



SERIES “AIRWAY REMODELLING: FROM BASIC SCIENCE TO CLINICAL PRACTICE”

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Number 4 in this Series

Clinical relevance of airway remodelling in airway diseases

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ABSTRACT: Asthma and chronic obstructive pulmonary disease (COPD) are characterised by airflow obstruction, airway remodelling (measurable structural change) and inflammation. The present review will examine the relationship between airway remodelling in these two conditions with respect to symptoms, abnormal lung function, airway hyperresponsiveness and decline in lung function. The potential for remodelling to be a protective response will also be discussed.

Asthma is associated with variable symptoms and changes in lung function and also fixed abnormalities of lung function and an increased rate of decline in lung function with age. There is a relative preservation of the relaxed airway lumen dimensions, prominent thickening of the smooth muscle layer and reduced airway distensibility. The severity of asthma is related to the degree of airway remodelling, which is most marked in cases of fatal asthma.

In COPD, symptoms are persistent and predictable but also progressive and are related to fixed abnormalities of lung function. Remodelling is associated with narrowing of the airway lumen and an increased thickness of the airway wall, although not usually to the extent seen in asthma. COPD is most often due to smoking where there is also remodelling of the parenchyma that may contribute to symptoms.

KEYWORDS: Asthma, clinical, chronic obstructive pulmonary disease, remodelling

The specific elements of remodelling which contribute to the clinical and functional manifestations of asthma and chronic obstructive pulmonary disease (COPD) remain poorly understood, due to the heterogeneity of these “diseases” and the interactions of numerous structural components within the airways. This article will examine the evidence that airway remodelling influences the clinical expression and natural evolution of airway diseases, predominantly asthma and COPD, and assess the relative protective and detrimental effects of this process. Previous articles in this series have defined remodelling and its relationship to airway function. This article will attempt to link what is known about airway remodelling to the acute and chronic clinical expression of airway disease, including symptoms, exacerbations, progression and severity of disease. This will not include the effects of remodelling in the integrated

airway tree, which is suggested by the overall reduction of airway complexity seen on casts from cases of fatal asthma [1] and by functional imaging studies demonstrating altered distribution of airflow in cases of asthma during bronchoconstriction with varying airway closure [2, 3].

Asthma is defined as “a chronic inflammatory disorder of the airways in which cells and cellular elements play a role. The chronic inflammation is associated with airway hyperresponsiveness that leads to recurrent episodes of wheezing, breathlessness, chest tightness and coughing, particularly at night or in the morning. These episodes are usually associated with widespread but variable airflow obstruction that is often reversible either spontaneously or with treatment” [4]. This less-than-perfect operative definition relates symptoms to airway hyperresponsiveness (AHR) and airway inflammation. In asthma, the pathology

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seems confined primarily to the conducting airways. Not only is airway inflammation present but there is also considerable structural remodelling of the airway wall, particularly the smooth muscle. The airway hyperresponsiveness that is characteristic of asthma is independent of baseline lung function. Symptoms and abnormalities of lung function generally (but not always) respond well to treatment with bronchodilators and inhaled or oral corticosteroids, but recur if treatment is withdrawn.

COPD is defined by the presence of persistently abnormal lung function, most often measured using forced expired volume in one second (FEV₁) [5–7]. The pathology of COPD involves both the conducting airways and the lung parenchyma, and COPD is associated with a systemic component [8]. It is characterised clinically by varying severity of cough and sputum production and by persistent, predictable breathlessness related to exertion. Symptoms and lung function deteriorate with time, especially in those who continue to smoke. Unlike in asthma, AHR in COPD is generally related to airway calibre and the disease responds poorly to treatment with bronchodilators or corticosteroids [9]. In a similar manner to asthma, exacerbations in COPD are frequently caused by viral infections [10]. COPD is most often, but not always, caused by smoking cigarettes. However, it should be remembered that some patients with asthma (who may or may not smoke cigarettes) may develop fixed or irreversible (by treatment) airflow obstruction and thus, by definition, also have COPD.

Remodelling and inflammation of the airways are characteristic of both asthma and COPD. The present article will regard remodelling as a quantifiable change in the dimensions of airway structure or in the relationship of airway structures to each other, compared with normal individuals. Remodelling in asthma and COPD and how this may be related to clinical outcomes will be separately examined, recognising that many of the clinical and pathological features of these two conditions overlap.

Although uncommon, asthma continues to cause death in some patients. Fatal asthma usually occurs on a background of chronic, severe symptoms often with fixed abnormalities of lung function, recurrent admissions to hospital, poor use of treatment or poor response to it, psychological factors and an underestimation of disease severity. A large body of qualitative and quantitative observations of airway structure in fatal asthma exists, and suggests that airway remodelling has a crucial role to play in fatal attacks. Much of the data regarding asthma pathology, especially remodelling, has been obtained from cases of fatal asthma. The extent to which these cases represent a uniquely remodelled phenotype or reflect clinically severe asthma will also be discussed, although this remains poorly understood.

Finally, it is largely assumed that airway remodelling is detrimental. There is plenty of evidence to support this view and this article will add to this impression. However, there is a growing interest in the possible protective role of remodelling which may be thought of as either adaptive remodelling that is designed to protect, yet in fact is detrimental, or remodelling that is truly protective and without which the patient would be even more severely compromised by symptoms and abnormal lung function.

REMODELLING: THE CHANGES IN ASTHMA AND COPD

Epithelium

Although desquamation of the epithelium is often reported as a pathological feature of asthma [11–15], quantification of the amount of damaged or missing epithelium in cases of mild or severe asthma has not shown any systematic significant differences from nonasthmatic control cases [16–18]. This is not surprising since damage to the epithelium in bronchial biopsies or as a *post mortem* artefact is considerable and likely to confound structural differences between cases of asthma and normal controls. What is clear is that in asthma the epithelium is thicker [17], there is hyperplasia of goblet cells (fig. 1) [19, 20] and there is increased turnover of epithelial cells, suggested by the more frequent appearance of free epithelial cells in bronchoalveolar lavage fluid [21, 22] or sputum [23], or the reduced viability of epithelial cells [24] in patients with asthma. These findings are not universal: some studies have shown no differences in the number of epithelial cells in the lavage of asthmatic patients [25] and it is apparent that significant desquamation of epithelial cells can occur in biopsy specimens from nonasthmatic subjects [26].

In COPD, epithelial thickness is also increased [27] and shows goblet cell hyperplasia [28, 29]. In addition, metaplasia of the epithelium is a more characteristic pathological change [30], compared with normal subjects or patients with asthma, although this is more an effect of cigarette smoking than the development of COPD.

The epithelium in asthma and COPD is a site of inflammation, displays altered permeability and is the source of a number of growth factors and cytokines which modulate inflammation and remodelling below the basement membrane [31]. These aspects of epithelial pathology are beyond the scope of this article. The role of epithelial remodelling in the clinical manifestations of airway disease is discussed below.

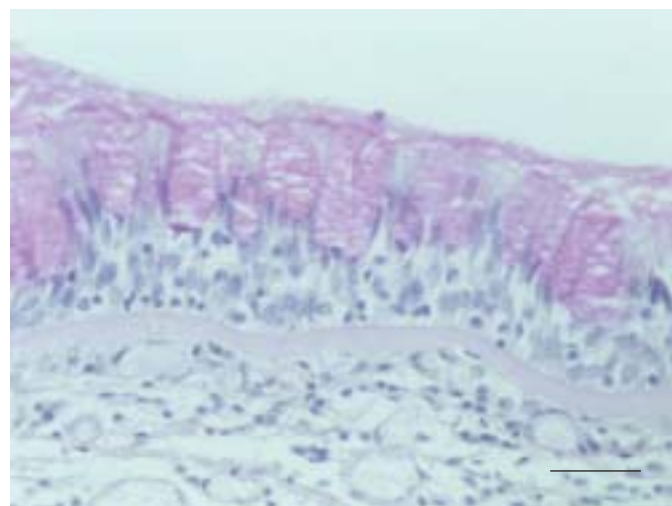


FIGURE 1. Section of airway epithelium from an asthma patient showing marked increase in goblet cell size and number and deposition of matrix material below the basement membrane. Periodic acid-Schiff stain. Scale bar=25 μ m.

Smooth muscle

The increased thickness of the smooth muscle contributes most to the increased airway wall thickness observed in asthma [32]. Increased thickness of the smooth muscle layer seen on transverse airway sections could result from more muscle cells (hyperplasia), larger muscle cells (hypertrophy) and/or more extracellular matrix (ECM) within the smooth muscle layer. A few studies have suggested that hyperplasia of smooth muscle is present in the central airways in some fatal cases [33, 34] and in clinically mild-to-moderate cases [35] of asthma. The latter study did not find any hypertrophy of smooth muscle or changes in contractile proteins that might suggest a hypercontractile smooth muscle cell phenotype. Conversely, EBINA and co-workers [33, 36] showed that in cases of fatal asthma where the smooth muscle was increased in both the central and peripheral airways, hyperplasia and hypertrophy were present in the central airways and hypertrophy was also present in the peripheral airways. These carefully conducted studies, undertaken using stereological methods, did not take into account the effects of the ECM between smooth muscle cells. This may differ systematically between cases of asthma and normal subjects, and estimation of the volume fraction of ECM may depend critically on the orientation of the smooth muscle layer that is used (fig. 2) [37]. For example, an increase in ECM in the smooth muscle layer may lead to an overestimation of smooth muscle volume and the degree of hypertrophy, since mean individual cell volume in tissues is calculated by dividing the number of cells (nuclei) into the estimated smooth muscle volume.

An increase in the volume of smooth muscle does not necessarily mean increased force development, since proliferating or secretory smooth muscle may have reduced contractility [38, 39]. A hypercontractile phenotype is observed *in vitro* after serum deprivation or other manipulations of cultured smooth muscle cells [39]. It is not clear whether the "normal" state of smooth muscle *in situ* is secretory, contractile or hypercontractile. The smooth muscle from patients with asthma has not been shown to contain more contractile proteins [35], but it is more sensitive to proliferative stimuli [40]. Theoretical studies have suggested that the smooth muscle in the thickened asthmatic airway would probably have to generate more force to narrow the thickened and

(theoretically) less compliant airway [41], although biological confirmation of these calculations does not yet exist.

Airway smooth muscle (ASM) has been assessed in a number of studies of COPD. HOGG *et al.* [42], in their landmark paper in 1968, showed with semi-quantitative pathology that smooth muscle was increased in the small airways in patients who had more severe airflow obstruction, were older and had smoked more cigarettes. Subsequent studies of smooth muscle in COPD have shown mixed results, with approximately half showing either an increase [27, 43–45] or no significant change [46–48]. In studies that also included comparisons with cases of asthma, the area of smooth muscle was less in COPD than in asthma [45, 49]. The study of TIDDENS *et al.* [48] was one of few to study large airways in patients who had been harmed by cigarette smoke and had developed airflow obstruction. However, unlike that of DUNNILL *et al.* [49], the former study found no significant differences in the amount of ASM in the large (or small) airways. In addition, it was found that, although maximum expiratory flow rates were related to overall wall area, they were not related to the area of ASM.

In summary, many studies show that there is more ASM in cases of asthma and COPD than in control cases, but there are some studies that do not agree and others that show heterogeneity between cases and between small and large airways. The contribution of the ECM to the smooth muscle layer in asthma and in COPD also remains to be determined.

ECM

Increased attention has recently been directed to the tissue between (and within) the structures of the airway wall: the ECM. This amorphous material is made up of fluids and a combination of proteoglycans and glycosaminoglycans. The ECM proteins have a number of functions that include: structural integrity; fluid balance; cellular migration; assembly and aggregation of structural proteins; elaboration of growth factors and cytokines; and osmotic activity. Changes in the relative amounts of these matrix proteins may therefore lead to changes in function. The increase in ECM between ASM cells has been suggested to cause the increased area of ASM observed in asthma (fig. 2) [37]; it may also alter airway wall compliance [50]. Studies of matrix proteins in asthma have shown increased deposition of: collagen, particularly sub-types

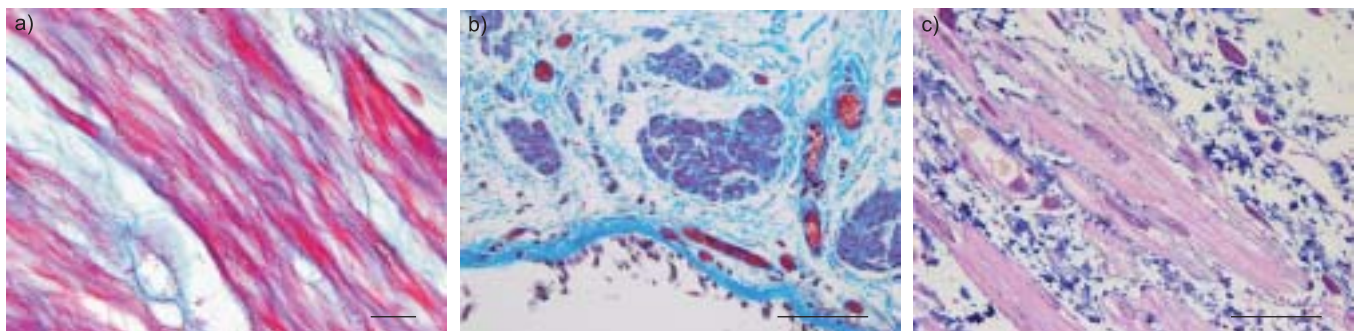


FIGURE 2. Airway smooth muscle stained with the Masson's trichrome technique and cut in: a) longitudinal section (4 μm) showing extensive overlap of the smooth muscle (red) and extracellular matrix (blue); b) transverse section showing separation of the smooth muscle and matrix; and c) thin longitudinal section (0.5 μm) also showing separation of muscle from the matrix. These images emphasise that changes in the amount and interaction of the matrix and smooth muscle in airway diseases are likely to affect airway function. Scale bars=50 μm .

I, III and IV [51–54]; decorin [55, 56]; laminin [57, 58]; tenascin [53, 59, 60]; versican [55, 56, 59]; biglycan [56, 59, 61]; perlecan [56, 61]; hyaluronan [50, 62]; and decreased lumican [55]. Reductions in elastin have been observed in a number of studies [63–65]. Increased collagen in the airway submucosa has also been observed in occupational asthma [66]. Recently, researchers have begun to examine the ECM within the bundles of ASM, although the results are only currently available in abstract form. These show that elastin [67] is increased in the smooth muscle layer in large airways in cases of fatal asthma, compared with control cases. No significant differences in total collagen content were observed [68]. PINI *et al.* [69] showed that, while levels of proteoglycans were unchanged in the subepithelial layer, they were increased in the smooth muscle layer in moderate asthma, compared with severe cases of asthma.

Fewer studies have been undertaken in COPD but a number have reported “fibrosis” in the airway wall of smokers with COPD [43, 46, 70]. Interestingly, there are no studies that have shown changes in airway wall collagen in COPD. Increased deposition of tenascin and laminin has been reported in asymptomatic smokers [71] and has been related to the numbers of mast cells in the airway wall [72]. VAN STRAATEN *et al.* [73] showed that airways from smokers with severe emphysema had diminished staining in peribronchial tissues for the proteoglycans decorin and biglycans, without changes in collagen, fibronectin or laminin, compared with nonsmokers or those with mild emphysema.

Remodelling of other structures in asthma and COPD

A number of studies have examined the vasculature of the airway wall in asthma and have shown an increase in the number [74] or area [75, 76] of the submucosal small blood vessels. These changes are thought to be due to both dilatation and congestion of existing airways but also to the formation of new vessels, possibly in response to local inflammation and the elaboration of growth factors [77]. Remodelling of large peribronchial vessels has also been observed [78], along with thickening of the arterial media in cases of fatal asthma. KUWANO *et al.* [45] found no significant differences between blood vessel dimensions in cases of COPD and control cases. The effects of the reported changes in blood vessels in asthma on clinical manifestations may be many. First, congested vessels will occupy space and may contribute to the exaggerated airway narrowing that occurs in response to smooth muscle shortening around a thickened airway [79, 80]. However, this effect is likely to be small given the extent of the changes observed and the absence of large vessels between the ASM and airway lumen [76]. Secondly, dilatation of vessels or rapid movement of fluid from vessels into peribronchial tissues may dissociate the elastic load of the lung parenchyma from contracting smooth muscle [81]. Thirdly, exudation of fluid may also contribute to increased thickness of the inner airway wall. Fourthly, it has been shown that reduced bronchial blood flow may prolong and accentuate airway responses to bronchoconstricting stimuli, probably by delaying the removal of an inhaled stimulus [82]. The extent to which these effects operate in asthma or COPD is unknown.

