



## Early View

### Review

## **Anti-alarmins in asthma: targeting the airway epithelium with next-generation biologics**

Celeste M. Porsbjerg, Asger Sverrild, Clare M. Lloyd, Andrew N. Menzies-Gow, Elisabeth H. Bel

Please cite this article as: Porsbjerg CM, Sverrild A, Lloyd CM, *et al.* Anti-alarmins in asthma: targeting the airway epithelium with next-generation biologics. *Eur Respir J* 2020; in press (<https://doi.org/10.1183/13993003.00260-2020>).

This manuscript has recently been accepted for publication in the *European Respiratory Journal*. It is published here in its accepted form prior to copyediting and typesetting by our production team. After these production processes are complete and the authors have approved the resulting proofs, the article will move to the latest issue of the ERJ online.

# **Anti-alarmins in asthma: targeting the airway epithelium with next-generation biologics**

Celeste M. Porsbjerg<sup>1</sup>, Asger Sverrild<sup>1</sup>, Clare M. Lloyd<sup>2</sup>, Andrew N. Menzies-Gow<sup>3</sup> and Elisabeth H. Bel<sup>4</sup>

Affiliations: <sup>1</sup>Department of Respiratory Medicine, Bispebjerg Hospital, Copenhagen, Denmark. <sup>2</sup>National Heart and Lung Institute, Imperial College London, London, UK. <sup>3</sup>Royal Brompton Hospital, London, UK. <sup>4</sup>Department of Respiratory Medicine, Amsterdam University Medical Centre, University of Amsterdam, Amsterdam, Netherlands.

**Correspondence:** Celeste Porsbjerg MD, Department of Respiratory Medicine, Bispebjerg Hospital, Bispebjerg Bakke 23, 2400 Copenhagen, Denmark.

Email: [Celeste.Porsbjerg@regionh.dk](mailto:Celeste.Porsbjerg@regionh.dk)

Telephone: +45 38 63 53 84

Fax: +45 35 31 21 79

## **Take-home message**

Blocking epithelial alarmins, upstream mediators triggered early in the asthma inflammatory response that orchestrate broad inflammatory effects, is a promising alternative approach to asthma treatment, which may be effective in a broad patient population.

## ABSTRACT

Monoclonal antibody therapies have significantly improved treatment outcomes for patients with severe asthma; however, a significant disease burden remains. Available biologic treatments, including anti-immunoglobulin (Ig) E, anti-interleukin (IL)-5, anti-IL-5R $\alpha$  and anti-IL-4R $\alpha$ , reduce exacerbation rates in study populations by approximately 50% only. Furthermore, there are currently no effective treatments for patients with severe, type 2 (T2)-low asthma. Existing biologics target immunologic pathways that are downstream in the T2 inflammatory cascade, which may explain why exacerbations are only partly abrogated. For example, T2 airway inflammation results from several inflammatory signals in addition to IL-5. Clinically, this can be observed in how fractional exhaled nitric oxide (Fe<sub>NO</sub>), which is driven by IL-13, may remain unchanged during anti-IL-5 treatment despite reduction in eosinophils, and how eosinophils may remain unchanged during anti-IL-4R $\alpha$  treatment despite reduction in Fe<sub>NO</sub>. The broad inflammatory response involving cytokines including IL-4, IL-5 and IL-13 that ultimately results in the classic features of exacerbations (eosinophilic inflammation, mucus production and bronchospasm) is initiated by release of 'alarmins' thymic stromal lymphopoietin (TSLP), IL-33 and IL-25 from the airway epithelium in response to triggers. The central, upstream role of these epithelial cytokines has identified them as strong potential therapeutic targets to prevent exacerbations and improve lung function in patients with T2-high and T2-low asthma. This article describes the effects of alarmins and discusses the potential role of anti-alarmins in the context of existing biologics. Clinical phenotypes of patients who may benefit from these treatments are also discussed, including how biomarkers may help identify potential responders.

**Keywords:** Cytokines; Interleukin-25; Interleukin-33; Thymic stromal lymphopoietin

## Introduction

Asthma is a chronic, inflammatory disease of the airways that affects over 300 million individuals worldwide [1]. Approximately 5–10% of these individuals have severe asthma [2, 3]. Despite a regimen of multiple maintenance medications, patients with severe asthma carry a substantial burden of disease, experiencing frequent exacerbations and having high rates of healthcare resource use, which are associated with substantial costs [4, 5]. These patients also have severely impaired health-related quality of life, with numerous aspects adversely affected, including sleep, work, study, exercise and daily activities. Comorbidities in severe asthma are common and include chronic rhinosinusitis, nasal polyposis and atopic dermatitis [6].

The management of severe asthma is a considerable challenge, particularly in patients with recurrent exacerbations, because these patients are often receiving maximal inhaled therapy and may require regular oral corticosteroid (OCS) treatment [7]. Until relatively recently, there were few alternatives to regular OCS in these patients, but the advent of monoclonal antibody therapies provided physicians with important additional options for exacerbation prevention, potentially enabling tapering or discontinuation of OCS [8]. These biologic therapies target the signalling pathways involved in type 2 (T2) inflammation, mediated by the action of cytokines, such as interleukin (IL)-4, IL-5 and IL-13. These pathways play an important role in the pathogenesis of T2-high asthma, which is characterised by high levels of T2 inflammation, as measured using biomarkers such as blood and sputum eosinophils, and fractional exhaled nitric oxide (F<sub>e</sub>NO) [9]. However, currently available biologic therapies, comprising anti-immunoglobulin (Ig)E, anti-IL-5, anti-IL-5R $\alpha$  and anti-IL-4R $\alpha$  monoclonal antibodies (the latter blocking the IL-4 and IL-13 pathways),

decrease exacerbation rates in study populations by approximately 50% only [10-12]. Furthermore, anti-IgE, anti-IL-5 and anti-IL-4/IL-13 therapies typically produce variable improvements in lung function and symptom scores, although some individuals may experience clinically significant improvements [10-13]. An explanation for this lack of complete efficacy may be that each therapy targets only some of the elements of the pathways that regulate T2 inflammation, leaving other elements of the disease pathophysiology untreated; hence, there is a need for treatments with broader effects on T2 inflammation. Furthermore, we urgently need treatments for patients with T2-low asthma, in whom the disease shows less involvement of T2 pathways [9]. Defining and therefore diagnosing T2-low asthma is made challenging by the lack of reliable biomarkers. Although sputum neutrophilia (in the absence of sputum eosinophilia) may be present in some patients, diagnosis of this asthma subtype is generally inferred from the absence of T2 biomarker elevation [14]. Related to this, it is important to emphasise that the use of corticosteroids, as recommended in patients with severe asthma, reduces eosinophil counts and can therefore confound T2-low classification [15]. For the purposes of this paper, we define T2-low asthma as asthma with low T2 biomarker levels.

## **Inflammatory pathways in asthma**

Within the T2-high subtype, the major pathways involved in airway inflammation (*i.e.* IL-4, IL-13 and IL-5 signalling) are driven by T helper type 2 (Th2) cells and type 2 innate lymphoid cells (ILC2) [16]. IL-4 signalling is central to B cell class switching and triggers the release of IgE from B cells, resulting in airway hyperresponsiveness among other effects [17]. IL-13 is believed to be a central regulator of IgE synthesis, as well as of mucus hypersecretion, airway hyperresponsiveness and fibrosis (an

element of airway remodelling) [18, 19]. IL-5 is a key effector of eosinophilic inflammation [20], which results in airway remodelling [21, 22].

The different T2 pathways have varying prominence across clinical phenotypes, resulting in distinct patterns of inflammatory biomarkers [23]. For instance, patients with early-onset allergic asthma have high serum levels of allergen-specific IgE and total IgE, and elevated Fe<sub>NO</sub> [23, 24], which is characteristic of inflammation predominantly driven by Th2 cells and/or IL-4/IL-13. In contrast, late/adult-onset eosinophilic asthma is a particularly severe form of the disease characterised by higher levels of sputum eosinophils than early-onset asthma [25], as well as elevated blood eosinophils [24, 26], related predominantly to IL-5-mediated inflammation. Elevated Fe<sub>NO</sub>, driven by IL-13, is also characteristic of this phenotype [27]. In the clinic, it may be observed that Fe<sub>NO</sub> remains high in these patients during anti-IL-5 treatment, despite the reduction in eosinophils [28, 29]. Similarly, anti-IL-4R $\alpha$  treatment can reduce Fe<sub>NO</sub>, but transient eosinophilia may be observed [8]; caution should therefore be taken when considering anti-IL-4R $\alpha$  for patients with a history of hypereosinophilic conditions [30]. Alternatively, mixed granulocytic asthma may be present, in which there are elevated levels of both eosinophils and neutrophils [24]. In adults with non-eosinophilic asthma, within the T2-low subtype, there is airway inflammation with a lack of eosinophils [24]. Instead, the dominant inflammatory cell types in blood and sputum may include neutrophils, or there may be very few inflammatory cells, termed paucigranulocytic inflammation [24, 31, 32].

