen NV; Merck Sharp & Dohme Corp.

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Members of the ethics board: Jan-Bas Prins, biomedical research, LUMC, the Netherlands; Martina Gahlemann, clinical care, BI, Germany; Luigi Visintin, legal affairs, LIAF, Italy; Hazel Evans, paediatric care, Southampton, UK; Martine Puhl, patient representation (co-chair), NAF, the Netherlands; Lina Buzermaniene, patient representation, EFA, Lithuania; Val Hudson, patient representation, Asthma UK; Laura Bond, patient representation, Asthma UK; Pim de Boer, patient representation and pathobiology, IND; Guy Widdershoven, research ethics, VUMC, the Netherlands; Ralf Sigmund, research methodology and biostatistics, BI, Germany.

The patient input platform: Amanda Roberts, UK; David Supple (chair), UK; Dominique Hamerlijnck, The Netherlands; Jenny Negus, UK; Juliëtte Kamphuis, The Netherlands; Lehanne Sergison, UK; Luigi Visintin, Italy; Pim de Boer (co-chair), The Netherlands; Susanne Onstein, The Netherlands.

Members of the safety monitoring board: William MacNee, clinical care; Renato Bernardini, clinical pharmacology; Louis Bont, paediatric care and infectious diseases; Per-Ake Wecksell, patient representation; Pim de Boer, patient representation and pathobiology (chair); Martina Gahlemann, patient safety advice and clinical care (co-chair); Ralf Sigmund, bio-informatician.

References

- 1 Chung KF, Wenzel SE, Brozek JL, et al. International ERS/ATS guidelines on definition, evaluation and treatment of severe asthma. Eur Respir J 2014; 43: 343–373.
- Bel EH, Sousa A, Fleming L, et al. Diagnosis and definition of severe refractory asthma: an international consensus statement from the Innovative Medicine Initiative (IMI). Thorax 2011; 66: 910–917.
- Pavord ID, Korn S, Howarth P, et al. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. Lancet 2012; 380: 651–659.
- Wenzel S, Ford L, Pearlman D, et al. Dupilumab in persistent asthma with elevated eosinophil levels. N Engl J Med 2013; 368: 2455–2466.
- Wu W, Bleecker E, Moore W, et al. Unsupervised phenotyping of Severe Asthma Research Program participants using expanded lung data. J Allergy Clin Immunol 2014; 133: 1280–1288.
- Haldar P, Pavord ID, Shaw DE, et al. Cluster analysis and clinical asthma phenotypes. Am J Respir Crit Care Med 2008: 178: 218–224.
- 7 Wenzel SE. Asthma phenotypes: the evolution from clinical to molecular approaches. Nat Med 2012; 18: 716–725.
- Wheelock CE, Goss VM, Balgoma D, et al. Application of 'omics technologies to biomarker discovery in inflammatory lung diseases. Eur Respir J 2013; 42: 802–825.
- 9 Auffray C, Adcock IM, Chung KF, et al. An integrative systems biology approach to understanding pulmonary diseases. Chest 2010; 137: 1410–1416.
- 10 Bousquet J. Global initiative for asthma (GINA) and its objectives. Clin Exp Allergy 2000; 30: Suppl. 1, 2-5.
- 11 Pavord ID, Pizzichini MM, Pizzichini E, et al. The use of induced sputum to investigate airway inflammation. Thorax 1997; 52: 498–501.
- 12 Szalma S, Koka V, Khasanova T, et al. Effective knowledge management in translational medicine. J Transl Med 2010; 8: 68.
- 13 Juniper EF, O'Byrne PM, Guyatt GH, et al. Development and validation of a questionnaire to measure asthma control. Eur Respir J 1999; 14: 902–907.
- Juniper EF, Guyatt GH, Ferrie PJ, et al. Measuring quality of life in asthma. Am Rev Respir Dis 1993; 147:
- 15 Zigmond AS, Snaith RP. The hospital anxiety and depression scale. Acta Psychiatr Scand 1983; 67: 361-370.
- Morley AD, Sharp HR. A review of sinonasal outcome scoring systems which is best? Clin Otolaryngol 2006; 31: 103–109.
- 17 Johns MW. A new method for measuring daytime sleepiness: the Epworth sleepiness scale. Sleep 1991; 14: 540-545.
- 18 Cohen JL, Mann DM, Wisnivesky JP, et al. Assessing the validity of self-reported medication adherence among inner-city asthmatic adults: the Medication Adherence Report Scale for Asthma. Ann Allergy Asthma Immunol 2009; 103: 325–331.
- 19 The ENFUMOSA cross-sectional European multicentre study of the clinical phenotype of chronic severe asthma. *Eur Respir J* 2003; 22: 470–477.
- 20 Moore WC, Bleecker ER, Curran-Everett D, et al. Characterization of the severe asthma phenotype by the National Heart, Lung, and Blood Institute's Severe Asthma Research Program. J Allergy Clin Immunol 2007; 119: 405–413
- 21 Schleich FN, Manise M, Sele J, et al. Distribution of sputum cellular phenotype in a large asthma cohort: predicting factors for eosinophilic vs neutrophilic inflammation. BMC Pulm Med 2013; 13: 11.
- 22 Chaudhuri R, Livingston E, McMahon AD, et al. Cigarette smoking impairs the therapeutic response to oral corticosteroids in chronic asthma. Am J Respir Crit Care Med 2003; 168: 1308–1311.
- 23 Silva GE, Sherrill DL, Guerra S, et al. Asthma as a risk factor for COPD in a longitudinal study. Chest 2004; 126: 59–65.
- 24 Bleecker ER. Similarities and differences in asthma and COPD. The Dutch hypothesis. Chest 2004; 126: Suppl., 93S-95S.
- 25 Galli J, Montuschi P, Passàli GC, et al. Exhaled nitric oxide measurement in patients affected by nasal polyposis. Otolaryngol Head Neck Surg 2012; 147: 351–356.
- 26 Gevaert P, Calus L, Van Zele T, et al. Omalizumab is effective in allergic and nonallergic patients with nasal polyps and asthma. J Allergy Clin Immunol 2013; 131: 110–116.