Quantitative studies of the local innervation of the airways in asthma have shown conflicting responses regarding vaso-active

intestinal peptide-positive nerves [83, 84]. Substance P-immunoreactive nerves were increased in cases of cough without any difference between cases of asthma and normal controls [85]. There are no published quantitative studies of nerve tissues in COPD, although it has been suggested that there is more evidence for disrupted neural control in asthma than in COPD [86].

The cartilage may act as a significant load, which limits shortening of the ASM, because its removal or a reduction in its stiffness results in excessive airway narrowing following stimulation of the ASM [87, 88]. The amount of cartilage has been shown to be increased or unchanged in asthma [32], and one study showed reduced cartilage in COPD [89].

EVIDENCE THAT REMODELLING INFLUENCES THE CLINICAL EXPRESSION OF AIRWAY DISEASES

Symptoms

The diagnosis of asthma is based largely on a characteristic history of intermittent symptoms, predominantly wheezing, chest tightness, shortness of breath, cough and mucus production. The variability of these symptoms, their precipitation by factors such as exercise, cold air, allergens, viral respiratory infections or specific agents such as aspirin, their relief by bronchodilators and control by inhaled corticosteroids, form the basis of the clinical diagnosis of asthma. Doctor-diagnosed asthma is the current gold standard for diagnosing asthma in most population studies. It is considered that wheeze, chest tightness and breathlessness (and possibly cough) result from excessive narrowing of large and small airways and subsequent air-trapping in asthma. Although COPD is defined by abnormalities of lung function, the recent Global Initiative for Chronic Obstructive Lung Diseases (GOLD) classification [5] recognises that chronic bronchitis (daily cough and sputum production) may be present with or without airflow obstruction. The symptoms of COPD are in part related to the remodelling of airways and in part related to changes in parenchymal structure and function. The latter include reduced elastic recoil, reduced alveolar surface area and ventilation-perfusion inequality due to emphysema, altered chest wall mechanics due to chronic hyperinflation, and systemic problems such as muscle wasting in severe cases. However, it is clear that in COPD narrowing of the airway lumen also contributes to symptoms.

Cough and sputum production

In asthma, cough may be dry or associated with the production of clear or discoloured sputum. Dry cough commonly accompanies shortness of breath, chest tightness and wheeze, and is thought to be related to airway narrowing although the exact mechanisms in relation to airway remodelling remain unclear. The secretion of mucus into the airway lumen will also cause symptoms based on the resultant narrowing of the airway lumen, as will be discussed in the following section. Stimulation of cough receptors in the central airways may be important. However, it is also conceivable that stimulation is due to air-trapping/hyperinflation with activation of stretch fibres, some of which may be due to mucus occlusion of the small airways [90]. Productive cough in asthma, with phlegm that is normally clear or white in colour, is readily accounted for by the increased volume of goblet cells and submucosal

mucous glands [13, 17, 20, 91]. Discolouration of the sputum may be due to the accumulation of neutrophils in relation to superimposed infection or, occasionally in noninfectious exacerbations, due to neutrophils or eosinophils. Persistent productive cough is prominent in some patients with asthma and may overlap with syndromes associated with bronchiectasis such as allergic bronchopulmonary fungal disease. As mentioned previously, most asthmatic patients do not spontaneously produce sputum, although the inflammatory cell profiles of spontaneously produced and induced sputum are similar [92].

Daily cough and sputum production, or chronic bronchitis, is a cardinal feature of COPD [5]. This symptom was one of the first to be clearly related to airway remodelling. The Reid index, a measure of increased volume of submucosal mucous glands, was shown to correlate with chronic productive cough in a number of studies of COPD and smoking [49, 91, 93–96]. However, mucous gland size is not the only determinant of sputum production: MULLEN *et al.* [97] showed that symptoms in smokers with chronic bronchitis were more strongly related to central airway inflammation rather than the Reid index, and SAETTA *et al.* [98] also showed a relationship of chronic bronchitis symptoms to airway inflammation, specifically neutrophils, macrophages and CD8+ lymphocytes.

Wheeze, cough, chest tightness and shortness of breath: mechanisms of airway narrowing

The lumen of an airway can be narrowed due to: 1) accumulation of fluid, cells or mucus; 2) encroachment of a thickened airway wall with or without smooth muscle shortening; 3) shortening of the smooth muscle surrounding the airway; or 4) collapse of the airway wall (fig. 3a–d, respectively).

Accumulation of mucus and cells in the airway lumen

The accumulation of mucus in the airways in cases of asthma is well recognised [11, 13, 19, 99]. It is associated with hypertrophy and hyperplasia of epithelial goblet cells (fig. 1) [13, 20] and hypertrophy of submucosal glands (fig. 4) [13, 17]. Remodelling of the mucus-secreting glands in asthma will result in increased mucus production, but it is likely that accumulation and pooling of secreted mucus over time is required to produce excessive airway narrowing [19, 100]. Smooth muscle shortening around the airway will amplify these effects [101]. Although quantitative studies of the airway epithelium in asthma do not show significant differences in epithelial loss in asthma [17, 18], the increased epithelial cells observed in intraluminal mucus [23], in induced sputum, and in bronchoalveolar lavage from cases of mild asthma

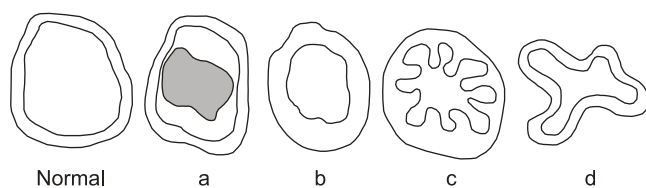


FIGURE 3. Schematic cross-section of the airway. Narrowing of the airway lumen may be due to: a) mucus, cells or other material within the lumen, b) thickening of the airway wall that encroaches on the lumen, c) shortening of smooth muscle around the lumen, d) collapse of the airway wall into the lumen.

suggest increased epithelial turnover in asthma. Impaired clearance of mucus is present during exacerbations of asthma [102] and is a particular feature of fatal asthma [19]. The rheological properties of mucus are altered in asthma [103, 104] which may lead to reduced clearance and accumulation of mucus within the airways. Although increased luminal mucus is seen in *post mortem* cases of mild asthma [105] and is almost invariably a feature of cases of fatal asthma [99], increased mucus production is not a common clinical feature of mild-to-moderate asthma. In fact, the collection of spontaneously produced sputum samples from asthma is difficult and induction with agents such as hypertonic saline is often required [106].

In COPD, mucus can accumulate in the airway causing narrowing of the lumen [107] and this accumulation has been correlated with the degree of airflow obstruction [27]. Cigarette smoking is also associated with changes in the epithelium including goblet cell hyperplasia [29] and metaplasia and inflammation of the submucosal mucous glands [97, 98].

Thickened airway wall encroaching on the airway lumen

Numerous studies have shown that the airway walls are thicker in asthma in both mild and severe cases [32]. Thickening of the airway wall in asthma is related to severity and involves all components of the airway, including the epithelium, reticular basement membrane, submucosa, smooth muscle and mucous glands [17]. Contributing factors include oedema and bronchial vessel congestion and proliferation [74, 75, 108]. Despite these changes, the increased thickness of the airway wall is not usually associated with narrowing of the airway lumen in asthma [109, 110]. It is calculated that, by itself, the increased thickness of the airway wall observed in asthma will increase airway resistance by <10% of values observed in nonasthmatic airways [79, 101]. Not surprisingly, the majority of individuals with asthma, who have mild-to-moderate symptoms, have lung function that is within the normal range. It seems unlikely that there is a mechanism for sudden thickening or enlargement of the submucosa which

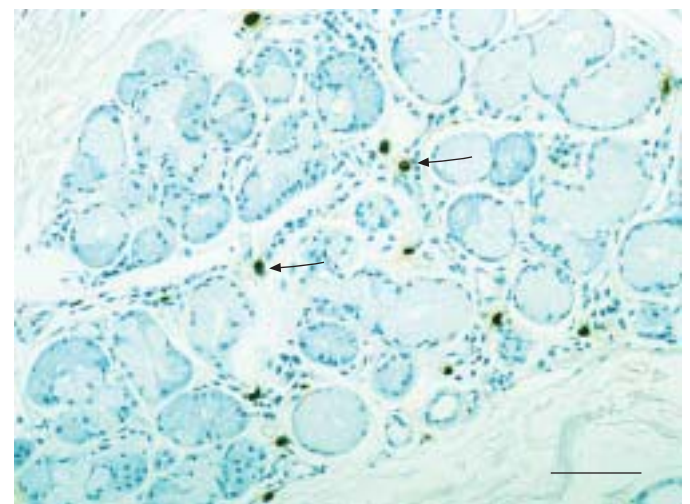


FIGURE 4. Detail of submucosa of a large airway from a case of fatal asthma. Submucosal mucous gland with numerous acini and interspersed intact mast cells (arrows). Tryptase antibody with haematoxylin counterstain. Scale bar=50 μ m.

might encroach upon the lumen. In particular, unlike in the nose, there does not appear to be any large capacitance vessels in the submucosa in humans with or without asthma [75]. There are more [74] and bigger [75] vessels below the airway epithelium in asthma but these have a relatively small space-occupying effect.

In COPD, the lumen of the airways are significantly narrowed, compared with subjects without airflow obstruction and compared with asthma cases [109]. Many studies have shown a correlation between abnormal lung function and lumen dimensions of the small airways in patients with COPD [27, 43, 44, 46, 111–114]. In general, these studies have shown a reduction in the diameter of small airways or an increase in the number of small airways with reduced internal diameters, compared with controls. These studies confirm the important role of small airways remodelling in COPD [44, 115]. Due to the large total cross-sectional area of the small airways [116], considerable inflammation and/or remodelling may occur before symptoms and abnormalities of lung function become apparent [117]. There have been few studies of the larger airways in COPD. TIDDENS *et al.* [48] studied cartilaginous airways from patients with airflow obstruction and showed that the thickness of the airway wall, although not the amount of smooth muscle, correlated inversely with measures of airflow. They also showed that these structural and functional changes were related to airway inflammation.

The increased thickness of the airway wall in COPD is not as prominent as that seen in asthma [44]. However, in a similar manner to asthma, the increased thickness of the airways in COPD involves the epithelium, reticular basement membrane, ASM and submucosal mucous glands in an inflammatory and fibrotic process [43, 70]. The fixed encroachment of the airway wall on the airway lumen in COPD is not reversed by bronchodilators and accounts for at least some of the persistently increased resistance to airflow. Therefore, in COPD, airway remodelling increases the thickness of the airway wall and directly results in airway narrowing. In contrast, in most (but not all) cases of asthma the increased thickness of the airway wall is greater than that in COPD but the effect on luminal size is much less, so that the functional effects of airway remodelling are only manifest with smooth muscle shortening. Of course increased airway wall thickness may also result in fixed airway narrowing and irreversible airflow obstruction in some patients with asthma [118].

Smooth muscle shortening

Shortening of the smooth muscle around the airway causes narrowing of the lumen. The amount of narrowing that occurs is dependent on three factors: 1) the proportion, relative to the resting length, that the muscle shortens; 2) the proportion of the airway perimeter that is made up of smooth muscle; and 3) the thickness of the airway wall (including smooth muscle, submucosa, epithelium and surface mucus and fluid) within the outer boundary of the smooth muscle layer [101]. The relationship of airway lumen size to smooth muscle shortening and wall thickness has been explored by MORENO *et al.* [101] and expanded in a number of mathematical models [119–121] and is discussed in more detail elsewhere in this series. In asthma, the increased thickness of the airway wall has a space-occupying effect that encroaches on the lumen, minimally

(asymptomatic) at rest and markedly (mild to severe symptoms) when the surrounding smooth muscle shortens, even if the degree of shortening is the same as that in nonasthmatic airways [79, 101, 119, 122]. It should be pointed out, however, that there may be opposing effects, due to the increased load associated with increased airway wall thickness, on the relationship between changes in lung function and dose of inhaled stimulus, as suggested by the study of NIIMI *et al.* [123].

The loads that oppose shortening of ASM may be critical in determining how much the airway wall can encroach on the airway lumen as the smooth muscle develops force [101]. These include external (to the smooth muscle layer) and internal (within the smooth muscle layer) loads. External loads include the mechanical properties of the airway wall or surrounding lung parenchyma that resist deformation. The structural properties of the airway wall and how these may change in asthma are dealt with in the subsequent section.

Loss of elastic recoil of the lung parenchyma has been observed in the lungs of some patients with asthma, even after successful treatment has restored normal lung function [124–126]. The structural bases for these changes are not known, although loss of alveolar attachments and peribronchiolar damage to elastin has been reported in cases of fatal asthma [64]. Abnormal elastic recoil of the lung, present in many subjects with moderate or severe persistent asthma, has not been associated with abnormalities of the lung parenchyma assessed by computed tomography (CT) scan or measurement of diffusing capacity [127]. Although elastic recoil of the lung in patients with asthma has not been shown to be related to duration or severity of asthma [128], abnormal elastic recoil has been identified as a risk factor for near fatal asthma [129].

Less is known about the internal loads that limit muscle shortening, but it is likely that remodelling has the potential to greatly alter them. The internal mechanical properties of structures within the smooth muscle [130–132] may determine both maximum shortening and the velocity of shortening, both of which will influence maximal airway narrowing. In addition, the mechanical properties of increased ECM surrounding smooth muscle cells [62] will affect smooth muscle shortening by altering the freedom of movement between cells and matrix and by constraining muscle cell width [133].

Airway wall collapse

The compliance of the airway wall will influence the loads that oppose ASM shortening. The mechanical properties of the airway wall are partly determined by the properties of the constituents including the epithelium, ECM, glands, blood vessels and cartilage (where present), by the tone of the ASM [134] and possibly by the dynamic changes in fluid and matrix interactions, as seen in articular cartilage [62]. The study of BRACKEL *et al.* [135] showed that the compliance of the central airways (from the trachea to the mid-part of the right lower lobe) was reduced in patients with asthma. Reduced distensibility of the airways has been observed in patients with asthma [136], has been related to the thickness of subepithelial fibrosis [137] and has been attributed to increased numbers of fibroblasts [54] and collagen deposition [52, 54, 138, 139] below the basement membrane in asthma. The increase in submucosal collagen in

asthma is more likely to resist airway dilatation than compression, due to the fibrillar nature of collagen [140].

It is also possible that changes in the ECM within the airway wall and in the external loads may increase collapsibility. Disruption of collagen due to the presence of metalloproteinases [141] may alter its mechanical properties. This is supported by *in vitro* studies showing increased responsiveness of human ASM after treatment with collagenase [131]. Loss of elastin may contribute to altered airway mechanics in asthma, possibly increasing deformability, with a tendency towards increased airway narrowing [63, 65]. Thus, in some asthma cases the airways could be more collapsible, which will increase the tendency for airway closure.

In addition to persistent narrowing of the airways in COPD, there is a component of dynamic collapse that further increases airway narrowing and limits maximal airflow [142]. This is probably due to loss of elastic recoil of the lung parenchyma [125] and loss of attachments to the airway wall [143] but may also be due to reduced stiffness of the airway wall. TIDDENS *et al.* [134] studied isolated airways from smokers who underwent surgery. They found that the amount and tone of the smooth muscle, but not total airway wall area, were determinants of airway mechanical properties including hysteresis, compliance and collapsibility. In an *in vitro* study, OPAZO SAEZ *et al.* [144] found that small airways from smokers with airflow obstruction had more smooth muscle and were able to generate more force and stress (force/cross-sectional area of ASM) *in vitro* in response to acetylcholine than those from smokers without airflow obstruction. Therefore, it is likely that in COPD, if airway wall compliance is normal or decreased, that smooth muscle stimulation will lead to increased airway narrowing. These studies show that interactions between the ECM and smooth muscle within the airway wall in subjects with COPD will determine the mechanical properties of the airway wall and the degree to which the airway lumen can narrow, either passively or as a result of smooth muscle shortening.