Our understanding of the role of the airway epithelium in driving asthma exacerbations has advanced considerably in recent years, from a simple passive

barrier function to an immunologically active front-line, initiating early responses to external triggers and orchestrating the resulting inflammatory cascade. As the interface between the external environment and the tissue of the bronchi, the airway epithelium is in constant contact with an array of stimuli, such as infectious agents, environmental allergens and atmospheric pollutants. The epithelium has been shown to mediate complex inflammatory processes in response to these allergic and non-allergic triggers, including the release of a trio of epithelial cytokines, known as 'alarmins' [33]. The alarmins thymic stromal lymphopoietin (TSLP), IL-33 and IL-25 instigate inflammatory responses via numerous downstream pathways, including T2 (IL-4, IL-13 and IL-5) (**figure 1**) and others such as T1 or T17 (IL-17), resulting in various pathophysiologic outcomes that may lead to asthma symptoms and exacerbations [33]. It should be noted here that the term 'alarmin' is not limited to these three epithelial cytokines; other proteins produced in response to microbial invasion or tissue injury may also be referred to as alarmins [34].

## **Understanding the pathophysiology of acute exacerbations: a path to better treatments?**

To improve management options for patients with asthma, we require a better understanding of the mechanisms driving acute asthma exacerbations.

Exacerbations are characterised by increasing symptoms, including dyspnoea, wheezing, cough and sputum, owing to increased inflammation and bronchospasm. They may be triggered by exposure to environmental allergens or pollutants but are most often provoked by viral infection (most commonly with rhinovirus) [35-37]. As with the overall patterns of chronic inflammation in patients with asthma described above, the pattern of inflammatory pathways activated during acute exacerbations is

also heterogeneous, with some patients having eosinophilic inflammation and others having neutrophilic, mixed or paucigranulocytic inflammation [38-40]. In patients with eosinophilic exacerbations, inflammation appears to be driven by multiple, simultaneously activated T2 pathways (e.g. both IL-5 and IL-4/IL-13). This can be inferred from human allergen challenge studies that show increases in airway IL-5, IL-4 and IL-13 levels during virus-induced asthma exacerbations [41, 42], as well as proteomic data showing elevated sputum levels of both IL-5 and IL-13 in patients with eosinophilic asthma [43]. With regard to the considerable proportion of patients who have non-eosinophilic, neutrophilic inflammation in relation to an exacerbation [24], less is understood about the pathways involved in non-eosinophilic exacerbations, although they may involve IL-17 and IL-23 [44, 45]. There is also evidence of airway microbiome involvement because subclinical infection by particular bacterial species is associated with neutrophilic, steroid-resistant asthma [46, 47].

Potential associations between an inflammatory asthma phenotype and specific exacerbation triggers are also poorly understood. Virus-induced exacerbations are more common in patients with allergic asthma [35], as well as in patients with elevated  $F_{eNO}$  or blood eosinophils [48], suggesting a link between virus-induced exacerbations and T2 inflammation. Eosinophils, activated by IL-5, drive pathophysiology in exacerbating and stable eosinophilic asthma. Eosinophils have recently been found to play a role in the defence against viruses. However, in patients with asthma, the degree to which eosinophils can carry out this role correlates with disease severity [49], explaining why patients with asthma and high eosinophil counts still experience virus-induced exacerbations. This may also explain



the lack of efficacy of anti-IL-5 therapy in mild asthma. For example, in a virus challenge study in patients with mild asthma, anti-IL-5 altered elements of the immune response, including B lymphocytes, macrophages and neutrophils, as well as attenuated eosinophils, but there were no associated improvements in lung function [50]. Less is known about potential links between asthma phenotypes and other associated exacerbation triggers.

There is a pressing need for better understanding of the drivers of exacerbations. One route towards achieving this is to study the impact of specific biologics on patient exacerbations with respect to their phenotypes and triggers. A retrospective analysis of exacerbations in a trial in patients with severe eosinophilic asthma receiving anti-IL-5 treatment with mepolizumab found that exacerbations occurring during treatment were associated with a lower induced sputum eosinophil count and less wheezing than those occurring in patients receiving placebo. This was particularly apparent in patients who had not started rescue treatment with corticosteroids, suggesting that exacerbations were less severe with mepolizumab [51]. This finding is corroborated by previous observations that non-eosinophilic exacerbations are less severe than eosinophilic exacerbations [52]. Data from ongoing studies designed to explore the nature of exacerbations in patients being treated with biologics, such as anti-IL-5 monoclonal antibodies [53], are keenly anticipated. With our current limited knowledge, without full appreciation of the heterogeneity of inflammation in acute exacerbations, prescription of treatments targeting specific inflammatory pathways may not effectively prevent all exacerbations. Therefore, targeting multiple inflammatory pathways may be a more effective approach.

## **Epithelial alarmins: early orchestrators of airway inflammation in acute asthma**

### ***Role of TSLP and IL-33 in T2-high asthma***

There is substantial evidence that the alarmins, TSLP and IL-33, play key roles in driving T2 inflammation in asthma. TSLP levels in bronchial lavage fluid and biopsies are elevated in patients with asthma compared with healthy individuals, and correlate with disease severity (including a negative correlation with lung function defined by forced expiratory volume in 1 second [FEV<sub>1</sub>]) [54-57]. In addition, genetic studies of TSLP have identified alleles that are associated with asthma [58]. Similarly, expression levels of IL-33 and its receptor ST2 in serum and bronchial biopsy samples are higher in patients with asthma than in healthy individuals, and positively correlate with disease severity [59-61]. Additionally, patients with allergic and eosinophilic asthma phenotypes have higher serum levels of IL-33 than those with non-allergic and non-eosinophilic phenotypes [62]. Furthermore, in patients with allergic asthma, ST2 expression on eosinophils from blood and sputum is significantly upregulated after allergen inhalation challenge [54]. Higher serum soluble ST2 levels are associated with an increased risk of exacerbations [63]. Certain alleles of IL-33 and its receptor are associated with asthma [58], and a rare IL-33 loss-of-function mutation reduces blood eosinophil counts and protects from asthma [64]. Bronchial allergen challenge directly increases airway expression of all three alarmins (TSLP, IL-33 and IL-25) in patients with allergic asthma, to a degree correlating with the degree of airway obstruction [42].

TSLP has been shown to drive various elements of asthma pathophysiology, including airway hyperresponsiveness, mucus overproduction and airway remodelling, via effects triggered downstream. However, it should be noted that not all these elements have been demonstrated conclusively in humans. During T2 inflammatory responses, TSLP potently activates dendritic cells and induces production of the Th2-attracting chemokines thymus and activation-regulated chemokine (TARC; also known as CCL17) and macrophage-derived chemokine (CCL22). TSLP-activated dendritic cells prime naïve Th cells to produce various inflammatory mediators, including IL-4, IL-5 and IL-13 [65]. The IL-5 secreted from polarised Th2 cells mediates eosinophilic inflammation via effects on eosinophil recruitment, maturation and survival [66]. When activated, eosinophils release cysteinyl leukotrienes, which are potent bronchoconstrictors [22]. They also induce airway remodelling through airway smooth muscle cell proliferation [21] and the release of mediators, such as transforming growth factor- $\beta$ , cationic proteins and cytokines, as well as through interactions with mast cells and epithelial cells [22]. Mast cells also release acute-phase inflammatory mediators, including cysteinyl leukotrienes, histamine and prostaglandins [67]. IL-4 and IL-13 secreted from polarised Th2 cells activate B cells, which produce IgE, resulting in airway hyperresponsiveness [18] and triggering mast cell degranulation, leading to vascular permeability [68]. IL-13 also increases mucus production via goblet cell proliferation, and airway inflammation via increased epithelial production of FeNO [19]. In addition to Th2 cells, TSLP activates multiple other cell types that release IL-13, including mast cells, basophils and ILC2s [69-71].

Meanwhile, IL-33 is thought to act as a positive regulator of TSLP dendritic cell signalling, initiating and maintaining Th2 cell-mediated inflammatory responses [72]. IL-33 drives airway hyperresponsiveness through IL-13-mediated mast cell–airway smooth muscle crosstalk. Human lung mast cells are stimulated by IL-33 to release histamine and IL-13, resulting in airway constriction [60]. Further roles of IL-33 include promoting mast cell survival and cytokine production [73], eosinophil survival [74], basophil activation, survival and proliferation [71], and increasing IL-13 release from ILC2s [75].