1320 DOI: 10.1183/13993003.00779-2015

- Heaney LG, Brightling CE, Menzies-Gow A, *et al.* Refractory asthma in the UK: cross-sectional findings from a UK multicentre registry. *Thorax* 2010; 65: 787–794.
- Schleich F, Brusselle G, Louis R, et al. Heterogeneity of phenotypes in severe asthmatics. The Belgian Severe Asthma Registry (BSAR). Respir Med 2014; 108: 1723–1732.
- 29 Reddel HK, Taylor DR, Bateman ED, et al. An official American Thoracic Society/European Respiratory Society statement: asthma control and exacerbations: standardizing endpoints for clinical asthma trials and clinical practice. Am J Respir Crit Care Med 2009; 180: 59–99.
- 30 McGrath KW, Icitovic N, Boushey HA, et al. A large subgroup of mild-to-moderate asthma is persistently noneosinophilic. Am J Respir Crit Care Med 2012; 185: 612–619.
- 31 Jatakanon A, Uasuf C, Maziak W, et al. Neutrophilic inflammation in severe persistent asthma. Am J Respir Crit Care Med 1999; 160: 1532–1539.
- Moore WC, Hastie AT, Li X, et al. Sputum neutrophil counts are associated with more severe asthma phenotypes using cluster analysis. J Allergy Clin Immunol 2014; 133: 1557–1563.
- Dixon AE, Pratley RE, Forgione PM, et al. Effects of obesity and bariatric surgery on airway hyperresponsiveness, asthma control, and inflammation. J Allergy Clin Immunol 2011; 128: 508–515.
- 34 Arterburn DE, Olsen MK, Smith VA, et al. Association between bariatric surgery and long-term survival. JAMA 2015; 313: 62–70.