Loss of parenchymal support is evident in patients with COPD where lung recoil is reduced due to emphysema [145, 146] and where the number of alveolar attachments around the airways is reduced due to emphysematous destruction and peribronchiolar inflammation [143]. Loss of alveolar attachments has been related to decreased airflow [147], to the reduced bronchodilating effect of a deep inspiration [148, 149] and to abnormal distribution of ventilation [150].

AIRWAY HYPERRESPONSIVENESS

Early dissertations on asthma described the characteristic increased sensitivity of asthmatic patients to inhaled allergens and other irritants [151] that has become known as AHR and has been shown to be a diagnostic feature of asthma [152–154]. An early study showed that AHR was related to severity of symptoms over long periods (months) rather than those in more recent days or weeks [155], suggesting that AHR is at least partly determined by remodelling, which probably only changes over long periods [156–158]. It should be recognised however, that some factors which influence AHR may change acutely with seasonal asthma [159] and following viral infections [160]. Similarly, the reversibility of airflow obstruction by

bronchodilators or in response to corticosteroid therapy has been used as a definition of asthma [153], differentiating it from COPD. AHR in asthma can be demonstrated to a wide range of stimuli such as methacholine, histamine, cold/dry air, exercise, nonisotonic stimuli or specific allergens to which the patient is sensitised [161]. The degree of AHR is loosely related to clinical severity of asthma, although not to baseline lung function, and severe AHR may be seen in patients with normal lung function [153].

The dose–response curve: its relationship to remodelling

It has been suggested that different aspects of airway wall remodelling may independently influence the sensitivity (position relative to the horizontal/dose axis), the reactivity (slope of the linear part of the curve) and the maximum response [162] of the dose–response curve. For example, inflammation and altered permeability of the airway wall may influence the sensitivity, whereas thickness of the airway wall may influence the reactivity and maximal response. Increasing the deposition of an inhaled stimulus may increase the sensitivity of the response without changing the maximal response [163]. NIIMI *et al.* [123] assessed airway inflammation using induced sputum, and airway wall thickness using CT scans in subjects with asthma and related these measures to aspects of the dose–response to inhaled methacholine. While sputum eosinophils were related to airway sensitivity, it was found that reactivity was inversely related to airway wall thickness. The reduced reactivity of the response may have resulted from increased stiffness of the thicker airway wall. An alternative explanation is that reduced access to the smooth muscle by the inhaled methacholine due to increased wall thickness may have reduced the stimulation of the smooth muscle. The findings of NIIMI *et al.* [123] highlight the need to consider not only the space-occupying effects of remodelling, but also the changes in the mechanical properties that remodelling may induce. LITTLE *et al.* [164] found no correlation between AHR sensitivity and increased airway wall thickness assessed by CT scan. BOULET *et al.* [165] found a correlation between airway wall thickness measured on CT scan and methacholine sensitivity in patients with asthma who had irreversible airflow obstruction, but not in patients with asthma who had near-normal lung function. Therefore, the reported data on the relation of airway structure by CT scan and AHR remain somewhat conflicting.

AHR and remodelling in asthma

Some studies suggest that easier access of the stimulus to epithelial and submucosal sites may enhance AHR. OHASHI *et al.* [166] found a positive correlation between the loss of epithelial tight junctions and AHR, while the number of epithelial cells in bronchoalveolar lavage fluid has also been reported to correlate with AHR [20]. JEFFREY *et al.* [15] observed an association of epithelial loss and AHR although this was not replicated in another study [16]. AHR has also been shown to be related to the thickness of the reticular basement membrane measured on biopsy [26, 158, 167–169]. However, a number of studies have not shown such a relationship [139, 170]. The number of subepithelial fibroblasts was suggested to be related to AHR in one study [171]. The results of studies correlating ASM function measured *in vitro* with airway responsiveness measured *in vivo* have varied [172–176]. Few studies have

attempted to correlate smooth muscle dimensions with AHR. WOODRUFF *et al.* [35] studied smooth muscle in biopsies from asthmatic and nonasthmatic subjects. Although they found that the volume fraction of smooth muscle in the submucosa did not correlate with AHR when the asthmatic subjects were analysed separately, there was a significant correlation when control and asthmatic subjects were analysed as a group. This supports the suggestion that the amount of smooth muscle has a direct effect on AHR in asthma. The results of many studies have not shown consistent relationships between airway remodelling and AHR. It is possible that this variation may be due to spurious relationships between individual elements of remodelled airways and the integrated behaviour of the remodelled airways that results in AHR.

COPD, AHR and remodelling

AHR to inhaled stimuli such as histamine and methacholine occurs in patients with COPD [177]. In contrast to patients with asthma without fixed airflow obstruction, the response to bronchoconstrictors is related to the degree of baseline airflow obstruction in COPD [9, 178, 179]. This is predictable since the response is measured as a percentage change from baseline and, by definition, patients with COPD start with a lower baseline level of lung function. Therefore, it is likely that the AHR seen in COPD simply reflects normal smooth muscle shortening around an airway that is already narrowed due to airway wall remodelling. This is supported by the study of RIESS *et al.* [180] who showed that airway responsiveness in 77 smokers who underwent lung resection was negatively related not only to baseline lung function, but also to lung recoil, and was positively related to airway wall thickness. These findings support theoretical models based on measured airway dimensions [140]. The bronchodilator response has also been examined in COPD and was negatively related to bronchiolar cell metaplasia and emphysema, although not related to ASM [181]. In a study that compared patients with centrilobular emphysema and patients with panlobular emphysema, FINKLESTEIN *et al.* [169] found that only those with centrilobular emphysema showed a relationship between wall thickness and airway responsiveness. Therefore, as in asthma, increased airway wall thickness could contribute to AHR in COPD by exaggerating the effects of smooth muscle shortening. However, in COPD, airway remodelling also contributes to reduced baseline airway calibre which in itself artefactually influences AHR. Finally, reduced lung recoil may also contribute to AHR in COPD.

AHR: asthma versus COPD

There are a number of differences in AHR between asthma and COPD. The dose–response curves are generally more left-shifted (increased sensitivity) in patients with asthma, compared with patients with COPD [182]. AHR in patients with asthma is characterised by the loss of the maximal or plateau response that is observed in normal subjects [182, 183]. One study has shown that plateau responses occur in patients with COPD [184] and another has not [185]. In addition, while patients with asthma respond to both direct smooth muscle/nerve stimulation and to indirect stimuli, patients with COPD generally respond only to direct smooth muscle stimulation with histamine or methacholine [9, 184]. The differences between AHR in asthma and COPD are intriguing, because if

stimulated airway segments from patients with COPD can produce more force than in nonobstructed patients, as discussed previously [144], then all subjects with COPD should have a loss of plateau response or demonstrate AHR to direct and indirect smooth muscle stimuli *in vivo*. The differences may lie in the relative mechanical properties of the smooth muscle (including length-tension and velocity of shortening considerations) and the airway wall in asthma and COPD. Other factors unrelated to remodelling, such as specific sensitisation, allergic airway inflammation, airway permeability and local neuro-humoral responses also need to be considered. Differences in the responses to adenosine between smokers and nonsmokers with COPD [186] suggest that inflammation may also modulate airway responses in some COPD patients.

CLINICAL SEVERITY OF AIRWAY DISEASE

Although many factors contribute to asthma severity, remodelling is almost certainly important. CT scans and morphometric studies of the proximal airways in asthma have shown a relationship between increased severity of disease and increased thickness of the airway wall [17, 110, 164, 187]. Pathological changes include thickness of the total airway wall, inner airway wall, the smooth muscle and mucous glands [17, 188]. Deposition of ECM proteins such as collagen [188] results in increased thickness of the reticular basement membrane in asthma, which has been shown to be related to severity in most [170, 189, 190] but not all [139] studies. More severe cases have been characterised by increased ASM [17, 188]. The degree of air-trapping, or hyperinflation (measured physiologically or radiologically), and unspecified remodelling also appear to contribute to asthma severity [170]. Finally, it is likely that genetic influences help determine the severity of asthma specifically through effects on airway remodelling [191, 192] and/or smooth muscle function [193].

There is a reproducible relationship between the clinical severity of asthma and AHR [152, 194]. To the extent that airway remodelling contributes to AHR as discussed previously, more marked changes of remodelling will result in more severe asthma. However, this is only a broad generalisation, since some aspects of remodelling may exaggerate airway narrowing whereas others may protect against airway narrowing (see below).

FIXED AIRFLOW OBSTRUCTION AND DECLINE IN LUNG FUNCTION IN ASTHMA

Fixed airflow obstruction

Asthma is associated with reduced lung function in large population studies, even those where most subjects have mild asthma [195–198]. The effect is more marked in asthmatics with more severe disease (fig. 5) [197] and in those with persistent symptoms [195]. Remodelling may result in abnormal lung function in patients with asthma early in life (or at the onset of disease) by preventing full lung growth or by accelerating the decline in lung function with age (see below). It should be noted that some patients with asthma also develop fixed airflow obstruction with predictable exertional dyspnoea, as is the case in patients with COPD [199, 200]. This overlap led Dutch investigators to suggest a common link between asthma and COPD and the hypothesis that smokers with features of asthma would be more likely to develop COPD [201].

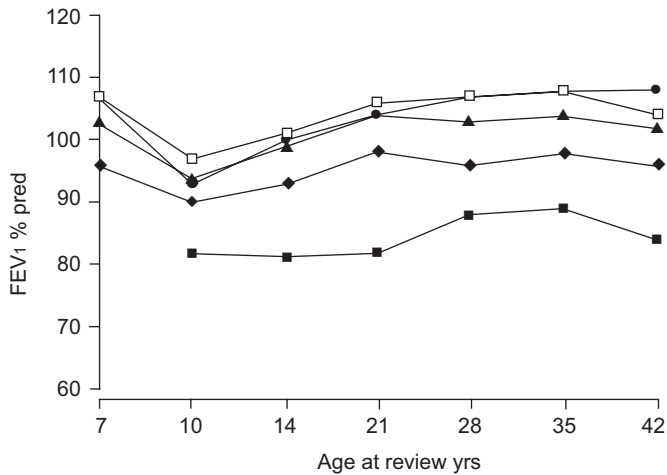


FIGURE 5. Data from the Melbourne cohort study [197] showing the per cent predicted (% pred) value of the forced expiratory volume in one second (FEV₁) from childhood to middle-age for subjects classified by status at first study as nonasthmatic (□), mild wheezy bronchitis (●), wheezy bronchitis (▲), mild asthma (◆) and severe asthma (■). The differences in lung function between the groups remained much the same over the course of 35 yrs.

The reduction in lung function in asthma appears to be related to duration and clinical severity [199, 200, 202–205]. In some studies of chronic, severe asthma, persistent airflow obstruction has been shown to be related both to inflammation, based upon exhaled nitric oxide and blood eosinophilia, and to remodelling, shown as increased wall thickness on CT scan [206]. Similarly, increased wall thickness on CT scan has been related to a decrease FEV₁ in asthma in some studies [110] but not others [164].

Decline in lung function

The rate of decline in lung function in adult patients with asthma appears to be increased relative to those without asthma [198, 207–209]. The Busselton health studies, which have followed a general population cohort since 1966, showed

an increased rate of decline in FEV₁ in asthmatic subjects [208], as was seen in the Copenhagen health studies [207], which also demonstrated that the decline in asthma was independent of cigarette smoking. Further study of the Busselton population [198] agreed with these findings but also showed that lung function at age 20 yrs was reduced in asthmatics but not in smokers (fig. 6). Some individuals with asthma may be genetically predisposed to a more rapid decline in lung function [210, 211] although the pathogenetic pathways related to this association are not established.

Risk factors for progressive loss of lung function in patients with asthma include smoking, atopy, adult onset, increased reversibility of obstruction, increased numbers of airway eosinophils and AHR [212]. TEN BRINKE *et al.* [213] showed that sputum eosinophilia, adult onset of asthma and AHR predicted fixed airflow obstruction in patients with asthma, although only sputum eosinophilia was significant by multivariate analyses. VONK *et al.* [214] found a relationship with pre-bronchodilator FEV₁, AHR and change in FEV₁ for irreversible airflow obstruction. The study of BROWN *et al.* [199] showed that fixed airflow obstruction, present after 2 weeks' treatment with oral corticosteroids, was related to the duration and severity of asthma. Whether these abnormalities are truly irreversible is questioned by studies that show improvements in lung function with high-dose corticosteroids [215], and highlights the need to distinguish "severe", "steroid-resistant", "poorly controlled" and "difficult-to-control" classifications of asthma [216]. These considerations are important since STRACHAN *et al.* [217] found that lung function (pre- and post-bronchodilator) in a cohort followed from childhood to 35 yrs of age was reduced most in those with persistent wheeze, less in those with intermittent (but current) wheeze and not at all in those with transient wheeze (no reported wheeze in the previous 12 months), which suggests that current symptoms partly account for deficits in lung function and, therefore, apparent increased decline in lung function. A similar conclusion was reached by ULRIK and LANGE [209], who found that the greatest decline in lung function occurred in patients with new (recent symptoms)

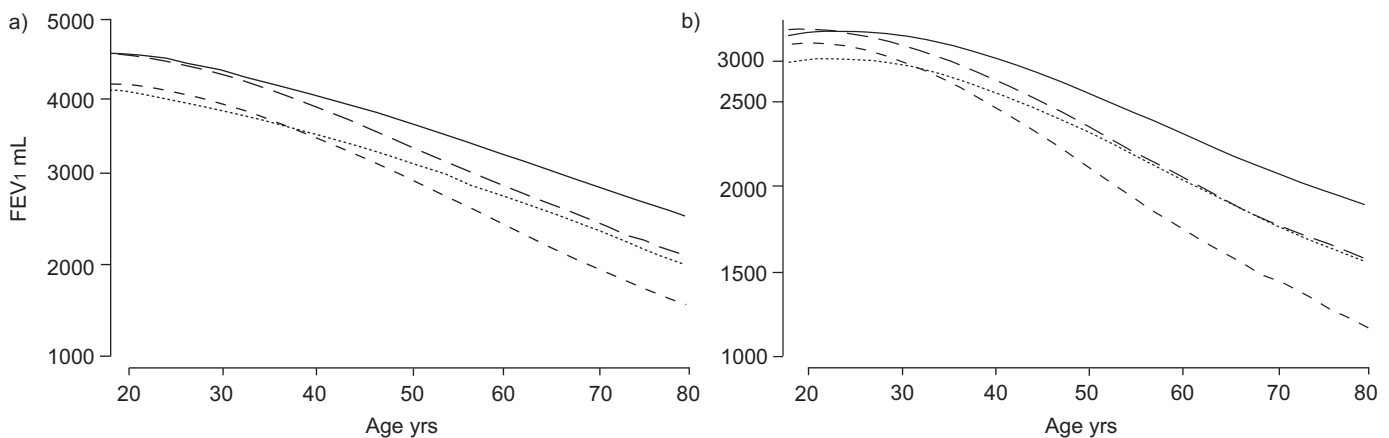


FIGURE 6. Data from the Busselton Health Study in a) males and b) females, showing height-corrected forced expiratory volume in one second (FEV₁) versus age for nonasthmatic nonsmokers (—), asthmatic nonsmokers (·····), nonasthmatic smokers (---) and asthmatic smokers (-.-). Subjects who have asthma and smoke have the fastest rate of decline of FEV₁ with age. Smoking is associated with a faster rate of decline compared with nonsmokers, but not a deficit in lung function at age 19 yrs. Asthma is associated with both an increased rate of decline of FEV₁ and a deficit in lung function at age 19 yrs. Reproduced from [198] with permission.

asthma. In this regard, decrements in lung function may result from exacerbations, which may be amenable to treatment [218].