### ***Role of IL-25 in T2-high asthma***

Of the three alarmins, the actions of IL-25 in patients with asthma are probably the least understood. IL-25 appears to play a key role in allergic inflammation, particularly as a driver of the T2 inflammatory response during virus-induced asthma exacerbations [76]. Elevated plasma levels of IL-25 are associated with the allergic asthma phenotype [77], and IL-25 concentration in sputum correlates with disease severity [78]. Sputum IL-25 is also increased in atopic versus non-atopic asthma patients [78]. In addition, a rare allele of a component of the IL-25 receptor, IL-17RB, is associated with a reduced incidence of asthma [79].

After inhaled allergen exposure, there is increased expression of IL-25 and its receptor in the bronchial mucosa and dermis (collected by skin biopsy) [80], and in circulating eosinophils and airway dendritic cells [81, 82] in patients with allergic asthma. Bronchial epithelial expression of IL-25, but not IL-33 or TSLP, is heterogeneous in patients with asthma. Those expressing high levels of IL-25 have greater airway hyperresponsiveness and remodelling, increased blood and airway

eosinophilia, and higher allergen skin-test reactivity, with elevated IgE, than those expressing low levels of IL-25 [83]. Aside from epithelial cells, which form the principal location of all three alarmins, IL-25 release appears to be prominently localised to eosinophils [42]. IL-25 is hypothesised to play a role in allergen-induced trafficking of eosinophil-lineage committed progenitor cells to the airways, and local differentiation, promoting tissue eosinophilia during asthmatic responses [84].

### ***Role of TSLP in T2-low asthma***

Much less is known about the role of alarmins in T2-low asthma than in T2-high asthma; most of our understanding comes from animal studies, with relatively few studies having been conducted in humans or human cells. Of the three alarmins, TSLP has the most compelling evidence for an effect in T2-low asthma. Anti-TSLP treatment has been shown to reduce exacerbations in patients with severe, uncontrolled asthma and low levels of blood eosinophils and  $F_{eNO}$  (as discussed later) [85]. TSLP is thought to play a role in neutrophilic, T2-low airway inflammation by activating dendritic cells to induce polarization of naïve T cells towards a Th17 phenotype [86]. Th17 polarization and subsequent IL-17 release promotes neutrophilic inflammation and a non-allergic/non-eosinophilic response [87]. TSLP, but not IL-33 or IL-25, expression in human bronchoalveolar lavage fluid has been shown to correlate most closely with neutrophil infiltration [54]. Nevertheless, future studies are warranted to understand whether there is a broader role for IL-33 in the pathogenesis of T2-low asthma.

### ***Role of the alarmins in steroid-refractory asthma***

Airway levels of ILC2s are elevated in several phenotypes of asthma, including late-onset eosinophilic asthma [25, 88-90], and may be the key driver of eosinophilic inflammation in these patients. Although individual alarmins do not directly promote proliferation of ILC2s, TSLP promotes the longevity of ILC2s, while ILC2 activation is promoted by IL-33, especially when in combination with TSLP (or IL-2) [70]. In a recent, *in vitro* study of human blood and lung ILC2s from patients with asthma and healthy controls, IL-25- and IL-33-stimulated expression of T2 cytokines by blood ILC2s was inhibited by steroid treatment; however, TSLP-stimulated T2 cytokine expression was not [91]. TSLP levels were elevated in bronchoalveolar lavage fluid in patients with asthma versus healthy controls and correlated with the level of steroid resistance. TSLP was shown to induce steroid resistance in the ILC2s [91]. TSLP-initiated T2 signalling may therefore be particularly important in severe asthma patients with steroid-refractory disease and both Th2- and ILC2-driven inflammation. Non-eosinophilic asthma is also associated with steroid resistance [92], linked to high IL-17 and high numbers of neutrophils in these patients, potentially resulting from TSLP signalling (as described above) [44, 87].

## **Targeting the airway alarmins with biologic therapies**

The inability of current biologic therapies to prevent all asthma exacerbations and the complexity of the inflammatory pathways involved in asthma, provide a strong rationale for finding alternative, more effective means of reducing airway inflammation and thereby preventing exacerbations. The central, upstream role of alarmins makes them attractive potential therapeutic targets in this regard. Current biologics are not disease modifying, and available evidence suggests that asthma severity returns to pre-treatment levels after stopping treatment with them [93, 94].

Blocking alarmins, however, has the potential to inhibit airway hyperresponsiveness and remodelling [95] and produce sustained reductions in disease activity. Biologics targeting the alarmins are now in development, with anti-TSLP and anti-IL-33 therapies having entered clinical trials. The current evidence for the effects of alarmin blockade in humans is summarised in **table 1** and **figure 2**.

### ***Anti-TSLP***

Tezepelumab, an anti-TSLP human monoclonal antibody that prevents TSLP interacting with its receptor complex [96], is the anti-alarmin that has reached the furthest stage of clinical development (**table 1**). In a proof-of-concept allergen challenge study in patients with mild allergic asthma, tezepelumab 700 mg was administered every 4 weeks for 12 weeks, with an inhaled allergen challenge at baseline and after 6 and 12 weeks [96]. FEV<sub>1</sub> reductions post-allergen challenge at week 6 and week 12 were significantly smaller with tezepelumab than with placebo (34% and 46%, respectively). In addition, patients receiving tezepelumab had significant decreases in levels of blood and sputum eosinophils before and after allergen challenge. Similarly, Fe<sub>NO</sub> levels dropped throughout the study, and post-allergen Fe<sub>NO</sub> increases were significantly reduced. These results paved the way for a phase 2b study in patients with moderate-to-severe uncontrolled asthma, who were administered tezepelumab 70 mg every 4 weeks, 210 mg every 4 weeks or 280 mg every 2 weeks, or placebo every 2 weeks, over 52 weeks [85]. Tezepelumab was well tolerated and significantly reduced annualised asthma exacerbation rates in the tezepelumab groups versus placebo (by 61%, 71% and 66%, respectively). Similar results were observed regardless of patients' blood eosinophil counts at enrolment. Pre-bronchodilator FEV<sub>1</sub> at week 52 improved within the first month of tezepelumab treatment relative to placebo and persisted throughout the study, as did

improvements in patient-reported outcomes. In an exploratory analysis of the phase 2b data, tezepelumab was found to reduce annualised asthma exacerbation rates irrespective of patients' baseline blood eosinophil count, level of Fe<sub>NO</sub>, or serum levels of IgE, IL-5, IL-13, periostin or TARC [97]. In addition, levels of these proinflammatory biomarkers and cytokines were reduced as early as 4 weeks after treatment initiation; the reductions were then maintained throughout the 52-week treatment period.

Taken together, these findings indicate that tezepelumab has broad inhibitory effects on multiple T2 inflammatory mediators of asthma, suggesting that anti-TSLP treatment may be beneficial in patients with a range of inflammatory phenotypes (**table 2**). As described above, because TSLP is postulated to initiate T2 inflammation via both Th2 and ILC2 cells, blocking TSLP with tezepelumab may effectively reduce inflammation in patients with steroid-refractory asthma.

Furthermore, the observation that tezepelumab was efficacious in patients with low blood eosinophils and low Fe<sub>NO</sub> indicates that it may be effective in the T2-low asthma subtype. By contrast, the existing biologics targeting T2 pathways, such as anti-IL-4R and anti-IL-5, have shown less efficacy in patients with low versus high eosinophil counts [98-100]. Whether tezepelumab is effective in patients with T2-low asthma is yet to be confirmed and will depend on how T2-low asthma is defined in terms of biomarkers in future clinical studies.

Several clinical investigations assessing the efficacy and safety of tezepelumab in different patient populations are ongoing, with results yet to be published (**table 3**).

Two phase 2 (NCT02698501 and NCT03688074) and two phase 3 studies are



ongoing: one investigating the efficacy and safety of tezepelumab in reducing OCS use in adults with OCS-dependent asthma (NCT03406078), and one investigating the efficacy and safety of tezepelumab in adults and adolescents with severe, uncontrolled asthma (NCT03347279).

In addition to tezepelumab, CSJ117, an anti-TSLP monoclonal antibody fragment delivered by inhalation, is in phase 1 development (NCT03138811), with results not yet published.

### ***Anti-IL-33***

Monoclonal antibodies targeting IL-33 or ST2 are in clinical development, with several in phase 2 trials (**table 3**), but results from these studies have not yet been published in peer-reviewed journals. AMG282 (RG6149/MSTT1041A) is being evaluated in a 12-month phase 2b study in patients with uncontrolled, severe asthma, with the primary outcome being the rate of exacerbations (NCT02918019). REGN3500 (SAR440340) was recently trialled alone and in combination with dupilumab in a 12-week phase 2a study in patients with moderate-to-severe asthma (NCT03387852). Top-line findings from this study are available but are yet to be published in a peer-reviewed journal. Although the anti-IL-33 antibody met its primary endpoint of reducing the incidence of 'loss of asthma control' events compared with placebo, outcomes were no better than those seen with dupilumab monotherapy. In addition, combined REGN3500 and dupilumab did not demonstrate increased benefit compared with dupilumab alone [101]. Two earlier phase 1 studies of REGN3500 have also been completed (NCT03112577, NCT02999711). Etokimab (ANB020) has recently been evaluated in a proof-of-concept phase 2a study in

adults with severe eosinophilic asthma, including a primary outcome of change in blood eosinophil count (NCT03469934). GSK3772847 (CNT07160) has recently completed a phase 2a study in patients with moderately severe asthma, with the primary outcome being loss of asthma control events at week 16 (NCT03207243). A 28-week phase 2b study in patients with moderate-to-severe asthma with allergic fungal airway disease is ongoing, with primary outcomes of change from baseline in blood eosinophil count and Fe<sub>NO</sub> levels at week 13 (NCT03393806).

### ***Anti-IL-25***

Although IL-25 appears to play a key role in allergic inflammation, particularly during virus-induced asthma exacerbations (as described above), no clinical studies of anti-IL-25 antibodies are in progress.

### ***Patients who may benefit from anti-alarmin treatment***

From a clinical perspective, there are several groups of patients who may benefit from the potentially broad anti-inflammatory effects of anti-alarmins, such as those with multiple activated T2 pathways. For example, in patients with allergic eosinophilic asthma, it may be difficult to discern whether exacerbations are primarily driven by allergen exposure or are related to eosinophilic inflammation. The ability to target pathways driving both IgE and eosinophils with a single treatment may be a more attractive option than anti-IgE or anti-IL-5. A second example is those patients with non-allergic eosinophilic asthma who have very high Fe<sub>NO</sub>, indicating concomitant activation of both IL-5 and IL-13, perhaps through activation of ILC2 cells. Although there is presently no evidence available to inform us whether these patients could achieve sufficient control on either anti-IL-5 or anti-IL-4/13, it appears

plausible that some need a broader suppression of the T2 pathways. Thirdly, patients with severe eosinophilic asthma and comorbidities, such as nasal polyposis or atopic dermatitis, that do not improve with current biologic therapies may also have co-activation of cells releasing both IL-5 and IL-4/IL-13. This suggests that these patients may benefit from an upstream-targeted treatment. Other patients who may benefit from anti-alarmin therapy could include those with predominantly mast cell-driven disease or those with no clear pattern of T2 inflammation.

### ***Outstanding questions regarding anti-alarmins***

Although tezepelumab has thus far been found to be well tolerated in clinical studies, questions remain regarding the long-term safety of blocking the alarmins and the benefit–risk of anti-alarmin treatment compared with that of cumulative OCS exposure, for which the side effects are well-known and can lead to significant morbidity [102]. One area for potential exploration is the antimicrobial activity of TSLP, in particular the short form of TSLP (sfTSLP), which has been shown to exert potent antimicrobial activity [103]. It is therefore conceivable that blocking sfTSLP could result in a greater risk of infection.

Upregulation of alarmins has been implicated in a number of human cancers, although their role is controversial; in some neoplasms, TSLP appears to play a pro-tumorigenic role, while in others, TSLP seems to be protective [104]. As such, further studies are required to determine the precise role of TSLP in human cancer.

Similarly, further studies are warranted to determine the effect of blocking IL-33 on cardiovascular disease, because IL-33 is thought to be cardioprotective [105].

An additional and important question is whether suppressing one alarmin alone is sufficient to suppress all inflammatory pathways induced by the alarmins. In this context, it is clearly important to understand the potential interactions between the alarmins and the relative importance of each alarmin in inflammatory responses. Using chronic models of helminth infection and T2 cytokine-driven lung inflammation, targeting all three alarmins (TSLP, IL-25 and IL-33) was found to be more efficacious than blocking a single alarmin alone [106]. Furthermore, disruption of all three mediators in a model of chronic house dust mite-induced allergic lung inflammation resulted in reduced inflammation, mucus production and lung remodelling. The authors concluded that the results of these studies suggest redundant roles for TSLP, IL-25 and IL-33 in the maintenance of T2 pathology and that these alarmins interact to potentiate their individual effects. An additional study found that both IL-33 and TSLP were required for epithelial cell IL-25 expression, mucus metaplasia and ILC2 expansion after early-life rhinovirus infection in mice [107]. The authors concluded that generation of mucus metaplasia involves a complex interplay among IL-25, IL-33 and TSLP. Nevertheless, blocking the TSLP pathway alone appears sufficient to suppress inflammation, as shown so far in the tezepelumab studies, albeit other anti-TSLP biologics such as RG7258 were not successfully developed into asthma therapies [108]. To date, no clinical studies have investigated the potential interactions between alarmins in patients with asthma, but it would be of importance to understand such interactions, both in terms of effects on the efficacy and safety of these treatments.

Of further interest is the mechanism by which anti-alarmins prevent exacerbations. It has been postulated that omalizumab exerts its beneficial effects through restoration

of innate antiviral immunity in plasmacytoid dendritic cells and through increasing the release of interferon- $\alpha$  on rhinovirus exposure, which is deficient in the presence of high IgE [109]. It is unknown whether anti-alarmins have an effect on antiviral immunity, and it could be speculated that such a mechanism may result in treatment benefit for patients with both T2-high and T2-low asthma. Mechanistic studies are required to determine how these new treatments work.

## **Conclusions and future perspective**

Monoclonal antibody therapies represent a new treatment era in severe asthma, offering patients who are exacerbating despite high-dose anti-inflammatory treatment a safer and more effective alternative option. However, because blocking specific T2 pathways with currently available biologics fails to prevent all exacerbations, a significant burden of disease remains. This limitation of current biologics may be a consequence of blocking pathways downstream in the immunological cascade, leaving others still active, as the partial suppression of T2 pathways may be insufficient to abrogate exacerbations in some patients. Blocking alarmins, upstream mediators triggered early in the inflammatory response that orchestrate broad T2 inflammatory effects, is a promising alternative approach that may be effective in a broader patient population.

Further research into the role of alarmins in asthma pathogenesis will improve our understanding of disease severity and the heterogeneity in response to current treatments. This research should include studies of differences between the roles of the alarmins in the different clinical phenotypes of severe asthma, such as late-onset eosinophilic asthma versus early-onset allergic asthma, as well as their relationship

with comorbidities such as atopic dermatitis and nasal polyposis. Understanding the clinical effect of each alarmin in patients with specific phenotypes, and in patients with a combination of asthma, atopic dermatitis and nasal polyposis, will be crucial not only for informing our ability to correctly select the appropriate anti-alarmin as a treatment, but also to our understanding of disease mechanisms in asthma. In addition to this, more studies in which exacerbations are phenotyped are also required to help to identify which types of exacerbation are reduced or otherwise by a given biologic treatment. Finally, mechanistic studies in humans are required to improve our understanding of how next-generation biologics work. This will help clinicians to select the most appropriate treatments for their patients.

## **Acknowledgements**

The authors thank Richard Claes, PhD of PharmaGenesis London, London, UK for providing medical writing support, which has been funded by AstraZeneca, Cambridge, UK, in accordance with Good Publication Practice 3 (GPP3) guidelines (<http://www.ismpp.org/gpp3>).

## **Conflict of interest**

CMP has received unrestricted research grants and honoraria for scientific presentations and consultancies from AstraZeneca, Boehringer Ingelheim, GlaxoSmithKline, Icepharma, Janssen, Nigaard, Nopharma, Novartis, Pharmaxis, Roche, Sandoz, Sanofi and Teva. AS has attended advisory board for AstraZeneca and Sanofi Genzyme and has received speaker fees from AstraZeneca and Novartis. CML is a Wellcome Senior Fellow in Basic Biomedical Science

(107059/15/Z). AMG has attended advisory boards for AstraZeneca, Boehringer Ingelheim, GlaxoSmithKline, Novartis, Sanofi and Teva, and has received speaker fees from AstraZeneca, Boehringer Ingelheim, Novartis, Roche, Teva and Vectura. He has participated in research with AstraZeneca for which his institution has been remunerated and has attended international conferences with Teva. He has had consultancy agreements with AstraZeneca, Sanofi and Vectura. EB has received personal fees from AstraZeneca, Chiesi, GlaxoSmithKline, Novartis, Sanofi, Sterna and Teva.