The beginnings of remodelling: longitudinal studies of lung function

The deficits in lung function in subjects with asthma arise early in the course of the disease [196, 197]. RASMUSSEN *et al.* [196] found that abnormal lung function at age 26 in asthmatics was related to male sex, AHR at age 9 yrs, early onset of asthma and reduced lung function by age 9 yrs. SEARS *et al.* [195], reporting on the same cohort, showed that the risk of persistent wheeze from age 9–26 yrs was related to reduced lung function. In studies of children [196, 197] the group data do not suggest a faster decline in lung function, which supports the argument that deficits in lung function occur early in the disease and then remain stable (fig. 5). However, COVAR *et al.* [210], in a re-analysis of the Childhood Asthma Management Program data, suggested that ~25% of children had a persistent decline of lung function over the 5 yrs of the study that was not impacted by corticosteroid therapy. These children were younger, more likely to be male and were less atopic. The Tucson study [219] showed that abnormalities of lung function in infancy were more common in children with transient wheeze associated with maternal smoking and viral illness, than in those who later developed asthma. However, infants with persistent wheeze and subsequent asthma initially had normal lung function but developed abnormalities of airway function in association with asthma [219]. In a study from Perth, Western Australia, children with abnormalities of lung function and persistent wheeze at age 6 yrs had normal measures of airflow but increased airway responsiveness as infants [220]. Therefore, it seems likely that children with asthma may have relatively normal lung function as infants but develop fixed abnormalities of lung function early in the course of the disease, and that some have a progressive decline in lung function.

The beginnings of remodelling: pathological studies

Reticular basement membrane thickness is increased in the airways of children with mild-to-severe asthma, aged 6–16 yrs [221]. Biopsies of airways of younger children with wheezing illnesses and abnormal lung function have not shown changes of remodelling although these infants did not clearly have asthma [222]. Another study showed that in infants who subsequently developed asthma, airway pathology was evident before the onset of their disease [223], which supports the concept that changes of inflammation and remodelling occur early in the disease, even before symptoms are evident. From the few data available it would appear that remodelling (suggested by increased thickness of the reticular basement membrane) may begin early in asthma that arises in children, and may even precede the onset of symptoms. There are no data regarding the smooth muscle in infants or young children who are destined to develop asthma.

Studies of airway morphology in adult-onset asthma are few with virtually no longitudinal data. In patients with occupational asthma [224] and in elite skiers who develop asthma-like symptoms [225] elements of remodelling are observed, but the relation to onset and duration of disease is less clear. LAPRISE and BOULET [226] followed up a group of subjects with

asymptomatic AHR, and found that those who became symptomatic at follow-up had focal, rather than continuous, thickening of the reticular basement membrane. The reticular basement membrane increased in thickness with the development of symptoms and this was associated with increased numbers of activated CD4+ T cells. In a study of asthmatics who were asymptomatic for some years and not receiving treatment, VAN DEN TOORN *et al.* [227] found increased eosinophil numbers and thickening of the reticular basement membrane, to a level between that of nonasthmatic control subjects and symptomatic cases of asthma.

There are few studies of the effects of age *versus* duration on airway remodelling. BAI *et al.* [228] showed that older subjects with asthma had more smooth muscle and a trend towards more ECM, compared with younger subjects with asthma. These studies demonstrate progression of some aspects of airway remodelling in patients with new asthma symptoms and persistence of these changes in the absence of symptoms. The latter suggests that remodelling may remain asymptomatic for some time. The reasons for this are intriguing, although possibilities might include the requirement for an additional factor (such as inflammation), absence of an adequate stimulus to contract smooth muscle or the protective effect of remodelling itself (see below).

FIXED AIRFLOW OBSTRUCTION AND DECLINE IN LUNG FUNCTION IN COPD

COPD is defined by abnormal lung function. The GOLD classification [5, 7] categorises patients on the basis of symptoms and lung function. Previously, stage 0 was characterised by normal lung function but with chronic symptoms of cough and sputum production. Stages 1–4 are characterised as “mild”, “moderate”, “severe” and “very severe”, based upon decreasing levels of lung function. The fixed abnormalities of lung function in COPD are due to a variety of physiological abnormalities that include changes intrinsic to the airway wall as outlined previously. The thickness of the airway wall, including the epithelium, lamina propria, smooth muscle and adventitia are all related to the severity of airflow obstruction in patients with COPD [27, 43, 44, 46, 111–114].

Accelerated loss of lung function develops over time in susceptible smokers [229, 230], and is most likely due to genetic mechanisms [231]. Smoking cessation leads to normalisation of the rate of decline of lung function and restarting increases the rate of decline [229, 230]. Airway inflammation persists in patients with COPD and chronic bronchitis who have stopped smoking [232]. It has been suggested that mucus hypersecretion itself contributes to an increased decline of lung function, even after adjustment for the effects of cigarette smoking [233]. Whether this is due to persistent inflammation or progressive remodelling of the airways is not clear. The progressive loss of lung function associated with continued smoking is probably due, in large part, to continued parenchymal destruction by activated neutrophils, compromised lung defence mechanisms such as damage to alveolar type-II cells, and to airway inflammation [234].

AHR has been associated with a faster decline in lung function in subjects with COPD [235–237]. This association may be due

to progressive airway remodelling, although there are no longitudinal studies in COPD to assess this hypothesis, and other influences such as inflammation must be considered [27]. In this regard, the relationship between exacerbations of symptoms and rate of decline in lung function is interesting. It was initially considered that following an exacerbation, subjects with COPD tended to return to the level of lung function that was present prior to the exacerbation [238]. More recently, however, there is evidence to show that more frequent exacerbations are associated with a more rapid decline in FEV₁ [239] and it has been observed that some subjects do not fully recover, even 6 months following an exacerbation [240]. The rate of decline may, in part, be due to persistently increased airway inflammation [241] which may be related to reduced viral clearance [242] or to transient increases in inflammation [243], both of which may result in incremental airway remodelling and loss of lung function.

REVERSIBILITY OF REMODELLING AND ABNORMAL LUNG FUNCTION IN ASTHMA AND COPD

The degree to which abnormal lung function and airway remodelling are reversible is unclear. Some studies have shown that prolonged treatment with inhaled corticosteroids may reduce thickness of the reticular basement membrane [157, 158]. The use of lower doses and/or shorter periods of treatment has not shown any effect [158, 244]. While these changes appear to be related to remodelling processes across the airway wall, including changes in ASM [187, 189], no studies have directly addressed the effect of treatment on ASM volume. Using CT scans, NIMI *et al.* [245] showed partial reversibility of airway wall thickness in patients with asthma. The authors suggested that there were steroid-responsive (possibly inflammatory) and steroid-unresponsive components (possibly remodelling) that contributed to increased airway wall thickness in asthma. It is generally regarded that remodelling is an irreversible process, but, as suggested previously, this may be related to dose of treatment. Duration of treatment is also likely to be important since improvements in AHR may continue for periods of ≥ 6 months, despite treatment alleviating symptoms within weeks [156, 158, 246]. The study of WARD *et al.* [158] showed that although inflammation improved after 3 months of therapy, increased thickness of the reticular membrane was improved only at the second time-point of observation, which was 12 months after treatment onset. More studies of this nature are needed to examine the response of airway remodelling to treatment. This will also require the development of less invasive means of monitoring remodelling. Finally, the observations that the resistance of inflammation and remodelling to treatment in patients with COPD may be related to acetylation of DNA [247], shows that there are mechanisms that may account for nonreversible changes that may be targeted in the future.

FATAL ASTHMA

The pathology of asthma was initially described in case series of fatal asthma [13, 248–251]. These largely qualitative descriptions highlighted common features, including: the infiltration of the airway wall with eosinophils, neutrophils and lymphomononuclear cells; occlusion of airway lumens with mucus, cellular debris and eosinophil products (Charcot–Leiden

crystals); and thickening of the airway wall with prominence of the smooth muscle and mucous glands both in the epithelium and in the submucosa. The classic publication by HUBER and KOESSLER in 1922 [11] was the first to attempt to quantify the degree of remodelling in fatal asthma and showed that the airway wall was thicker, as was the smooth muscle, in both small and large airways in cases of asthma (fig. 7). Airway sizes from cases and controls were matched using the outer diameter of airways cut in transverse section. This might tend to over-estimate the wall thickness in asthma, since it is likely that, in these cases, airways from the same generation may artefactually have smaller outer diameters due to muscle contraction. This problem was overcome later by using the basement membrane perimeter to match airway size [252]. However, the results of HUBER and KOESSLER [11] showed large quantitative differences between controls and cases of fatal asthma. The paper (all 92 pages) is still well worth reading!

Since these early studies, quantification of remodelling in fatal asthma [32] has confirmed increased airway wall thickness, increased area of smooth muscle, increased mucus in the airway lumen, increased area of the submucosal mucous glands, increased deposition of ECM proteins and increased number and size of blood vessels within the airway wall, compared not only with nonasthmatic control cases but also with nonfatal, clinically mild-to-moderate cases of asthma. A recent study that used fractal geometry to describe measurements from airway casts showed that fatal asthma is associated with reduced complexity of the bronchial tree compared with control cases [1], although they were not significantly different from nonfatal cases.

Is fatal asthma just severe asthma or is it a distinct entity?

Clinical characteristics of those who die from asthma can be compared with those with near-fatal asthma, those with severe nonfatal asthma and those with mild-to-moderate asthma. The clinical severity of asthma is based upon: symptom frequency

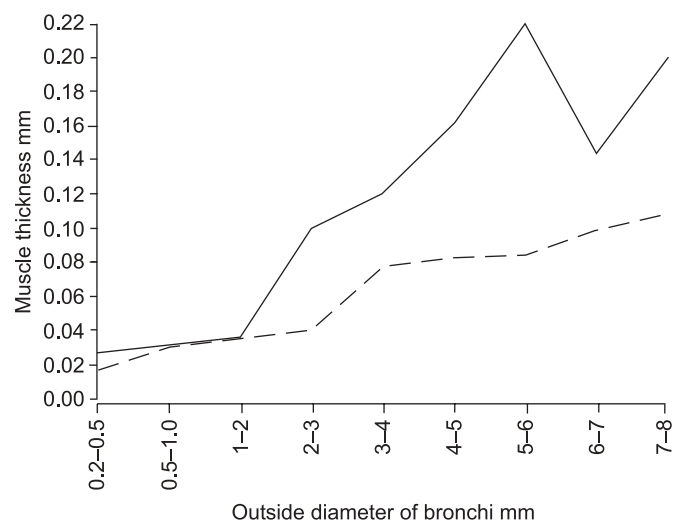


FIGURE 7. Thickness of airway smooth muscle plotted against outside bronchial diameter, measured in 1922 in *post mortem* tissues from nonasthmatic subjects (---) and in cases of fatal asthma (—). Modified from [11] with permission.

during the day or at night; treatment requirements, particularly oral corticosteroids; lung function; the frequency of emergency treatment and attacks (exacerbations); the frequency of admission to hospital; interruption of usual activities by asthma; and the history of near-fatal attacks [253]. Comparisons using these parameters suggest, almost universally, that those who die from asthma have a clinical history comparable with cases of severe asthma [17, 99, 254]. Comparisons of near-fatal and fatal cases of asthma show similar levels of clinical markers of severity and similar reported psychiatric problems [255], although whether these are higher than the general population (except for depression and anxiety) is controversial [256]. Another study showed that sudden onset of symptoms could occur in both groups [257]. These observations suggest that overall, clinically severe asthma is not obviously different from fatal asthma. Therefore, additional factors are required for an increased risk of death from asthma [258].

Potential physiological differences between asthma that is severe and asthma that is life-threatening or fatal include the increased response of the forced vital capacity to inhaled bronchoconstrictors in near-fatal asthma, compared with severe but not near-fatal cases, as well as a lower starting level of lung function [259]. This finding raises the possibility that remodelling may determine the nature and extent of airway narrowing and, therefore, the degree of the response and the likelihood of a fatal or near-fatal event. Interestingly, one of the strongest risk factors for a near-fatal or fatal asthma attack is a previous near-fatal event, which supports an underlying inflammatory or structural change in the lungs that contributes to the risk. There have been very few studies that have compared the genetics of fatal or near-fatal cases with less severe cases. Although adverse reactions to β -agonists have been suggested to be genetically driven [193], WEIR *et al.* [260] did not find differences between asthma groups of different severity with regard to polymorphisms of the β -adrenergic receptor.

Pathologically, cases of fatal asthma have shown similar changes to those seen in severe asthma. SYNEK *et al.* [261] showed increased numbers of eosinophils and epithelial CD3+ cells in large, but not small, airways in fatal asthma compared with mild-to-moderate asthma cases that died from nonrespiratory causes. No differences in numbers of mast cells, monocyte/macrophages or neutrophils or airway dimensions were reported. SOBONYA [262] studied six cases of severe allergic asthma with persistent airflow obstruction that had died from nonrespiratory causes. It was found that severe asthma cases had increased thickness of the reticular basement membrane and two cases of asthma had reduced luminal area of small airways associated with fibrosis, compared with nonasthma controls. In contrast to studies of fatal cases, increased thickness of the airway wall or the ASM was not observed, which suggests this factor may differentiate fatal from severe asthma, but further studies comparing airway dimensions in cases of clinically severe (but not fatal) asthma with cases of fatal asthma would better answer this question.

Is fatal asthma an extreme exacerbation?

Exposure to a number of stimuli can result in exacerbations of asthma and, in some cases, death. These include viral

respiratory infection [263, 264], allergens such as *Alternaria* [265], soybeans [266], pollen exposure associated with thunderstorms [267, 268] and specific occupational exposures [269]. These exposures will induce airway narrowing, the magnitude of which may be proportional to the underlying airway remodelling. In support of this, COCKCROFT and co-workers [270, 271] twice demonstrated that the response to inhaled allergen is related not only to the degree of allergic sensitisation shown by the skin weal response, but also to the degree of remodelling shown by the nonspecific response of the airways to inhaled histamine. The effects of stimulus strength also agree with responses predicted by models using airway dimensions of asthmatic and nonasthmatic subjects [101, 272].

What features of remodelling contribute to a fatal attack?

A number of studies have examined cases of fatal asthma in relation to the duration of the attack [273–276]. All studies showed a clear dichotomy of times to death from the onset of symptoms of the fatal attack and consistent differences in the ratio of numbers of eosinophils to neutrophils in the airway walls. Short-course cases demonstrated a decreased ratio of eosinophils to neutrophils, compared with long-course cases. This led SUR *et al.* [274] to postulate that a specific phenotype of asthmatic exacerbation that responded with an influx of neutrophils was predisposed to sudden death from asthma. CARROLL *et al.* [275] observed that the mucous gland area was greater in the short-course cases, and proposed that increased airway remodelling contributed to a rapid death after exposure to an inflammatory stimulus and that the decreased eosinophil-to-neutrophil ratio was due to capturing the early neutrophilic response that is characteristic of the usual response to an inflammatory stimulus [277]. It should be noted that the ratios of neutrophils to eosinophils will also be partly determined by the day-to-day level of eosinophilic inflammation present in the chronic state.

Multiple additional pathological features have been observed in fatal asthma. Luminal mucus and muscle shortening are likely to contribute to asthma death, as noted in studies where the lungs were not fixed in inflation [99, 258]. Increased shortening of ASM persisted despite increased *post mortem* serum levels of salbutamol in the short-course cases and there was more mucus accumulation in the airway lumens of long course-cases [276]. Finally, MAUAD *et al.* [64] reported loss of alveolar attachments with accompanying loss of small airway elastin in fatal asthma cases, perhaps leading to airway closure due to loss of parenchymal tethering. Thus, different mechanisms of excessive airway narrowing (mucus accumulation, loss of alveolar attachments or smooth muscle shortening) may account for death in fatal asthma.

IS REMODELLING ONLY DELETERIOUS OR CAN IT BE PROTECTIVE?

This question was recently addressed in a review by MCPARLAND *et al.* [278]. Much of the previous discussion suggests that remodelling may only have deleterious effects, such as excessive airway narrowing and symptoms, increased airway responsiveness, fixed airflow obstruction and accelerated decline in lung function. However, it should be considered that the increased thickness of the airway wall, smooth muscle, mucous glands and deposition of ECM

proteins may have beneficial effects, without which the effects of airway inflammation or the response of the airways to bronchoconstricting stimuli such as allergen would be even more detrimental to the patient. It is conceivable that an initial abnormality that impacts on airway structure and/or function is followed by a secondary remodelling process which minimises the effect of the initial change [279]. The latter may occur over a short or long period but may, in fact, be protective (fig. 8). Primary events might include increased ASM that is present at birth or develops within the first few years of life or to airway inflammation due to hypersensitivity to inhaled stimuli or an inherent abnormality of the epithelium [280].