## References

1. Global Asthma Network. Global Asthma Network. Global Asthma Report 2018. <http://www.globalasthmareport.org/> Date last updated: January 2018. Date last accessed: January 17, 2020.
2. Backman H, Jansson SA, Stridsman C, Eriksson B, Hedman L, Eklund BM, Sandstrom T, Lindberg A, Lundback B, Ronmark E. Severe asthma-A population study perspective. *Clin Exp Allergy* 2019; 49: 819–828.
3. Chung KF, Wenzel SE, Brozek JL, Bush A, Castro M, Sterk PJ, Adcock IM, Bateman ED, Bel EH, Bleecker ER, Boulet LP, Brightling C, Chanez P, Dahlen SE, Djukanovic R, Frey U, Gaga M, Gibson P, Hamid Q, Jajour NN, Mauad T, Sorkness RL, Teague WG. International ERS/ATS guidelines on definition, evaluation and treatment of severe asthma. *Eur Respir J* 2014; 43: 343–373.
4. Chipps BE, Zeiger RS, Borish L, Wenzel SE, Yegin A, Hayden ML, Miller DP, Bleecker ER, Simons FE, Szeffler SJ, Weiss ST, Haselkorn T, Group TS. Key findings and clinical implications from The Epidemiology and Natural History of Asthma: Outcomes and Treatment Regimens (TENOR) study. *J Allergy Clin Immunol* 2012; 130: 332–342.
5. Zeiger RS, Schatz M, Dalal AA, Qian L, Chen W, Ngor EW, Suruki RY, Kawatkar AA. Utilization and costs of severe uncontrolled asthma in a managed-care setting. *J Allergy Clin Immunol Pract* 2016; 4: 120–129.
6. Boulet LP. Influence of comorbid conditions on asthma. *Eur Respir J* 2009; 33: 897–906.
7. Ramsahai JM, Hansbro PM, Wark PAB. Mechanisms and management of asthma exacerbations. *Am J Respir Crit Care Med* 2019; 199: 423–432.
8. McGregor MC, Krings JG, Nair P, Castro M. Role of biologics in asthma. *Am J Respir Crit Care Med* 2019; 199: 433–445.
9. Fahy JV. Type 2 inflammation in asthma--present in most, absent in many. *Nat Rev Immunol* 2015; 15: 57–65.

10. Normansell R, Walker S, Milan SJ, Walters EH, Nair P. Omalizumab for asthma in adults and children. *Cochrane Database Syst Rev* 2014: CD003559.
11. Farne HA, Wilson A, Powell C, Bax L, Milan SJ. Anti-IL5 therapies for asthma. *Cochrane Database Syst Rev* 2017: 9: CD010834.
12. Zayed Y, Kheiri B, Banifadel M, Hicks M, Aburahma A, Hamid K, Bachuwa G, Chandran A. Dupilumab safety and efficacy in uncontrolled asthma: a systematic review and meta-analysis of randomized clinical trials. *J Asthma* 2018: 1–10.
13. Xiong XF, Zhu M, Wu HX, Fan LL, Cheng DY. Efficacy and safety of dupilumab for the treatment of uncontrolled asthma: a meta-analysis of randomized clinical trials. *Respir Res* 2019: 20: 108.
14. Fricker M, Heaney LG, Upham JW. Can biomarkers help us hit targets in difficult-to-treat asthma? *Respirology* 2017: 22: 430–442.
15. Cowan DC, Cowan JO, Palmay R, Williamson A, Taylor DR. Effects of steroid therapy on inflammatory cell subtypes in asthma. *Thorax* 2010: 65: 384–390.
16. Gurram RK, Zhu J. Orchestration between ILC2s and Th2 cells in shaping type 2 immune responses. *Cell Mol Immunol* 2019: 16: 225–235.
17. Steinke JW, Borish L. Th2 cytokines and asthma. Interleukin-4: its role in the pathogenesis of asthma, and targeting it for asthma treatment with interleukin-4 receptor antagonists. *Respir Res* 2001: 2: 66–70.
18. Rael EL, Lockey RF. Interleukin-13 signaling and its role in asthma. *World Allergy Organ J* 2011: 4: 54–64.
19. Corren J. Role of interleukin-13 in asthma. *Curr Allergy Asthma Rep* 2013: 13: 415–420.
20. Garcia G, Taille C, Laveneziana P, Bourdin A, Chanez P, Humbert M. Anti-interleukin-5 therapy in severe asthma. *Eur Respir Rev* 2013: 22: 251–257.
21. Halwani R, Vazquez-Tello A, Sumi Y, Pureza MA, Bahammam A, Al-Jahdali H, Soussi-Gounni A, Mahboub B, Al-Muhsen S, Hamid Q. Eosinophils induce airway smooth muscle cell proliferation. *J Clin Immunol* 2013: 33: 595–604.
22. Possa SS, Leick EA, Prado CM, Martins MA, Tiberio IF. Eosinophilic inflammation in allergic asthma. *Front Pharmacol* 2013: 4: 46.
23. Kim H, Ellis AK, Fischer D, Noseworthy M, Olivenstein R, Chapman KR, Lee J. Asthma biomarkers in the age of biologics. *Allergy Asthma Clin Immunol* 2017: 13: 48.
24. Carr TF, Zeki AA, Kraft M. Eosinophilic and noneosinophilic asthma. *Am J Respir Crit Care Med* 2018: 197: 22–37.
25. Miranda C, Busacker A, Balzar S, Trudeau J, Wenzel SE. Distinguishing severe asthma phenotypes: role of age at onset and eosinophilic inflammation. *J Allergy Clin Immunol* 2004: 113: 101–108.
26. de Groot JC, Storm H, Amelink M, de Nijs SB, Eichhorn E, Reitsma BH, Bel EH, Ten Brinke A. Clinical profile of patients with adult-onset eosinophilic asthma. *ERJ Open Res* 2016: 2.
27. Hirano T, Matsunaga K. Late-onset asthma: current perspectives. *J Asthma Allergy* 2018: 11: 19–27.



28. Yancey SW, Keene ON, Albers FC, Ortega H, Bates S, Bleecker ER, Pavord I. Biomarkers for severe eosinophilic asthma. *J Allergy Clin Immunol* 2017; 140: 1509–1518.
29. Berry A, Busse WW. Biomarkers in asthmatic patients: Has their time come to direct treatment? *J Allergy Clin Immunol* 2016; 137: 1317–1324.
30. Brooks GD. Updated evaluation of dupilumab in the treatment of asthma: patient selection and reported outcomes. *Ther Clin Risk Manag* 2020; 16: 181–187.
31. Tliba O, Panettieri RA, Jr. Paucigranulocytic asthma: Uncoupling of airway obstruction from inflammation. *J Allergy Clin Immunol* 2019; 143: 1287–1294.
32. Moore WC, Hastie AT, Li X, Li H, Busse WW, Jarjour NN, Wenzel SE, Peters SP, Meyers DA, Bleecker ER, National Heart L, Blood Institute's Severe Asthma Research P. Sputum neutrophil counts are associated with more severe asthma phenotypes using cluster analysis. *J Allergy Clin Immunol* 2014; 133: 1557–1563.
33. Mitchell PD, O'Byrne PM. Epithelial-derived cytokines in asthma. *Chest* 2017; 151: 1338–1344.
34. Oppenheim JJ, Tewary P, de la Rosa G, Yang D. Alarmins initiate host defense. *Adv Exp Med Biol* 2007; 601: 185–194.
35. Murray CS, Simpson A, Custovic A. Allergens, viruses, and asthma exacerbations. *Proc Am Thorac Soc* 2004; 1: 99–104.
36. Heymann PW, Carper HT, Murphy DD, Platts-Mills TA, Patrie J, McLaughlin AP, Erwin EA, Shaker MS, Hellems M, Peerzada J, Hayden FG, Hatley TK, Chamberlain R. Viral infections in relation to age, atopy, and season of admission among children hospitalized for wheezing. *J Allergy Clin Immunol* 2004; 114: 239–247.
37. Nicholson KG, Kent J, Ireland DC. Respiratory viruses and exacerbations of asthma in adults. *BMJ* 1993; 307: 982–986.
38. Wark PA, Gibson PG. Asthma exacerbations. 3: Pathogenesis. *Thorax* 2006; 61: 909–915.
39. Fahy JV, Kim KW, Liu J, Boushey HA. Prominent neutrophilic inflammation in sputum from subjects with asthma exacerbation. *J Allergy Clin Immunol* 1995; 95: 843–852.
40. Wang F, He XY, Baines KJ, Gunawardhana LP, Simpson JL, Li F, Gibson PG. Different inflammatory phenotypes in adults and children with acute asthma. *Eur Respir J* 2011; 38: 567–574.
41. Jackson DJ, Makrinioti H, Rana BM, Shamji BW, Trujillo-Torrallbo MB, Footitt J, Jerico D-R, Telcian AG, Nikonova A, Zhu J, Aniscenko J, Gogsadze L, Bakhsoliani E, Traub S, Dhariwal J, Porter J, Hunt D, Hunt T, Hunt T, Stanciu LA, Khaitov M, Bartlett NW, Edwards MR, Kon OM, Mallia P, Papadopoulos NG, Akdis CA, Westwick J, Edwards MJ, Cousins DJ, Walton RP, Johnston SL. IL-33-dependent type 2 inflammation during rhinovirus-induced asthma exacerbations in vivo. *Am J Respir Crit Care Med* 2014; 190: 1373–1382.
42. Wang W, Li Y, Lv Z, Chen Y, Li Y, Huang K, Corrigan CJ, Ying S. Bronchial allergen challenge of patients with atopic asthma triggers an alarmin (IL-33, TSLP, and IL-25) response in the airways epithelium and submucosa. *J Immunol* 2018; 201: 2221–2231.
43. Schofield JPR, Burg D, Nicholas B, Strazzeri F, Brandsma J, Staykova D, Folisi C, Bansal AT, Xian Y, Guo Y, Rowe A, Corfield J, Wilson S, Ward J, Lutter R, Shaw DE, Bakke PS, Caruso M, Dahlen SE, Fowler SJ, Horvath I, Howarth P, Krug N, Montuschi P, Sanak M, Sandstrom T, Sun K, Pandis I, Riley J, Auffray C, De Meulder B, Lefaudeux D, Sousa AR, Adcock IM, Chung KF, Sterk PJ, Skipp PJ, Djukanovic R, Group UBS. Stratification of asthma phenotypes by airway proteomic signatures. *J Allergy Clin Immunol* 2019; 144: 70–82.