These events may give rise to secondary protective events. The increase in ECM that occurs in asthma could serve to minimise shortening by adding to (rather than reducing) the loads against which smooth muscle shortens, thereby limiting airway narrowing. Such loads might include increased stiffness of the airway wall and increased series elastic loads that inhibit mucosal folding [41, 281]. MCPARLAND *et al.* [282] showed that loads opposing shortening of ASM might be substantial in relation to the ability of the smooth muscle to generate force. In addition, MILANESE *et al.* [283] showed an inverse relationship between thickness of the reticular basement membrane and airway responsiveness. The increase in ECM around bundles of ASM [62] may result in radial constraint that prevents thickening of smooth muscle cells as they shorten [133]. The distensibility of the airways is generally decreased in asthma [135–137] which may indicate increased airway wall stiffness and resistance to deformation. NIIMI *et al.* [123] showed an inverse relationship of airway reactivity (but not sensitivity) to airway wall thickness seen on CT scan. This

study supports the concept that airway wall thickening in asthma may serve as a protective response, at least in milder cases.

There are few studies addressing these relationships in COPD. The most relevant is the study of COLEBATCH *et al.* [284] which showed that some patients with COPD and “primary narrowing of airways” had reduced airway conductance relative to elastic recoil of the lung, suggesting reduced airway distensibility. Whether this equates to a reduced tendency to collapse was not examined. TIDDENS *et al.* [285] examined *in vivo* lung function in patients undergoing surgery and found that airway collapsibility, estimated *in vivo* as the transpulmonary pressure at zero flow, was not related to the level of pre-operative airflow obstruction or to airway wall dimensions, measured post-operatively in airways from the resected specimen of lung. In a similar study, TIDDENS *et al.* [134] found that although the airway wall thickness was related to measures of airflow obstruction it was not related to *in vitro* measurements of collapsibility of airway segments. This suggests that although remodelling reduces airflow and airway distensibility in COPD, it does not resist collapse of the airway and therefore does not have a protective role.

More studies are needed to try and establish the relative importance of remodelling as protective or deleterious. These studies will require the use of reliable but noninvasive methods of monitoring to allow longitudinal studies of the effects of interventions on remodelling and lung function. These studies are necessary because treatments aimed at remodelling may potentially reverse both harmful and helpful aspects of remodelling.

CONCLUSION

Although airway remodelling is closely related to airway inflammation, it has its own consequences with regard to symptoms, exacerbations (including fatal attacks of asthma), loss of lung function, decline in lung function and response to treatment. However, the specific elements of remodelling which contribute to the symptoms or progression of asthma and COPD remain poorly understood, partly due to the heterogeneity of these diseases and the complex interplay between numerous structural components within the airways. On the one hand, remodelling in asthma is associated with thicker and probably stiffer airway walls which encroach minimally on the airway lumen. The airways resist distension (except in mild cases), but smooth muscle shortening causes exaggerated airway narrowing. On the other hand, airway remodelling in COPD is less marked but encroaches on the airway lumen, limiting airflow and restricting dilatation, but probably not collapse. Changes in the epithelium, the smooth muscle and various ECM proteins appear to contribute to airway remodelling in both asthma and COPD. The relative amounts and interactions of these tissues are likely to account for the different natural histories of these two conditions.

Future directions for research in this area include: 1) establishing methods for assessing remodelling so that it can be more readily related to clinical aspects of airway disease, lung function, airway inflammation and responsiveness to treatment; 2) identifying the specific elements that contribute to airway remodelling, with particular with regard to the

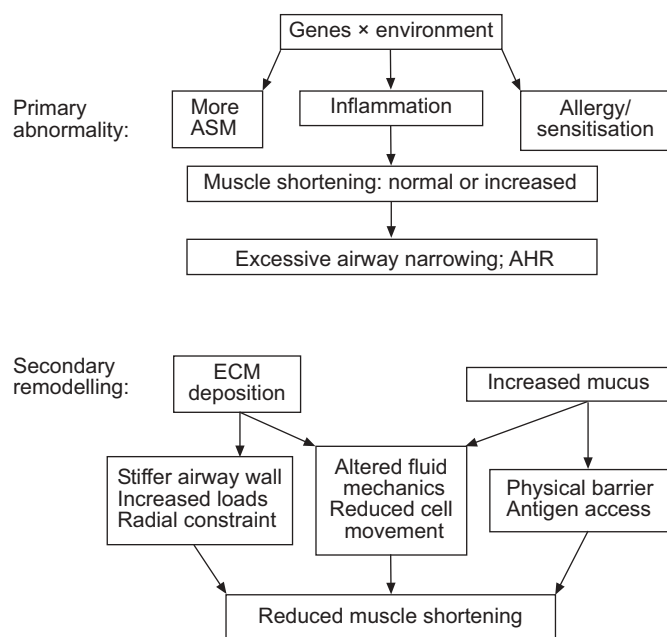


FIGURE 8. Schematic diagram of the relationships between remodelling that may result in symptoms and remodelling that may prevent or minimise symptoms. ASM: airway smooth muscle; AHR: airway hyperresponsiveness; ECM: extracellular matrix.

extracellular matrix (this seems to be an expanding area of current research); 3) comparing elements of remodelling in patients with chronic obstructive pulmonary disease with those in patients with asthma (who may or may not smoke cigarettes) who have fixed airflow obstruction; and 4) assessing longitudinal changes in remodelling.

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REFERENCES

- 1 Boser SR, Park H, Perry SF, Menache MG, Green FH. Fractal geometry of airway remodelling in human asthma. *Am J Respir Crit Care Med* 2005; 172: 817–823.
- 2 King GG, Eberl S, Salome CM, Young IH, Woolcock AJ. Differences in airway closure between normal and asthmatic subjects measured with single-photon emission computed tomography and technegas. *Am J Respir Crit Care Med* 1998; 158: 1900–1906.
- 3 Samee S, Altes T, Powers P, *et al.* Imaging the lungs in asthmatic patients by using hyperpolarized helium-3 magnetic resonance: assessment of response to methacholine and exercise challenge. *J Allergy Clin Immunol* 2003; 111: 1205–1211.
- 4 Global Strategy for Asthma Management and Prevention. NIH publication No 02-3659. www.ginasthma.org. Date last updated: 2005. Date last accessed: December, 12: 2006.
- 5 Pauwels RA, Buist AS, Calverley PM, Jenkins CR, Hurd SS. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease. NHLBI/WHO Global Initiative for Chronic Obstructive Lung Disease (GOLD) Workshop summary. *Am J Respir Crit Care Med* 2001; 163: 1256–1276.
- 6 McKenzie DK, Frith PA, Burdon JG, Town GI, Australian Lung Foundation, Thoracic Society of Australia and New Zealand. The COPDX Plan: Australian and New Zealand Guidelines for the Management of Chronic Obstructive Pulmonary Disease 2003. *Med J Aust* 2003; 178: Suppl., S7–S39.
- 7 National Institute of Health, National Heart, Lung and Blood Institute. Global Strategy for the Diagnosis, Management and Prevention of Chronic Obstructive Pulmonary Disease (GOLD). Bethesda, National Heart, Lung and Blood Institute. NIH publication No. 2701; NHLBI/WHO Workshop report, 2003.
- 8 Sin DD, Man SF. Skeletal muscle weakness, reduced exercise tolerance, and COPD: is systemic inflammation the missing link? *Thorax* 2006; 61: 1–3.
- 9 Woolcock AJ, Anderson SD, Peat JK, *et al.* Characteristics of bronchial hyperresponsiveness in chronic obstructive pulmonary disease and in asthma. *Am Rev Respir Dis* 1991; 143: 1438–1443.
- 10 Proud D, Chow CW. Role of viral infections in asthma and COPD. *Am J Respir Cell Mol Biol* 2006; 35: 513–518.
- 11 Huber HL, Koessler KK. The pathology of fatal asthma. *Arch Intern Med* 1922; 30: 689–760.
- 12 Houston JC, De Navasquez S, Trounce JR. A clinical and pathological study of fatal cases of status asthmaticus. *Thorax* 1953; 8: 207–213.
- 13 Dunnill MS. The pathology of asthma, with special reference to changes in the bronchial mucosa. *J Clin Path* 1960; 13: 27–33.
- 14 Laitinen LA, Heino M, Laitinen A, Kava T, Haahtela T. Damage of the airway epithelium and bronchial reactivity in patients with asthma. *Am Rev Respir Dis* 1985; 131: 599–606.
- 15 Jeffery PK, Wardlaw AJ, Nelson FC, Collins JV, Kay AB. Bronchial biopsies in asthma. An ultrastructural, quantitative study and correlation with hyperreactivity. *Am Rev Respir Dis* 1989; 140: 1745–1753.
- 16 Lozewicz S, Wells C, Gomez E, *et al.* Morphological integrity of the bronchial epithelium in mild asthma. *Thorax* 1990; 45: 12–15.
- 17 Carroll N, Elliot J, Morton A, James A. The structure of large and small airways in nonfatal and fatal asthma. *Am Rev Respir Dis* 1993; 147: 405–410.
- 18 Ordonez C, Ferrando R, Hyde DM, Wong HH, Fahy JV. Epithelial desquamation in asthma: artifact or pathology? *Am J Respir Crit Care Med* 2000; 162: 2324–2329.
- 19 Aikawa T, Shimura S, Sasaki H, Ebina M, Takishima T. Marked goblet cell hyperplasia with mucus accumulation in the airways of patients who died of severe acute asthma attack. *Chest* 1992; 101: 916–921.
- 20 Ordonez CL, Khashayar R, Wong HH, *et al.* Mild and moderate asthma is associated with airway goblet cell hyperplasia and abnormalities in mucin gene expression. *Am J Respir Crit Care Med* 2001; 163: 517–523.
- 21 Wardlaw AJ, Dunnette S, Gleich GJ, Collins JV, Kay AB. Eosinophils and mast cells in bronchoalveolar lavage in subjects with mild asthma. Relationship to bronchial hyperreactivity. *Am Rev Respir Dis* 1988; 137: 62–69.
- 22 Montefort S, Roberts JA, Beasley R, Holgate ST, Roche WR. The site of disruption of the bronchial epithelium in asthmatic and non-asthmatic subjects. *Thorax* 1992; 47: 499–503.
- 23 Naylor B. The shedding of the mucosa of the bronchial tree in asthma. *Thorax* 1962; 17: 69–72.
- 24 Campbell AM, Chanez P, Vignola AM, *et al.* Functional characteristics of bronchial epithelium obtained by brushing from asthmatic and normal subjects. *Am Rev Respir Dis* 1993; 147: 529–534.
- 25 Kirby JG, Hargreave FE, Gleich GJ, O'Byrne PM. Bronchoalveolar cell profiles of asthmatic and nonasthmatic subjects. *Am Rev Respir Dis* 1987; 136: 379–383.
- 26 Boulet LP, Laviolette M, Turcotte H, *et al.* Bronchial subepithelial fibrosis correlates with airway responsiveness to methacholine. *Chest* 1997; 112: 45–52.
- 27 Hogg JC, Chu F, Utokaparch S, *et al.* The nature of small-airway obstruction in chronic obstructive pulmonary disease. *N Engl J Med* 2004; 350: 2645–2653.
- 28 Lumsden AB, McLean A, Lamb D. Goblet and Clara cells of human distal airways: evidence for smoking induced changes in their numbers. *Thorax* 1984; 39: 844–849.

- 29 Saetta M, Turato G, Baraldo S, *et al.* Goblet cell hyperplasia and epithelial inflammation in peripheral airways of smokers with both symptoms of chronic bronchitis and chronic airflow limitation. *Am J Respir Crit Care Med* 2000; 161: 1016–1021.
- 30 Lee JS, Lippman SM, Benner SE, *et al.* Randomized placebo-controlled trial of isotretinoin in chemoprevention of bronchial squamous metaplasia. *J Clin Oncol* 1994; 12: 937–945.
- 31 Holgate ST, Lackie PM, Howarth PH, *et al.* Invited lecture: activation of the epithelial mesenchymal trophic unit in the pathogenesis of asthma. *Int Arch Allergy Immunol* 2001; 124: 253–258.
- 32 James AL. Relationship between airway wall thickness and airway hyperresponsiveness. In: Stewart AG, ed. *Airway Wall Remodelling in Asthma*. New York, CRC Press Inc., 1997; pp. 1–28.
- 33 Ebina M, Takahashi T, Chiba T, Motomiya M. Cellular hypertrophy and hyperplasia of airway smooth muscles underlying bronchial asthma. A 3-D morphometric study. *Am Rev Respir Dis* 1993; 148: 720–726.
- 34 Heard BE, Hossain S. Hyperplasia of bronchial muscle in asthma. *J Pathol* 1973; 110: 319–331.
- 35 Woodruff PG, Dolganov GM, Ferrando RE, *et al.* Hyperplasia of smooth muscle in mild to moderate asthma without changes in cell size or gene expression. *Am J Respir Crit Care Med* 2004; 169: 1001–1006.
- 36 Ebina M, Yaegashi H, Takahashi T, Motomiya M, Tanemura M. Distribution of smooth muscles along the bronchial tree. A morphometric study of ordinary autopsy lungs. *Am Rev Respir Dis* 1990; 141: 1322–1326.
- 37 Thomson RJ, Bramley AM, Schellenberg RR. Airway muscle stereology: implications for increased shortening in asthma. *Am J Respir Crit Care Med* 1996; 154: 749–757.
- 38 Hirst SJ, Walker TR, Chilvers ER. Phenotypic diversity and molecular mechanisms of airway smooth muscle proliferation in asthma. *Eur Respir J* 2000; 16: 159–177.
- 39 Zhou L, Li J, Goldsmith AM, *et al.* Human bronchial smooth muscle cell lines show a hypertrophic phenotype typical of severe asthma. *Am J Respir Crit Care Med* 2004; 169: 703–711.
- 40 Johnson PR, Roth M, Tamm M, *et al.* Airway smooth muscle cell proliferation is increased in asthma. *Am J Respir Crit Care Med* 2001; 164: 474–477.
- 41 Lambert RK, Codd SL, Alley MR, Pack RJ. Physical determinants of bronchial mucosal folding. *J Appl Physiol* 1994; 77: 1206–1216.
- 42 Hogg JC, Macklem PT, Thurlbeck WM. Site and nature of airway obstruction in chronic obstructive lung disease. *N Engl J Med* 1968; 278: 1355–1360.
- 43 Nagai A, West WW, Thurlbeck WM. The National Institutes of Health Intermittent Positive-Pressure Breathing trial: pathology studies. II. Correlation between morphologic findings, clinical findings, and evidence of expiratory air-flow obstruction. *Am Rev Respir Dis* 1985; 132: 946–953.
- 44 Bosken CH, Wiggs BR, Pare PD, Hogg JC. Small airway dimensions in smokers with obstruction to airflow. *Am Rev Respir Dis* 1990; 142: 563–570.
- 45 Kuwano K, Bosken CH, Pare PD, Bai TR, Wiggs BR, Hogg JC. Small airways dimensions in asthma and in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1993; 148: 1220–1225.
- 46 Cosio M, Ghezzi H, Hogg JC, *et al.* The relations between structural changes in small airways and pulmonary-function tests. *N Engl J Med* 1978; 298: 1277–1281.
- 47 Petty TL, Silvers GW, Stanford RE, Baird MD, Mitchell RS. Small airway pathology is related to increased closing capacity and abnormal slope of phase III in excised human lungs. *Am Rev Respir Dis* 1980; 121: 449–456.
- 48 Tiddens HA, Pare PD, Hogg JC, Hop WC, Lambert R, de Jongste JC. Cartilaginous airway dimensions and airflow obstruction in human lungs. *Am J Respir Crit Care Med* 1995; 152: 260–266.
- 49 Dunnill MS, Massarella GR, Anderson JA. A comparison of the quantitative anatomy of the bronchi in normal subjects, in status asthmaticus, in chronic bronchitis, and in emphysema. *Thorax* 1969; 24: 176–179.
- 50 Seow CY, Schellenberg RR, Pare PD. Structural and functional changes in the airway smooth muscle of asthmatic subjects. *Am J Respir Crit Care Med* 1998; 158: Suppl. 5, S179–S186.
- 51 Benayoun L, Letuve S, Druilhe A, *et al.* Regulation of peroxisome proliferator-activated receptor gamma expression in human asthmatic airways: relationship with proliferation, apoptosis, and airway remodelling. *Am J Respir Crit Care Med* 2001; 164: 1487–1494.
- 52 Carroll NG, Perry S, Karkhanis A, *et al.* The airway longitudinal elastic fiber network and mucosal folding in patients with asthma. *Am J Respir Crit Care Med* 2000; 161: 244–248.
- 53 Durieu I, Peyrol S, Gindre D, Bellon G, Durand DV, Pacheco Y. Subepithelial fibrosis and degradation of the bronchial extracellular matrix in cystic fibrosis. *Am J Respir Crit Care Med* 1998; 158: 580–588.
- 54 Roche WR, Beasley R, Williams JH, Holgate ST. Subepithelial fibrosis in the bronchi of asthmatics. *Lancet* 1989; 1: 520–524.
- 55 de Medeiros Matsushita M, da Silva LF, dos Santos MA, *et al.* Airway proteoglycans are differentially altered in fatal asthma. *J Pathol* 2005; 207: 102–110.
- 56 Todorova L, Gurcan E, Miller-Larsson A, Westergren-Thorsson G. Lung fibroblast proteoglycan production induced by serum is inhibited by budesonide and formoterol. *Am J Respir Cell Mol Biol* 2005; 34: 92–100.
- 57 Amin K, Ludviksdottir D, Janson C, *et al.* Inflammation and structural changes in the airways of patients with atopic and nonatopic asthma. BHR Group. *Am J Respir Crit Care Med* 2000; 162: 2295–2301.
- 58 Altraja A, Laitinen A, Virtanen I, *et al.* Expression of laminins in the airways in various types of asthmatic patients: a morphometric study. *Am J Respir Cell Mol Biol* 1996; 15: 482–488.
- 59 Huang J, Olivenstein R, Taha R, Hamid Q, Ludwig M. Enhanced proteoglycan deposition in the airway wall of atopic asthmatics. *Am J Respir Crit Care Med* 1999; 160: 725–729.
- 60 Amin K, Janson C, Boman G, Venge P. The extracellular deposition of mast cell products is increased in hypertrophic airways smooth muscles in allergic asthma but not in nonallergic asthma. *Allergy* 2005; 60: 1241–1247.

- 61 Westergren-Thorsson G, Chakir J, Lafreniere-Allard MJ, Boulet LP, Tremblay GM. Correlation between airway responsiveness and proteoglycan production by bronchial fibroblasts from normal and asthmatic subjects. *Int J Biochem Cell Biol* 2002; 34: 1256–1267.
- 62 Roberts C. Is asthma a fibrotic disease? *Chest* 1995; 107: Suppl. 3, 111S–117S.
- 63 Bousquet J, Lacoste JY, Chanez P, Vic P, Godard P, Michel FB. Bronchial elastic fibers in normal subjects and asthmatic patients. *Am J Respir Crit Care Med* 1996; 153: 1648–1654.
- 64 Mauad T, Silva LF, Santos MA, et al. Abnormal alveolar attachments with decreased elastic fiber content in distal lung in fatal asthma. *Am J Respir Crit Care Med* 2004; 170: 857–862.
- 65 Mauad T, Xavier AC, Saldiva PH, Dolhnikoff M. Elastosis and fragmentation of fibers of the elastic system in fatal asthma. *Am J Respir Crit Care Med* 1999; 160: 968–975.
- 66 Laitinen LA, Laitinen A, Altraja A, et al. Bronchial biopsy findings in intermittent or “early” asthma. *J Allergy Clin Immunol* 1996; 98: S3–S6.
- 67 Araujo BB, Silva LFF, Saldiva PHN, James AL, Dolhnikoff M, Mauad T. Elastic fiber content in airway smooth muscle in fatal asthma. *Am Rev Respir Dis* 2005; 2: A516.
- 68 Araujo BB, Silva LEF, Saldiva PHN, James A, Dolhnikoff M, Mauad T. Collagen content in airway smooth muscle in asthma. In: Society AT, ed. American Thoracic Society Annual Scientific Meeting, 2006; San Diego, USA. *Am J Respir Crit Care Med* 2006.
- 69 Pini L, Hamid Q, Shannon J, et al. Differences in proteoglycan deposition in the airways of moderate and severe asthmatics. *Eur Respir J* 2007; 37:188–196.
- 70 Wright JL, Lawson LM, Pare PD, Wiggs BJ, Kennedy S, Hogg JC. Morphology of peripheral airways in current smokers and ex-smokers. *Am Rev Respir Dis* 1983; 127: 474–477.
- 71 Amin K, Ekberg-Jansson A, Lofdahl CG, Venge P. Relationship between inflammatory cells and structural changes in the lungs of asymptomatic and never smokers: a biopsy study. *Thorax* 2003; 58: 135–142.
- 72 Ekberg-Jansson A, Amin K, Bake B, et al. Bronchial mucosal mast cells in asymptomatic smokers relation to structure, lung function and emphysema. *Respir Med* 2005; 99: 75–83.
- 73 van Straaten JF, Coers W, Noordhoek JA, et al. Proteoglycan changes in the extracellular matrix of lung tissue from patients with pulmonary emphysema. *Mod Pathol* 1999; 12: 697–705.
- 74 Li X, Wilson JW. Increased vascularity of the bronchial mucosa in mild asthma. *Am J Respir Crit Care Med* 1997; 156: 229–233.
- 75 Carroll NG, Cooke C, James AL. Bronchial blood vessel dimensions in asthma. *Am J Respir Crit Care Med* 1997; 155: 689–695.
- 76 Chu HW, Kraft M, Rex MD, Martin RJ. Evaluation of blood vessels and edema in the airways of asthma patients: regulation with clarithromycin treatment. *Chest* 2001; 120: 416–422.
- 77 Hoshino M, Takahashi M, Aoike N. Expression of vascular endothelial growth factor, basic fibroblast growth factor, and angiogenin immunoreactivity in asthmatic airways and its relationship to angiogenesis. *J Allergy Clin Immunol* 2001; 107: 295–301.
- 78 Green FH, Butt JC, James AL, Carroll NG. Abnormalities of the bronchial arteries in asthma. *Chest* 2006; 130: 1025–1033.
- 79 James AL, Pare PD, Hogg JC. The mechanics of airway narrowing in asthma. *Am Rev Respir Dis* 1989; 139: 242–246.
- 80 Wagner EM, Mitzner W. Effects of bronchial vascular engorgement on airway dimensions. *J Appl Physiol* 1996; 81: 293–301.
- 81 Uhlig T, Wildhaber JH, Carroll N, et al. Pulmonary vascular congestion selectively potentiates airway responsiveness in piglets. *Am J Respir Crit Care Med* 2000; 161: 1306–1313.
- 82 Wagner EM, Mitzner WA. Bronchial circulatory reversal of methacholine-induced airway constriction. *J Appl Physiol* 1990; 69: 1220–1224.
- 83 Ollerenshaw SL, Jarvis D, Sullivan CE, Woolcock AJ. Substance P immunoreactive nerves in airways from asthmatics and nonasthmatics. *Eur Respir J* 1991; 4: 673–682.
- 84 Howarth PH, Springall DR, Redington AE, Djukanovic R, Holgate ST, Polak JM. Neuropeptide-containing nerves in endobronchial biopsies from asthmatic and nonasthmatic subjects. *Am J Respir Cell Mol Biol* 1995; 13: 288–296.
- 85 Lee SY, Kim MK, Shin C, et al. Substance P-immunoreactive nerves in endobronchial biopsies in cough-variant asthma and classic asthma. *Respiration* 2003; 70: 49–53.
- 86 de Jongste JC, Jongejan RC, Kerrebijn KF. Control of airway caliber by autonomic nerves in asthma and in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1991; 43: 1421–1426.
- 87 James AL, Pare PD, Moreno RH, Hogg JC. Quantitative measurement of smooth muscle shortening in isolated pig trachea. *J Appl Physiol* 1987; 63: 1360–1365.
- 88 Moreno RH, Lisboa C, Hogg JC, Pare PD. Limitation of airway smooth muscle shortening by cartilage stiffness and lung elastic recoil in rabbits. *J Appl Physiol* 1993; 75: 738–744.
- 89 Tandon MK, Campbell AH. Bronchial cartilage in chronic bronchitis. *Thorax* 1969; 24: 607–612.
- 90 Widdicombe JG. Overview of neural pathways in allergy and asthma. *Pulm Pharmacol Ther* 2003; 16: 23–30.
- 91 Takizawa T, Thurlbeck WM. Muscle and mucous gland size in the major bronchi of patients with chronic bronchitis, asthma, and asthmatic bronchitis. *Am Rev Respir Dis* 1971; 104: 331–336.
- 92 Pizzichini MM, Popov TA, Efthimiadis A, et al. Spontaneous and induced sputum to measure indices of airway inflammation in asthma. *Am J Respir Crit Care Med* 1996; 154: 866–869.
- 93 Reid L. Measurement of the bronchial mucous gland layer: a diagnostic yardstick in chronic bronchitis. *Thorax* 1960; 15: 132–141.
- 94 Takizawa T, Thurlbeck WM. A comparative study of four methods of assessing the morphologic changes in chronic bronchitis. *Am Rev Respir Dis* 1971; 103: 774–783.
- 95 Vignola AM, Chanez P, Chiappara G, et al. Transforming growth factor-beta expression in mucosal biopsies in asthma and chronic bronchitis. *Am J Respir Crit Care Med* 1997; 156: 591–599.

- 96 Thurlbeck WM, Angus GE. A distribution curve for chronic bronchitis. *Thorax* 1964; 19: 436–442.
- 97 Mullen JB, Wright JL, Wiggs BR, Pare PD, Hogg JC. Reassessment of inflammation of airways in chronic bronchitis. *Br Med J (Clin Res Ed)* 1985; 291: 1235–1239.
- 98 Saetta M, Turato G, Facchini FM, *et al.* Inflammatory cells in the bronchial glands of smokers with chronic bronchitis. *Am J Respir Crit Care Med* 1997; 156: 1633–1639.
- 99 Kuyper LM, Pare PD, Hogg JC, *et al.* Characterisation of airway plugging in fatal asthma. *Am J Med* 2003; 115: 6–11.
- 100 James A. Theoretic effects of mucus gland discharge on airway gland resistance in asthma. *Chest* 1995; 107: Suppl. 3, 110S.
- 101 Moreno RH, Hogg JC, Pare PD. Mechanics of airway narrowing. *Am Rev Respir Dis* 1986; 133: 1171–1180.
- 102 Messina MS, O’Riordan TG, Smaldone GC. Changes in mucociliary clearance during acute exacerbations of asthma. *Am Rev Respir Dis* 1991; 143: 993–997.
- 103 Sheehan JK, Richardson PS, Fung DC, Howard M, Thornton DJ. Analysis of respiratory mucus glycoproteins in asthma: a detailed study from a patient who died in status asthmaticus. *Am J Respir Cell Mol Biol* 1995; 13: 748–756.
- 104 Shimura S, Sasaki T, Sasaki H, Takishima T. Chemical properties of bronchorrhea sputum in bronchial asthma. *Chest* 1988; 94: 1211–1215.
- 105 Carroll NG, Mutavdzic S, James AL. Increased mast cells and neutrophils in submucosal mucous glands and mucus plugging in patients with asthma. *Thorax* 2002; 57: 677–682.
- 106 Pin I, Gibson PG, Kolendowicz R, *et al.* Use of induced sputum cell counts to investigate airway inflammation in asthma. *Thorax* 1992; 47: 25–29.
- 107 Aikawa T, Shimura S, Sasaki H, Takishima T, Yaegashi H, Takahashi T. Morphometric analysis of intraluminal mucus in airways in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1989; 140: 477–482.
- 108 McDonald DM. Angiogenesis and remodelling of airway vasculature in chronic inflammation. *Am J Respir Crit Care Med* 2001; 164: Suppl. 10, S39–S45.
- 109 Nakano Y, Muller NL, King GG, *et al.* Quantitative assessment of airway remodelling using high-resolution CT. *Chest* 2002; 122: Suppl. 6, 271S–275S.
- 110 Niimi A, Matsumoto H, Amitani R, *et al.* Airway wall thickness in asthma assessed by computed tomography. Relation to clinical indices. *Am J Respir Crit Care Med* 2000; 162: 1518–1523.
- 111 Berend N, Woolcock AJ, Marlin GE. Correlation between the function and structure of the lung in smokers. *Am Rev Respir Dis* 1979; 119: 695–705.
- 112 Petty TL, Silvers GW, Stanford RE. Small airway dimension and size distribution in human lungs with an increased closing capacity. *Am Rev Respir Dis* 1982; 125: 535–539.
- 113 Matsuba K, Thurlbeck WM. The number and dimensions of small airways in nonemphysematous lungs. *Am Rev Respir Dis* 1971; 104: 516–524.
- 114 Matsuba K, Thurlbeck WM. The number and dimensions of small airways in emphysematous lungs. *Am J Pathol* 1972; 67: 265–275.
- 115 Yanai M, Sekizawa K, Ohru T, Sasaki H, Takishima T. Site of airway obstruction in pulmonary disease: direct measurement of intrabronchial pressure. *J Appl Physiol* 1992; 72: 1016–1023.
- 116 Weibel ER, Gomez DM. Architecture of the human lung. *Science* 1962; 137: 577–585.
- 117 Niewoehner DE, Kleinerman J, Rice DB. Pathologic changes in the peripheral airways of young cigarette smokers. *N Engl J Med* 1974; 291: 755–758.
- 118 Boulet LP, Turcotte H, Hudon C, Carrier G, Maltais F. Clinical, physiological and radiological features of asthma with incomplete reversibility of airflow obstruction compared with those of COPD. *Can Respir J* 1998; 5: 270–277.
- 119 Lambert RK, Wiggs BR, Kuwano K, Hogg JC, Pare PD. Functional significance of increased airway smooth muscle in asthma and COPD. *J Appl Physiol* 1993; 74: 2771–2781.
- 120 Lambert RK, Wilson TA, Hyatt RE, Rodarte JR. A computational model for expiratory flow. *J Appl Physiol* 1982; 52: 44–56.
- 121 Wiggs BR, Moreno R, Hogg JC, Hilliam C, Pare PD. A model of the mechanics of airway narrowing. *J Appl Physiol* 1990; 69: 849–860.
- 122 Wiggs BR, Bosken C, Pare PD, James A, Hogg JC. A model of airway narrowing in asthma and in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1992; 145: 1251–1258.
- 123 Niimi A, Matsumoto H, Takemura M, Ueda T, Chin K, Mishima M. Relationship of airway wall thickness to airway sensitivity and airway reactivity in asthma. *Am J Respir Crit Care Med* 2003; 168: 983–988.
- 124 Woolcock AJ, Read J. The static elastic properties of the lungs in asthma. *Am Rev Respir Dis* 1968; 98: 788–794.
- 125 Finucane KE, Colebatch HJ. Elastic behavior of the lung in patients with airway obstruction. *J Appl Physiol* 1969; 26: 330–338.
- 126 Gelb AF, Zamel N. Unsuspected pseudophysiologic emphysema in chronic persistent asthma. *Am J Respir Crit Care Med* 2000; 162: 1778–1782.
- 127 Gelb AF, Licuanan J, Shinar CM, Zamel N. Unsuspected loss of lung elastic recoil in chronic persistent asthma. *Chest* 2002; 121: 715–721.
- 128 Finucane KE, Greville HW, Brown PJ. Irreversible airflow obstruction. Evolution in asthma. *Med J Aust* 1985; 142: 602–604.
- 129 Gelb AF, Schein A, Nussbaum E, *et al.* Risk factors for near-fatal asthma. *Chest* 2004; 126: 1138–1146.
- 130 Fredberg JJ, Jones KA, Nathan M, *et al.* Friction in airway smooth muscle: mechanism, latch, and implications in asthma. *J Appl Physiol* 1996; 81: 2703–2712.
- 131 Bramley AM, Roberts CR, Schellenberg RR. Collagenase increases shortening of human bronchial smooth muscle *in vitro*. *Am J Respir Crit Care Med* 1995; 152: 1513–1517.
- 132 Bramley AM, Thomson RJ, Roberts CR, Schellenberg RR. Hypothesis: excessive bronchoconstriction in asthma is due to decreased airway elastance. *Eur Respir J* 1994; 7: 337–341.
- 133 Meiss RA, Pidaparti RM. Mechanical state of airway smooth muscle at very short lengths. *J Appl Physiol* 2004; 96: 655–667.