44. Chesne J, Braza F, Mahay G, Brouard S, Aronica M, Magnan A. IL-17 in severe asthma. Where do we stand? *Am J Respir Crit Care Med* 2014; 190: 1094–1101.
45. Nakajima H, Hirose K. Role of IL-23 and Th17 cells in airway inflammation in asthma. *Immune Netw* 2010; 10: 1–4.
46. Goleva E, Jackson LP, Harris JK, Robertson CE, Sutherland ER, Hall CF, Good JT, Jr., Gelfand EW, Martin RJ, Leung DY. The effects of airway microbiome on corticosteroid responsiveness in asthma. *Am J Respir Crit Care Med* 2013; 188: 1193–1201.
47. Earl CS, An SQ, Ryan RP. The changing face of asthma and its relation with microbes. *Trends Microbiol* 2015; 23: 408–418.
48. Bjerregaard A, Laing IA, Backer V, Sverrild A, Khoo SK, Chidlow G, Sikazwe C, Smith DW, Le Souef P, Porsbjerg C. High fractional exhaled nitric oxide and sputum eosinophils are associated with an increased risk of future virus-induced exacerbations: A prospective cohort study. *Clin Exp Allergy* 2017; 47: 1007–1013.
49. Sabogal Pineros YS, Bal SM, Dijkhuis A, Majoor CJ, Dierdorp BS, Dekker T, Hoefsmit EP, Bonta PI, Picavet D, van der Wel NN, Koenderman L, Sterk PJ, Ravanetti L, Lutter R. Eosinophils capture viruses, a capacity that is defective in asthma. *Allergy* 2019; 74: 1898–1909.
50. Sabogal Pineros YS, Bal SM, van de Pol MA, Dierdorp BS, Dekker T, Dijkhuis A, Brinkman P, van der Sluijs KF, Zwinderman AH, Majoor CJ, Bonta PI, Ravanetti L, Sterk PJ, Lutter R. Anti-IL-5 in mild asthma alters rhinovirus-induced macrophage, B-Cell, and neutrophil responses (MATERIAL). A placebo-controlled, double-blind study. *Am J Respir Crit Care Med* 2019; 199: 508–517.
51. Shrimanker R, Pavord ID, Yancey S, Heaney LG, Green RH, Bradding P, Hargadon B, Brightling CE, Wardlaw AJ, Haldar P. Exacerbations of severe asthma in patients treated with mepolizumab. *Eur Respir J* 2018; 52.
52. Jayaram L, Pizzichini MM, Cook RJ, Boulet LP, Lemiere C, Pizzichini E, Cartier A, Hussack P, Goldsmith CH, Laviolette M, Parameswaran K, Hargreave FE. Determining asthma treatment by monitoring sputum cell counts: effect on exacerbations. *Eur Respir J* 2006; 27: 483–494.
53. ClinicalTrials.gov. Exploring asthma exacerbations in mepolizumab treated patients. <https://clinicaltrials.gov/ct2/show/NCT03324230> Date last updated: July 10, 2019. Date last accessed: January 20, 2020.
54. Mitchell PD, Salter BM, Oliveria JP, El-Gammal A, Tworek D, Smith SG, Sehmi R, Gauvreau GM, PM OAB. IL-33 and its receptor ST2 after inhaled allergen challenge in allergic asthmatics. *Int Arch Allergy Immunol* 2018; 176: 133–142.
55. Ying S, O'Connor B, Ratoff J, Meng Q, Mallett K, Cousins D, Robinson D, Zhang G, Zhao J, Lee TH, Corrigan C. Thymic stromal lymphopoietin expression is increased in asthmatic airways and correlates with expression of Th2-attracting chemokines and disease severity. *J Immunol* 2005; 174: 8183–8190.
56. Shikotra A, Choy DF, Ohri CM, Doran E, Butler C, Hargadon B, Shelley M, Abbas AR, Austin CD, Jackman J, Wu LC, Heaney LG, Arron JR, Bradding P. Increased expression of immunoreactive thymic stromal lymphopoietin in patients with severe asthma. *J Allergy Clin Immunol* 2012; 129: 104–111.
57. Ying S, O'Connor B, Ratoff J, Meng Q, Fang C, Cousins D, Zhang G, Gu S, Gao Z, Shamji B, Edwards MJ, Lee TH, Corrigan CJ. Expression and cellular provenance of thymic stromal lymphopoietin and chemokines in patients with severe asthma and chronic obstructive pulmonary disease. *J Immunol* 2008; 181: 2790–2798.

58. Moffatt MF, Gut IG, Demenais F, Strachan DP, Bouzigon E, Heath S, von Mutius E, Farrall M, Lathrop M, Cookson W, Consortium G. A large-scale, consortium-based genomewide association study of asthma. *N Engl J Med* 2010; 363: 1211–1221.
59. Prefontaine D, Lajoie-Kadoch S, Foley S, Audusseau S, Olivenstein R, Halayko AJ, Lemiere C, Martin JG, Hamid Q. Increased expression of IL-33 in severe asthma: evidence of expression by airway smooth muscle cells. *J Immunol* 2009; 183: 5094–5103.
60. Kaur D, Gomez E, Doe C, Berair R, Woodman L, Saunders R, Hollins F, Rose FR, Amrani Y, May R, Kearley J, Humbles A, Cohen ES, Brightling CE. IL-33 drives airway hyper-responsiveness through IL-13-mediated mast cell: airway smooth muscle crosstalk. *Allergy* 2015; 70: 556–567.
61. Momen T, Ahanchian H, Reisi M, Shamsdin SA, Shahsanai A, Keivanfar M. Comparison of interleukin-33 serum levels in asthmatic patients with a control group and relation with the severity of the disease. *Int J Prev Med* 2017; 8: 65.
62. Gasiuniene E, Janulaityte I, Zemeckiene Z, Barkauskiene D, Sitkauskiene B. Elevated levels of interleukin-33 are associated with allergic and eosinophilic asthma. *Scand J Immunol* 2019; 89: e12724.
63. Watanabe M, Nakamoto K, Inui T, Sada M, Honda K, Tamura M, Ogawa Y, Yokoyama T, Saraya T, Kurai D, Ishii H, Takizawa H. Serum sST2 levels predict severe exacerbation of asthma. *Respir Res* 2018; 19: 169.
64. Smith D, Helgason H, Sulem P, Bjornsdottir US, Lim AC, Sveinbjornsson G, Hasegawa H, Brown M, Ketchum RR, Gavala M, Garrett L, Jonasdottir A, Jonasdottir A, Sigurdsson A, Magnusson OT, Eyjolfsson GI, Olafsson I, Onundarson PT, Sigurdardottir O, Gislason D, Gislason T, Ludviksson BR, Ludviksdottir D, Boezen HM, Heinzmann A, Krueger M, Porsbjerg C, Ahluwalia TS, Waage J, Backer V, Deichmann KA, Koppelman GH, Bonnelykke K, Bisgaard H, Masson G, Thorsteinsdottir U, Gudbjartsson DF, Johnston JA, Jonsdottir I, Stefansson K. A rare IL33 loss-of-function mutation reduces blood eosinophil counts and protects from asthma. *PLoS Genet* 2017; 13: e1006659.
65. Soumelis V, Reche PA, Kanzler H, Yuan W, Edward G, Homey B, Gilliet M, Ho S, Antonenko S, Lauerma A, Smith K, Gorman D, Zurawski S, Abrams J, Menon S, McClanahan T, de Waal-Malefy R, Bazan F, Kastelein RA, Liu YJ. Human epithelial cells trigger dendritic cell mediated allergic inflammation by producing TSLP. *Nat Immunol* 2002; 3: 673–680.
66. Karlen S, De Boer ML, Lipscombe RJ, Lutz W, Mordvinov VA, Sanderson CJ. Biological and molecular characteristics of interleukin-5 and its receptor. *Int Rev Immunol* 1998; 16: 227–247.
67. Bradding P, Walls AF, Holgate ST. The role of the mast cell in the pathophysiology of asthma. *J Allergy Clin Immunol* 2006; 117: 1277–1284.
68. Galli SJ, Tsai M. IgE and mast cells in allergic disease. *Nat Med* 2012; 18: 693–704.
69. Allakhverdi Z, Comeau MR, Jessup HK, Yoon BR, Brewer A, Chartier S, Paquette N, Ziegler SF, Sarfati M, Delespesse G. Thymic stromal lymphopoietin is released by human epithelial cells in response to microbes, trauma, or inflammation and potentially activates mast cells. *J Exp Med* 2007; 204: 253–258.
70. Camelo A, Rosignoli G, Ohne Y, Stewart RA, Overed-Sayer C, Sleeman MA, May RD. IL-33, IL-25, and TSLP induce a distinct phenotypic and activation profile in human type 2 innate lymphoid cells. *Blood Adv* 2017; 1: 577–589.
71. Salter BM, Oliveria JP, Nusca G, Smith SG, Tworek D, Mitchell PD, Watson RM, Sehmi R, Gauvreau GM. IL-25 and IL-33 induce type 2 inflammation in basophils from subjects with allergic asthma. *Respir Res* 2016; 17: 5.