- 134** Tiddens HA, Hofhuis W, Bogaard JM, *et al.* Compliance, hysteresis, and collapsibility of human small airways. *Am J Respir Crit Care Med* 1999; 160: 1110–1118.
- 135** Brackel HJ, Pedersen OF, Mulder PG, Overbeek SE, Kerrebijn KF, Bogaard JM. Central airways behave more stiffly during forced expiration in patients with asthma. *Am J Respir Crit Care Med* 2000; 162: 896–904.
- 136** Wilson JW, Li X, Pain MC. The lack of distensibility of asthmatic airways. *Am Rev Respir Dis* 1993; 148: 806–809.
- 137** Ward C, Johns DP, Bish R, *et al.* Reduced airway distensibility, fixed airflow limitation, and airway wall remodelling in asthma. *Am J Respir Crit Care Med* 2001; 164: 1718–1721.
- 138** Brewster CE, Howarth PH, Djukanovic R, Wilson J, Holgate ST, Roche WR. Myofibroblasts and subepithelial fibrosis in bronchial asthma. *Am J Respir Cell Mol Biol* 1990; 3: 507–511.
- 139** Chu HW, Halliday JL, Martin RJ, Leung DY, Szeffler SJ, Wenzel SE. Collagen deposition in large airways may not differentiate severe asthma from milder forms of the disease. *Am J Respir Crit Care Med* 1998; 158: 1936–1944.
- 140** Pare PD, Wiggs BR, James A, Hogg JC, Bosken C. The comparative mechanics and morphology of airways in asthma and in chronic obstructive pulmonary disease. *Am Rev Respir Dis* 1991; 143: 1189–1193.
- 141** Vignola AM, Riccobono L, Mirabella A, *et al.* Sputum metalloproteinase-9/tissue inhibitor of metalloproteinase-1 ratio correlates with airflow obstruction in asthma and chronic bronchitis. *Am J Respir Crit Care Med* 1998; 158: 1945–1950.
- 142** Hyatt RE. Expiratory flow limitation. *J Appl Physiol* 1983; 55: 1–7.
- 143** Saetta M, Ghezzi H, Kim WD, *et al.* Loss of alveolar attachments in smokers. A morphometric correlate of lung function impairment. *Am Rev Respir Dis* 1985; 132: 894–900.
- 144** Opazo Saez AM, Seow CY, Pare PD. Peripheral airway smooth muscle mechanics in obstructive airways disease. *Am J Respir Crit Care Med* 2000; 161: 910–917.
- 145** Lamb D, McLean A, Gillyool M, Warren PM, Gould GA, MacNee W. Relation between distal airspace size, bronchiolar attachments, and lung function. *Thorax* 1993; 48: 1012–1017.
- 146** Saetta M, Shiner RJ, Angus GE, *et al.* Destructive index: a measurement of lung parenchymal destruction in smokers. *Am Rev Respir Dis* 1985; 131: 764–769.
- 147** Petty TL, Silvers GW, Stanford RE. Radial traction and small airways disease in excised human lungs. *Am Rev Respir Dis* 1986; 133: 132–135.
- 148** Corsico A, Milanese M, Baraldo S, *et al.* Small airway morphology and lung function in the transition from normality to chronic airway obstruction. *J Appl Physiol* 2003; 95: 441–447.
- 149** Scichilone N, Bruno A, Marchese R, Vignola AM, Toghiani A, Bellia V. Association between reduced bronchodilatory effect of deep inspiration and loss of alveolar attachments. *Respir Res* 2005; 6: 55–61.
- 150** Nagai A, Yamawaki I, Takizawa T, Thurlbeck WM. Alveolar attachments in emphysema of human lungs. *Am Rev Respir Dis* 1991; 144: 888–891.
- 151** Persson CG. On the medical history of xanthines and other remedies for asthma: a tribute to HH Salter. *Thorax* 1985; 40: 881–886.
- 152** Hargreave FE, Ryan G, Thomson NC, *et al.* Bronchial responsiveness to histamine or methacholine in asthma: measurement and clinical significance. *J Allergy Clin Immunol* 1981; 68: 347–355.
- 153** Ryan G, Latimer KM, Dolovich J, Hargreave FE. Bronchial responsiveness to histamine: relationship to diurnal variation of peak flow rate, improvement after bronchodilator, and airway calibre. *Thorax* 1982; 37: 423–429.
- 154** Boushey HA, Holtzman MJ, Sheller JR, Nadel JA. Bronchial hyperreactivity. *Am Rev Respir Dis* 1980; 121: 389–413.
- 155** Makino S. Clinical significance of bronchial sensitivity to acetylcholine and histamine in bronchial asthma. *J Allergy* 1966; 38: 127–142.
- 156** Reddel HK, Jenkins CR, Marks GB, *et al.* Optimal asthma control, starting with high doses of inhaled budesonide. *Eur Respir J* 2000; 16: 226–235.
- 157** Sont JK, Willems LN, Bel EH, van Krieken JH, Vandenberghe JP, Sterk PJ. Clinical control and histopathologic outcome of asthma when using airway hyperresponsiveness as an additional guide to long-term treatment. The AMPUL Study Group. *Am J Respir Crit Care Med* 1999; 159: 1043–1051.
- 158** Ward C, Pais M, Bish R, *et al.* Airway inflammation, basement membrane thickening and bronchial hyperresponsiveness in asthma. *Thorax* 2002; 57: 309–316.
- 159** Britton J, Chinn S, Burney P, Papacosta AO, Tattersfield A. Seasonal variation in bronchial reactivity in a community population. *J Allergy Clin Immunol* 1988; 82: 134–139.
- 160** Sterk PJ. Virus-induced airway hyperresponsiveness in man. *Eur Respir J* 1993; 6: 894–902.
- 161** Sterk PJ, Fabbri LM, Quanjer PH, *et al.* Airway responsiveness. Standardized challenge testing with pharmacological, physical and sensitizing stimuli in adults. Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society. *Eur Respir J Suppl* 1993; 16: 53–83.
- 162** Sterk PJ, Bel EH. Bronchial hyperresponsiveness: the need for a distinction between hypersensitivity and excessive airway narrowing. *Eur Respir J* 1989; 2: 267–274.
- 163** Guillemi S, James AL, Pare PD. Effect of breathing pattern during inhalation challenge on the shape and position of the dose-response curve. *Lung* 1989; 167: 95–106.
- 164** Little SA, Sproule MW, Cowan MD, *et al.* High resolution computed tomographic assessment of airway wall thickness in chronic asthma: reproducibility and relationship with lung function and severity. *Thorax* 2002; 57: 247–253.
- 165** Boulet L, Belanger M, Carrier G. Airway responsiveness and bronchial-wall thickness in asthma with or without fixed airflow obstruction. *Am J Respir Crit Care Med* 1995; 152: 865–871.
- 166** Ohashi Y, Motojima S, Fukuda T, Makino S. Airway hyperresponsiveness, increased intracellular spaces of bronchial epithelium, and increased infiltration of eosinophils and lymphocytes in bronchial mucosa in asthma. *Am Rev Respir Dis* 1992; 145: 1469–1476.

- 167** Lee SY, Kim SJ, Kwon SS, *et al.* Relation of airway reactivity and sensitivity with bronchial pathology in asthma. *J Asthma* 2002; 39: 537–544.
- 168** Chetta A, Foresi A, Del Donno M, *et al.* Bronchial responsiveness to distilled water and methacholine and its relationship to inflammation and remodelling of the airways in asthma. *Am J Respir Crit Care Med* 1996; 153: 910–917.
- 169** Finkelstein R, Ma HD, Ghezzi H, Whittaker K, Fraser RS, Cosio MG. Morphometry of small airways in smokers and its relationship to emphysema type and hyperresponsiveness. *Am J Respir Crit Care Med* 1995; 152: 267–276.
- 170** Wenzel SE, Schwartz LB, Langmack EL, *et al.* Evidence that severe asthma can be divided pathologically into two inflammatory subtypes with distinct physiologic and clinical characteristics. *Am J Respir Crit Care Med* 1999; 160: 1001–1008.
- 171** Hoshino M, Nakamura Y, Sim J, Shimojo J, Isogai S. Bronchial subepithelial fibrosis and expression of matrix metalloproteinase-9 in asthmatic airway inflammation. *J Allergy Clin Immunol* 1998; 102: 783–788.
- 172** Vincenc KS, Black JL, Yan K, Armour CL, Donnelly PD, Woolcock AJ. Comparison of *in vivo* and *in vitro* responses to histamine in human airways. *Am Rev Respir Dis* 1983; 128: 875–879.
- 173** Armour CL, Lazar NM, Schellenberg RR, *et al.* A comparison of *in vivo* and *in vitro* human airway reactivity to histamine. *Am Rev Respir Dis* 1984; 129: 907–910.
- 174** de Jongste JC, Mons H, Bonta IL, Kerrebijn KF. Human asthmatic airway responses *in vitro* - a case report. *Eur J Respir Dis* 1987; 70: 1–7.
- 175** Goldie RG, Spina D, Henry PJ, Lulich KM, Paterson JW. *In vitro* responsiveness of human asthmatic bronchus to carbachol, histamine, beta-adrenoceptor agonists and theophylline. *Br J Clin Pharmacol* 1986; 22: 669–676.
- 176** An SS, Fabry B, Trepast X, Wang N, Fredberg JJ. Do biophysical properties of the airway smooth muscle in culture predict airway hyperresponsiveness? *Am J Respir Cell Mol Biol* 2006; 35: 55–64.
- 177** Ramsdell JW, Nachtwey FJ, Moser KM. Bronchial hyperreactivity in chronic obstructive bronchitis. *Am Rev Respir Dis* 1982; 126: 829–832.
- 178** van Schayck CP, Dompeling E, Molema J, Folgering H, van Grunsven PM, van Weel C. Does bronchial hyperresponsiveness precede or follow airway obstruction in asthma or COPD? *Neth J Med* 1994; 45: 145–153.
- 179** Ramsdale EH, Morris MM, Roberts RS, Hargreave FE. Bronchial responsiveness to methacholine in chronic bronchitis: relationship to airflow obstruction and cold air responsiveness. *Thorax* 1984; 39: 912–918.
- 180** Riess A, Wiggs B, Verburgt L, Wright JL, Hogg JC, Pare PD. Morphologic determinants of airway responsiveness in chronic smokers. *Am J Respir Crit Care Med* 1996; 154: 1444–1449.
- 181** Nagai A, Thurlbeck WM, Konno K. Responsiveness and variability of airflow obstruction in chronic obstructive pulmonary disease. Clinicopathologic correlative studies. *Am J Respir Crit Care Med* 1995; 151: 635–639.
- 182** Woolcock AJ, Salome CM, Yan K. The shape of the dose-response curve to histamine in asthmatic and normal subjects. *Am Rev Respir Dis* 1984; 130: 71–75.
- 183** James A, Loughheed D, Pearce-Pinto G, Ryan G, Musk AW. Maximal airway narrowing in a general population. *Am Rev Respir Dis* 1992; 146: 895–899.
- 184** Du Toit JI, Woolcock AJ, Salome CM, Sundrum R, Black JL. Characteristics of bronchial hyperresponsiveness in smokers with chronic air-flow limitation. *Am Rev Respir Dis* 1986; 134: 498–501.
- 185** Bel EH, Zwinderman AH, Timmers MC, Dijkman JH, Sterk PJ. The protective effect of a beta 2 agonist against excessive airway narrowing in response to bronchoconstrictor stimuli in asthma and chronic obstructive lung disease. *Thorax* 1991; 46: 9–14.
- 186** Oosterhoff Y, de Jong JW, Jansen MA, Koeter GH, Postma DS. Airway responsiveness to adenosine 5'-monophosphate in chronic obstructive pulmonary disease is determined by smoking. *Am Rev Respir Dis* 1993; 147: 553–558.
- 187** Kasahara K, Shiba K, Ozawa T, Okuda K, Adachi M. Correlation between the bronchial subepithelial layer and whole airway wall thickness in patients with asthma. *Thorax* 2002; 57: 242–246.
- 188** Benayoun L, Druilhe A, Drombet M, Aubier M, Pretolani M. Airway structural alterations selectively associated with severe asthma. *Am J Respir Crit Care Med* 2003; 167: 1360–1368.
- 189** James AL, Maxwell PS, Pearce-Pinto G, Elliot JG, Carroll NG. The relationship of reticular basement membrane thickness to airway wall remodelling in asthma. *Am J Respir Crit Care Med* 2002; 166: 1590–1595.
- 190** Miranda C, Busacker A, Balzar S, Trudeau J, Wenzel SE. Distinguishing severe asthma phenotypes: role of age at onset and eosinophilic inflammation. *J Allergy Clin Immunol* 2004; 113: 101–118.
- 191** Pulleyn LJ, Newton R, Adcock IM, Barnes PJ. TGFbeta1 allele association with asthma severity. *Hum Genet* 2001; 109: 623–627.
- 192** Szalai C, Kozma GT, Nagy A, *et al.* Polymorphism in the gene regulatory region of MCP-1 is associated with asthma susceptibility and severity. *J Allergy Clin Immunol* 2001; 108: 375–381.
- 193** Israel E, Chinchilli VM, Ford JG, *et al.* Use of regularly scheduled albuterol treatment in asthma: genotype-stratified, randomised, placebo-controlled cross-over trial. *Lancet* 2004; 364: 1505–1512.
- 194** Woolcock AJ, Jenkins CR. Assessment of bronchial responsiveness as a guide to prognosis and therapy in asthma. *Med Clin North Am* 1990; 74: 753–765.
- 195** Sears MR, Greene JM, Willan AR, *et al.* A longitudinal, population-based, cohort study of childhood asthma followed to adulthood. *N Engl J Med* 2003; 349: 1414–1422.
- 196** Rasmussen F, Taylor DR, Flannery EM, *et al.* Risk factors for airway remodelling in asthma manifested by a low postbronchodilator FEV1/vital capacity ratio: a longitudinal population study from childhood to adulthood. *Am J Respir Crit Care Med* 2002; 165: 1480–1488.
- 197** Phelan PD, Robertson CF, Olinsky A. The Melbourne Asthma Study: 1964–1999. *J Allergy Clin Immunol* 2002; 109: 189–194.
- 198** James AL, Palmer LJ, Kicic E, *et al.* Decline in lung function in the Busselton health study: the effects of