72. Murakami-Satsutani N, Ito T, Nakanishi T, Inagaki N, Tanaka A, Vien PT, Kibata K, Inaba M, Nomura S. IL-33 promotes the induction and maintenance of Th2 immune responses by enhancing the function of OX40 ligand. *Allergol Int* 2014; 63: 443–455.
73. Iikura M, Suto H, Kajiwara N, Oboki K, Ohno T, Okayama Y, Saito H, Galli SJ, Nakae S. IL-33 can promote survival, adhesion and cytokine production in human mast cells. *Lab Invest* 2007; 87: 971–978.
74. Johnston LK, Bryce PJ. Understanding interleukin 33 and its roles in eosinophil development. *Front Med (Lausanne)* 2017; 4: 51.
75. Barlow JL, Peel S, Fox J, Panova V, Hardman CS, Camelo A, Bucks C, Wu X, Kane CM, Neill DR, Flynn RJ, Sayers I, Hall IP, McKenzie AN. IL-33 is more potent than IL-25 in provoking IL-13-producing nuocytes (type 2 innate lymphoid cells) and airway contraction. *J Allergy Clin Immunol* 2013; 132: 933–941.
76. Beale J, Jayaraman A, Jackson DJ, Macintyre JDR, Edwards MR, Walton RP, Zhu J, Man Ching Y, Shamji B, Edwards M, Westwick J, Cousins DJ, Yi Hwang Y, McKenzie A, Johnston SL, Bartlett NW. Rhinovirus-induced IL-25 in asthma exacerbation drives type 2 immunity and allergic pulmonary inflammation. *Sci Transl Med* 2014; 6: 256ra134.
77. Tang W, Smith SG, Beaudin S, Dua B, Howie K, Gauvreau G, O'Byrne PM. IL-25 and IL-25 receptor expression on eosinophils from subjects with allergic asthma. *Int Arch Allergy Immunol* 2014; 163: 5–10.
78. Paplinska-Goryca M, Grabczak EM, Dabrowska M, Hermanowicz-Salamon J, Proboszcz M, Nejman-Gryz P, Maskey-Warzechowska M, Krenke R. Sputum interleukin-25 correlates with asthma severity: a preliminary study. *Postepy Dermatol Alergol* 2018; 35: 462–469.
79. Jung JS, Park BL, Cheong HS, Bae JS, Kim JH, Chang HS, Rhim T, Park JS, Jang AS, Lee YM, Kim KU, Uh ST, Na JO, Kim YH, Park CS, Shin HD. Association of IL-17RB gene polymorphism with asthma. *Chest* 2009; 135: 1173–1180.
80. Corrigan CJ, Wang W, Meng Q, Fang C, Eid G, Caballero MR, Lv Z, An Y, Wang YH, Liu YJ, Kay AB, Lee TH, Ying S. Allergen-induced expression of IL-25 and IL-25 receptor in atopic asthmatic airways and late-phase cutaneous responses. *J Allergy Clin Immunol* 2011; 128: 116–124.
81. Tang W, Smith SG, Salter B, Oliveria JP, Mitchell P, Nusca GM, Howie K, Gauvreau GM, O'Byrne PM, Sehmi R. Allergen-induced increases in interleukin-25 and interleukin-25 receptor expression in mature eosinophils from atopic asthmatics. *Int Arch Allergy Immunol* 2016; 170: 234–242.
82. Tworek D, Smith SG, Salter BM, Baatjes AJ, Scime T, Watson R, Obminski C, Gauvreau GM, O'Byrne PM. IL-25 receptor expression on airway dendritic cells after allergen challenge in subjects with asthma. *Am J Respir Crit Care Med* 2016; 193: 957–964.
83. Cheng D, Xue Z, Yi L, Shi H, Zhang K, Huo X, Bonser LR, Zhao J, Xu Y, Erle DJ, Zhen G. Epithelial interleukin-25 is a key mediator in Th2-high, corticosteroid-responsive asthma. *Am J Respir Crit Care Med* 2014; 190: 639–648.
84. Tang W, Smith SG, Du W, Gugilla A, Du J, Oliveria JP, Howie K, Salter BM, Gauvreau GM, O'Byrne PM, Sehmi R. Interleukin-25 and eosinophils progenitor cell mobilization in allergic asthma. *Clin Transl Allergy* 2018; 8: 5.
85. Corren J, Parnes JR, Wang L, Mo M, Roseti SL, Griffiths JM, van der Merwe R. Tezepelumab in adults with uncontrolled asthma. *N Engl J Med* 2017; 377: 936–946.
86. Tanaka J, Watanabe N, Kido M, Saga K, Akamatsu T, Nishio A, Chiba T. Human TSLP and TLR3 ligands promote differentiation of Th17 cells with a central memory phenotype under Th2-polarizing conditions. *Clin Exp Allergy* 2009; 39: 89–100.