- asthma and cigarette smoking. *Am J Respir Crit Care Med* 2005; 171: 109–114.
- 199** Brown PJ, Greville HW, Finucane KE. Asthma and irreversible airflow obstruction. *Thorax* 1984; 39: 131–136.
- 200** Backman KS, Greenberger PA, Patterson R. Airways obstruction in patients with long-term asthma consistent with "irreversible asthma". *Chest* 1997; 112: 1234–1240.
- 201** Orié NGM, Sluiter HJ, de Vries K, Tammelin GJ, Witkop J. The host factor in bronchitis. In: Orié NGM, Sluiter HJ, eds. *Bronchitis*. Assen, Royal van Gorcum, 1961; pp. 43–59.
- 202** Cibella F, Cuttitta G, Bellia V, et al. Lung function decline in bronchial asthma. *Chest* 2002; 122: 1944–1948.
- 203** Cassino C, Berger KI, Goldring RM, et al. Duration of asthma and physiologic outcomes in elderly nonsmokers. *Am J Respir Crit Care Med* 2000; 162: 1423–1428.
- 204** Boulet LP, Turcotte H, Turcot O, Chakir J. Airway inflammation in asthma with incomplete reversibility of airflow obstruction. *Respir Med* 2003; 97: 739–744.
- 205** Zeiger RS, Dawson C, Weiss S. Relationships between duration of asthma and asthma severity among children in the Childhood Asthma Management Program (CAMP). *J Allergy Clin Immunol* 1999; 103: 376–387.
- 206** Bumbacea D, Campbell D, Nguyen L, et al. Parameters associated with persistent airflow obstruction in chronic severe asthma. *Eur Respir J* 2004; 24: 122–128.
- 207** Lange P, Parner J, Vestbo J, Schnohr P, Jensen G. A 15-year follow-up study of ventilatory function in adults with asthma. *N Engl J Med* 1998; 339: 1194–1200.
- 208** Peat JK, Woolcock AJ, Cullen K. Decline of lung function and development of chronic airflow limitation: a longitudinal study of non-smokers and smokers in Busselton, Western Australia. *Thorax* 1990; 45: 32–37.
- 209** Ulrik CS, Lange P. Decline of lung function in adults with bronchial asthma. *Am J Respir Crit Care Med* 1994; 150: 629–634.
- 210** Covar RA, Spahn JD, Murphy JR, Szeffler SJ. Progression of asthma measured by lung function in the childhood asthma management program. *Am J Respir Crit Care Med* 2004; 170: 234–241.
- 211** Jongepier H, Boezen HM, Dijkstra A, et al. Polymorphisms of the ADAM33 gene are associated with accelerated lung function decline in asthma. *Clin Exp Allergy* 2004; 34: 757–760.
- 212** Ulrik CS. Outcome of asthma: longitudinal changes in lung function. *Eur Respir J* 1999; 13: 904–918.
- 213** ten Brinke A, Zwinderman AH, Sterk PJ, Rabe KF, Bel EH. Factors associated with persistent airflow limitation in severe asthma. *Am J Respir Crit Care Med* 2001; 164: 744–748.
- 214** Vonk JM, Jongepier H, Panhuysen CI, Schouten JP, Bleecker ER, Postma DS. Risk factors associated with the presence of irreversible airflow limitation and reduced transfer coefficient in patients with asthma after 26 years of follow up. *Thorax* 2003; 58: 322–327.
- 215** ten Brinke A, Zwinderman AH, Sterk PJ, Rabe KF, Bel EH. "Refractory" eosinophilic airway inflammation in severe asthma: effect of parenteral corticosteroids. *Am J Respir Crit Care Med* 2004; 170: 601–605.
- 216** Wenzel S. Severe asthma in adults. *Am J Respir Crit Care Med* 2005; 172: 149–160.
- 217** Strachan DP, Butland BK, Anderson HR. Incidence and prognosis of asthma and wheezing illness from early childhood to age 33 in a national British cohort. *BMJ* 1996; 312: 1195–1199.
- 218** Dompeling E, van Schayck CP, van Grunsven PM, et al. Slowing the deterioration of asthma and chronic obstructive pulmonary disease observed during bronchodilator therapy by adding inhaled corticosteroids. A 4-year prospective study. *Ann Intern Med* 1993; 118: 770–778.
- 219** Martinez FD. What have we learned from the Tucson Children's Respiratory Study? *Paediatr Respir Rev* 2002; 3: 193–197.
- 220** Palmer LJ, Rye PJ, Gibson NA, Burton PR, Landau LI, LeSouef PN. Airway responsiveness in early infancy predicts asthma, lung function, and respiratory symptoms by school age. *Am J Respir Crit Care Med* 2001; 163: 37–42.
- 221** Payne NR, Rogers AV, Adelroth E, et al. Early thickening of the reticular basement membrane in children with difficult asthma. *Am J Respir Crit Care Med* 2003; 167: 78–82.
- 222** Saglani S, Malmstrom K, Pelkonen AS, et al. Airway remodelling and inflammation in symptomatic infants with reversible airflow obstruction. *Am J Respir Crit Care Med* 2005; 171: 722–727.
- 223** Pohunek P, Warner JO, Turzikova J, Kudrman J, Roche WR. Markers of eosinophilic inflammation and tissue re-modelling in children before clinically diagnosed bronchial asthma. *Pediatr Allergy Immunol* 2005; 16: 43–51.
- 224** Saetta M, Di Stefano A, Maestrelli P, et al. Airway mucosal inflammation in occupational asthma induced by toluene diisocyanate. *Am Rev Respir Dis* 1992; 145: 160–168.
- 225** Karjalainen EM, Laitinen A, Sue-Chu M, Altraja A, Bjermer L, Laitinen LA. Evidence of airway inflammation and remodelling in ski athletes with and without bronchial hyperresponsiveness to methacholine. *Am J Respir Crit Care Med* 2000; 161: 2086–2091.
- 226** Laprise C, Boulet LP. Asymptomatic airway hyperresponsiveness: a three-year follow-up. *Am J Respir Crit Care Med* 1997; 156: 403–409.
- 227** van den Toorn LM, Overbeek SE, de Jongste JC, Leman K, Hoogsteden HC, Prins JB. Airway inflammation is present during clinical remission of atopic asthma. *Am J Respir Crit Care Med* 2001; 164: 2107–2113.
- 228** Bai TR, Cooper J, Koelmeyer T, Pare PD, Weir TD. The effect of age and duration of disease on airway structure in fatal asthma. *Am J Respir Crit Care Med* 2000; 162: 663–669.
- 229** Fletcher C, Peto R, Tinker C, Speizer FE. *The Natural History of Chronic Bronchitis and Emphysema*. Oxford, Oxford University Press; 1976.
- 230** Buist AS, Nagy JM, Sexton GJ. The effect of smoking cessation on pulmonary function: a 30-month follow-up of two smoking cessation clinics. *Am Rev Respir Dis* 1979; 120: 953–957.
- 231** Sandford AJ, Chagani T, Weir TD, Connett JE, Anthonisen NR, Pare PD. Susceptibility genes for rapid decline of lung function in the lung health study. *Am J Respir Crit Care Med* 2001; 163: 469–473.
- 232** Turato G, Di Stefano A, Maestrelli P, et al. Effect of smoking cessation on airway inflammation in chronic bronchitis. *Am J Respir Crit Care Med* 1995; 152: 1262–1267.
- 233** Vestbo J, Prescott E, Lange P. Association of chronic mucus hypersecretion with FEV1 decline and chronic

- obstructive pulmonary disease morbidity. Copenhagen City Heart Study Group. *Am J Respir Crit Care Med* 1996; 153: 1530–1535.
- 234** Hogg JC. Pathophysiology of airflow limitation in chronic obstructive pulmonary disease. *Lancet* 2004; 364: 709–721.
- 235** O'Connor GT, Sparrow D, Weiss ST. A prospective longitudinal study of methacholine airway responsiveness as a predictor of pulmonary-function decline: the Normative Aging Study. *Am J Respir Crit Care Med* 1995; 152: 87–92.
- 236** Tracey M, Villar A, Dow L, Coggon D, Lampe FC, Holgate ST. The influence of increased bronchial responsiveness, atopy, and serum IgE on decline in FEV1. A longitudinal study in the elderly. *Am J Respir Crit Care Med* 1995; 151: 656–662.
- 237** Rijcken B, Schouten JP, Xu X, Rosner B, Weiss ST. Airway hyperresponsiveness to histamine associated with accelerated decline in FEV1. *Am J Respir Crit Care Med* 1995; 151: 1377–1382.
- 238** Fletcher C, Peto R. The natural history of chronic airflow obstruction. *Br Med J* 1977; 1: 1645–1648.
- 239** Donaldson GC, Seemungal TA, Bhowmik A, Wedzicha JA. Relationship between exacerbation frequency and lung function decline in chronic obstructive pulmonary disease. *Thorax* 2002; 57: 847–852.
- 240** Seemungal TA, Donaldson GC, Bhowmik A, Jeffries DJ, Wedzicha JA. Time course and recovery of exacerbations in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2000; 161: 1608–1613.
- 241** Donaldson GC, Seemungal TA, Patel IS, *et al.* Airway and systemic inflammation and decline in lung function in patients with COPD. *Chest* 2005; 128: 1995–2004.
- 242** Wilkinson TM, Patel IS, Wilks M, Donaldson GC, Wedzicha JA. Airway bacterial load and FEV1 decline in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2003; 167: 1090–1095.
- 243** Aaron SD, Angel JB, Lunau M, *et al.* Granulocyte inflammatory markers and airway infection during acute exacerbation of chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 2001; 163: 349–355.
- 244** Jeffery PK, Godfrey RW, Adelroth E, Nelson F, Rogers A, Johansson SA. Effects of treatment on airway inflammation and thickening of basement membrane reticular collagen in asthma. A quantitative light and electron microscopic study. *Am Rev Respir Dis* 1992; 145: 890–899.
- 245** Niimi A, Matsumoto H, Amitani R, *et al.* Effect of short-term treatment with inhaled corticosteroid on airway wall thickening in asthma. *Am J Med* 2004; 116: 725–731.
- 246** Haahtela T, Jarvinen M, Kava T, *et al.* Comparison of a beta 2-agonist, terbutaline, with an inhaled corticosteroid, budesonide, in newly detected asthma. *N Engl J Med* 1991; 325: 388–392.
- 247** Ito K, Ito M, Elliott WM, *et al.* Decreased histone deacetylase activity in chronic obstructive pulmonary disease. *N Engl J Med* 2005; 352: 1967–1976.
- 248** Messer JW, Peters GA, Bennett WA. Causes of death and pathologic findings in 304 cases of bronchial asthma. *Dis Chest* 1960; 38: 616–624.
- 249** Earle BV. Fatal bronchial asthma. A series of fifteen cases with a review of the literature. *Thorax* 1953; 8: 195–206.
- 250** Richards W, Patrick JR. Death from asthma in children. *Am J Dis Child* 1965; 110: 4–21.
- 251** Cardell BS. Death in asthmatics. *Thorax* 1959; 14: 341–352.
- 252** James AL, Hogg JC, Dunn LA, Pare PD. The use of the internal perimeter to compare airway size and to calculate smooth muscle shortening. *Am Rev Respir Dis* 1988; 138: 136–139.
- 253** Masoli M, Fabian D, Holt S, Beasley R. The global burden of asthma: executive summary of the GINA Dissemination Committee report. *Allergy* 2004; 59: 469–478.
- 254** Hessel PA, Mitchell I, Tough S, *et al.* Risk factors for death from asthma. Prairie Provinces Asthma Study Group. *Ann Allergy Asthma Immunol* 1999; 83: 362–368.
- 255** Campbell DA, McLennan G, Coates JR, *et al.* A comparison of asthma deaths and near-fatal asthma attacks in South Australia. *Eur Respir J* 1994; 7: 490–497.
- 256** ten Brinke A, Ouwerkerk ME, Zwinderman AH, Spinhoven P, Bel EH. Psychopathology in patients with severe asthma is associated with increased health care utilization. *Am J Respir Crit Care Med* 2001; 163: 1093–1096.
- 257** Kunitoh H, Yahikozawa H, Kakuta T, *et al.* Fatal and near fatal asthma. *Ann Allergy* 1992; 69: 111–115.
- 258** Abramson MJ, Bailey MJ, Couper FJ, *et al.* Are asthma medications and management related to deaths from asthma? *Am J Respir Crit Care Med* 2001; 163: 12–18.
- 259** Lee P, Abisheganaden J, Chee CB, Wang YT. A new asthma severity index: a predictor of near-fatal asthma? *Eur Respir J* 2001; 18: 272–278.
- 260** Weir TD, Mallek N, Sandford AJ, *et al.* beta2-Adrenergic receptor haplotypes in mild, moderate and fatal/near fatal asthma. *Am J Respir Crit Care Med* 1998; 158: 787–791.
- 261** Synek M, Beasley R, Frew AJ, *et al.* Cellular infiltration of the airways in asthma of varying severity. *Am J Respir Crit Care Med* 1996; 154: 224–230.
- 262** Sobonya RE. Quantitative structural alterations in long-standing allergic asthma. *Am Rev Respir Dis* 1984; 130: 289–292.
- 263** Faul JL, Tormey VJ, Leonard C, *et al.* Lung immunopathology in cases of sudden asthma death. *Eur Respir J* 1997; 10: 301–307.
- 264** O'Sullivan S, Cormican L, Faul JL, *et al.* Activated, cytotoxic CD8(+) T lymphocytes contribute to the pathology of asthma death. *Am J Respir Crit Care Med* 2001; 164: 560–564.
- 265** O'Hollaren MT, Yunginger JW, Offord KP, *et al.* Exposure to an aeroallergen as a possible precipitating factor in respiratory arrest in young patients with asthma. *N Engl J Med* 1991; 324: 359–363.
- 266** Ferrer A, Torres A, Roca J, Sunyer J, Anto JM, Rodriguez-Roisin R. Characteristics of patients with soybean dust-induced acute severe asthma requiring mechanical ventilation. *Eur Respir J* 1990; 3: 429–433.
- 267** Bellomo R, Gigliotti P, Treloar A, *et al.* Two consecutive thunderstorm associated epidemics of asthma in the city of Melbourne. The possible role of rye grass pollen. *Med J Aust* 1992; 156: 834–837.
- 268** Marks GB, Colquhoun JR, Girgis ST, *et al.* Thunderstorm outflows preceding epidemics of asthma during spring and summer. *Thorax* 2001; 56: 468–471.

- 269** Fabbri LM, Danieli D, Crescioli S, *et al.* Fatal asthma in a subject sensitized to toluene diisocyanate. *Am Rev Respir Dis* 1988; 137: 1494–1498.
- 270** Cockcroft DW, Murdock KY, Kirby J, Hargreave F. Prediction of airway responsiveness to allergen from skin sensitivity to allergen and airway responsiveness to histamine. *Am Rev Respir Dis* 1987; 135: 264–267.
- 271** Cockcroft DW, Davis BE, Boulet LP, *et al.* The links between allergen skin test sensitivity, airway responsiveness and airway response to allergen. *Allergy* 2005; 60: 56–59.
- 272** Wiggs B, Moreno R, James A, Hogg JC, Pare P. A model of the mechanics of airway narrowing in asthma. In: Kaliner MA, Barnes PJ, Persson CGA, eds. *Asthma. Its Pathology and Treatment*. New York, Marcel Dekker, 1991; pp. 73–101.
- 273** Azzawi M, Johnston PW, Majumdar S, Kay AB, Jeffery PK. T lymphocytes and activated eosinophils in airway mucosa in fatal asthma and cystic fibrosis. *Am Rev Respir Dis* 1992; 145: 1477–1482.
- 274** Sur S, Crotty TB, Kephart GM, *et al.* Sudden-onset fatal asthma. A distinct entity with few eosinophils and relatively more neutrophils in the airway submucosa? *Am Rev Respir Dis* 1993; 148: 713–717.
- 275** Carroll N, Carello S, Cooke C, James A. Airway structure and inflammatory cells in fatal attacks of asthma. *Eur Respir J* 1996; 9: 709–715.
- 276** James AL, Elliot JG, Abramson MJ, Walters EH. Time to death, airway wall inflammation and remodelling in fatal asthma. *Eur Respir J* 2005; 26: 429–434.
- 277** Murlas CG, Roun JH. Sequence of pathologic changes in the airway mucosa of guinea pigs during ozone-induced bronchial hyperreactivity. *Am Rev Respir Dis* 1985; 131: 314–320.
- 278** McParland BE, Macklem PT, Pare PD. Airway wall remodelling: friend or foe? *J Appl Physiol* 2003; 95: 426–434.
- 279** Pascual RM, Peters SP. Airway remodelling contributes to the progressive loss of lung function in asthma: An overview. *J Allergy Clin Immunol* 2005; 116: 477–486.
- 280** Holgate ST, Davies DE, Lackie PM, Wilson SJ, Puddicombe SM, Lordan JL. Epithelial-mesenchymal interactions in the pathogenesis of asthma. *J Allergy Clin Immunol* 2000; 105: 193–204.
- 281** Wiggs BR, Hrousis CA, Drazen JM, Kamm RD. On the mechanism of mucosal folding in normal and asthmatic airways. *J Appl Physiol* 1997; 83: 1814–1821.
- 282** McParland BE, Wiggs B, Johnson PR, Armour C, Black JL. Airway mucosal folding: is it really a load to smooth muscle shortening? In: American Thoracic Society Annual Scientific Meeting, 1998. Chicago, Illinois, USA. American Thoracic Society, 1998; p. 157.
- 283** Milanese M, Crimi E, Scordamaglia A, *et al.* On the functional consequences of bronchial basement membrane thickening. *J Appl Physiol* 2001; 91: 1035–1040.
- 284** Colebatch HJ, Finucane KE, Smith MM. Pulmonary conductance and elastic recoil relationships in asthma and emphysema. *J Appl Physiol* 1973; 34: 143–153.
- 285** Tiddens HA, Bogaard JM, de Jongste JC, Hop WC, Coxson HO, Pare PD. Physiological and morphological determinants of maximal expiratory flow in chronic obstructive lung disease. *Eur Respir J* 1996; 9: 1785–1794.