87. Gao H, Ying S, Dai Y. Pathological roles of neutrophil-mediated inflammation in asthma and its potential for therapy as a target. *J Immunol Res* 2017: 2017: 3743048.
88. Nagakumar P, Denney L, Fleming L, Bush A, Lloyd CM, Saglani S. Type 2 innate lymphoid cells in induced sputum from children with severe asthma. *J Allergy Clin Immunol* 2016: 137: 624–626.
89. Smith SG, Chen R, Kjarsgaard M, Huang C, Oliveria JP, O'Byrne PM, Gauvreau GM, Boulet LP, Lemiere C, Martin J, Nair P, Sehmi R. Increased numbers of activated group 2 innate lymphoid cells in the airways of patients with severe asthma and persistent airway eosinophilia. *J Allergy Clin Immunol* 2016: 137: 75–86.
90. Phipatanakul W, Mauger DT, Sorkness RL, Gaffin JM, Holguin F, Woodruff PG, Ly NP, Bacharier LB, Bhakta NR, Moore WC, Bleecker ER, Hastie AT, Meyers DA, Castro M, Fahy JV, Fitzpatrick AM, Gaston BM, Jarjour NN, Levy BD, Peters SP, Teague WG, Fajt M, Wenzel SE, Erzurum SC, Israel E, Severe Asthma Research P. Effects of age and disease severity on systemic corticosteroid responses in asthma. *Am J Respir Crit Care Med* 2017: 195: 1439–1448.
91. Liu S, Verma M, Michalec L, Liu W, Sripada A, Rollins D, Good J, Ito Y, Chu H, Gorska MM, Martin RJ, Alam R. Steroid resistance of airway type 2 innate lymphoid cells from patients with severe asthma: The role of thymic stromal lymphopoietin. *J Allergy Clin Immunol* 2018: 141: 257–268.
92. Pavord ID, Brightling CE, Woltmann G, Wardlaw AJ. Non-eosinophilic corticosteroid unresponsive asthma. *Lancet* 1999: 353: 2213–2214.
93. Ortega H, Lemiere C, Llanos JP, Forshag M, Price R, Albers F, Yancey S, Castro M. Outcomes following mepolizumab treatment discontinuation: real-world experience from an open-label trial. *Allergy Asthma Clin Immunol* 2019: 15: 37.
94. Haldar P, Brightling CE, Singapuri A, Hargadon B, Gupta S, Monteiro W, Bradding P, Green RH, Wardlaw AJ, Ortega H, Pavord ID. Outcomes after cessation of mepolizumab therapy in severe eosinophilic asthma: a 12-month follow-up analysis. *J Allergy Clin Immunol* 2014: 133: 921–923.
95. Gauvreau GM, White L, Davis BE. Anti-alarmin approaches entering clinical trials. *Curr Opin Pulm Med* 2019: doi: 10.1097/MCP.0000000000000615. [Epub ahead of print].
96. Gauvreau GM, O'Byrne PM, Boulet LP, Wang Y, Cockcroft D, Bigler J, FitzGerald JM, Boedigheimer M, Davis BE, Dias C, Gorski KS, Smith L, Bautista E, Comeau MR, Leigh R, Parnes JR. Effects of an anti-TSLP antibody on allergen-induced asthmatic responses. *N Engl J Med* 2014: 370: 2102–2110.
97. Corren J, Garcia Gil E, Parnes JR, Pham T-H, Griffiths JM. Tezepelumab treatment effect on annualized rate of exacerbations by baseline biomarkers in uncontrolled severe asthma patients: phase 2b PATHWAY study. *Am J Respir Crit Care Med* 2019: 199: A2621.
98. Corren J, Weinstein S, Janka L, Zangrilli J, Garin M. Phase 3 study of reslizumab in patients with poorly controlled asthma: Effects across a broad range of eosinophil counts. *Chest* 2016: 150: 799–810.
99. Bleecker ER, FitzGerald JM, Chanez P, Papi A, Weinstein SF, Barker P, Sproule S, Gilmartin G, Aurivillius M, Werkstrom V, Goldman M, investigators Ss. Efficacy and safety of benralizumab for patients with severe asthma uncontrolled with high-dosage inhaled corticosteroids and long-acting beta2-agonists (SIROCCO): a randomised, multicentre, placebo-controlled phase 3 trial. *Lancet* 2016: 388: 2115–2127.
100. Castro M, Corren J, Pavord ID, Maspero J, Wenzel S, Rabe KF, Busse WW, Ford L, Sher L, FitzGerald JM, Katelaris C, Tohda Y, Zhang B, Staudinger H, Pirozzi G, Amin N, Ruddy M, Akinlade B, Khan A, Chao J, Martincova R, Graham NMH, Hamilton JD, Swanson BN, Stahl N, Yancopoulos GD, Teper A. Dupilumab efficacy and safety in moderate-to-severe uncontrolled asthma. *N Engl J Med* 2018: 378: 2486–2496.

101. Sanofi/Regeneron. Regeneron and Sanofi announce positive topline phase 2 results for IL-33 antibody in asthma. <https://investor.regeneron.com/news-releases/news-release-details/regeneron-and-sanofi-announce-positive-topline-phase-2-results> Date last updated: June 21, 2019. Date last accessed: January 20, 2020. 2019.
102. Bloechliger M, Reinau D, Spoenclin J, Chang SC, Kuhlbusch K, Heaney LG, Jick SS, Meier CR. Adverse events profile of oral corticosteroids among asthma patients in the UK: cohort study with a nested case-control analysis. *Respir Res* 2018; 19: 75.
103. Bjerkan L, Schreurs O, Engen SA, Jahnsen FL, Baekkevold ES, Blix IJ, Schenck K. The short form of TSLP is constitutively translated in human keratinocytes and has characteristics of an antimicrobial peptide. *Mucosal Immunol* 2015; 8: 49–56.
104. Varricchi G, Pecoraro A, Marone G, Criscuolo G, Spadaro G, Genovese A, Marone G. Thymic stromal lymphopoietin isoforms, inflammatory disorders, and cancer. *Front Immunol* 2018; 9: 1595.
105. Chen WY, Tsai TH, Yang JL, Li LC. Therapeutic strategies for targeting IL-33/ST2 signalling for the treatment of inflammatory diseases. *Cell Physiol Biochem* 2018; 49: 349–358.
106. Vannella KM, Ramalingam TR, Borthwick LA, Barron L, Hart KM, Thompson RW, Kindrachuk KN, Cheever AW, White S, Budelsky AL, Comeau MR, Smith DE, Wynn TA. Combinatorial targeting of TSLP, IL-25, and IL-33 in type 2 cytokine-driven inflammation and fibrosis. *Sci Transl Med* 2016; 8: 337ra365.
107. Han M, Rajput C, Hong JY, Lei J, Hinde JL, Wu Q, Bentley JK, Hershenson MB. The innate cytokines IL-25, IL-33, and TSLP cooperate in the induction of type 2 innate lymphoid cell expansion and mucous metaplasia in rhinovirus-infected immature mice. *J Immunol* 2017; 199: 1308–1318.
108. Snell NJ. Discontinued drug projects in the respiratory therapeutic area during 2012. *Expert Opin Investig Drugs* 2014; 23: 411–415.
109. Teach SJ, Gill MA, Togias A, Sorkness CA, Arbes SJ, Jr., Calatroni A, Wildfire JJ, Gergen PJ, Cohen RT, Pongracic JA, Kercksmar CM, Khurana Hershey GK, Gruchalla RS, Liu AH, Zoratti EM, Kattan M, Grindle KA, Gern JE, Busse WW, Szeffler SJ. Preseasonal treatment with either omalizumab or an inhaled corticosteroid boost to prevent fall asthma exacerbations. *J Allergy Clin Immunol* 2015; 136: 1476–1485.
110. Pham T-H, Ren P, Parnes JR, Griffiths JM. Tezepelumab reduces multiple key inflammatory biomarkers in patients with severe, uncontrolled asthma in the phase 2b PATHWAY study. *Am J Respir Crit Care Med* 2019; 199: A2677.
111. Baatjes AJ, Smith SG, Dua B, Watson R, Gauvreau GM, O'Byrne PM. Treatment with anti-OX40L or anti-TSLP does not alter the frequency of T regulatory cells in allergic asthmatics. *Allergy* 2015; 70: 1505–1508.
112. Wenzel S, Ford L, Pearlman D, Spector S, Sher L, Skobieranda F, Wang L, Kirkesseli S, Rocklin R, Bock B, Hamilton J, Ming JE, Radin A, Stahl N, Yancopoulos GD, Graham N, Pirozzi G. Dupilumab in persistent asthma with elevated eosinophil levels. *N Engl J Med* 2013; 368: 2455–2466.
113. Rabe KF, Nair P, Brusselle G, Maspero JF, Castro M, Sher L, Zhu H, Hamilton JD, Swanson BN, Khan A, Chao J, Staudinger H, Pirozzi G, Antoni C, Amin N, Ruddy M, Akinlade B, Graham NMH, Stahl N, Yancopoulos GD, Teper A. Efficacy and safety of dupilumab in glucocorticoid-dependent severe asthma. *N Engl J Med* 2018; 378: 2475–2485.
114. Bhutani M, Yang WH, Hebert J, de Takacs F, Stril JL. The real world effect of omalizumab add on therapy for patients with moderate to severe allergic asthma: The ASTERIX Observational study. *PLoS One* 2017; 12: e0183869.

115. Busse W, Corren J, Lanier BQ, McAlary M, Fowler-Taylor A, Cioppa GD, van As A, Gupta N. Omalizumab, anti-IgE recombinant humanized monoclonal antibody, for the treatment of severe allergic asthma. *J Allergy Clin Immunol* 2001; 108: 184–190.
116. Massanari M, Holgate ST, Busse WW, Jimenez P, Kianifard F, Zeldin R. Effect of omalizumab on peripheral blood eosinophilia in allergic asthma. *Respir Med* 2010; 104: 188–196.
117. Djukanovic R, Wilson SJ, Kraft M, Jarjour NN, Steel M, Chung KF, Bao W, Fowler-Taylor A, Matthews J, Busse WW, Holgate ST, Fahy JV. Effects of treatment with anti-immunoglobulin E antibody omalizumab on airway inflammation in allergic asthma. *Am J Respir Crit Care Med* 2004; 170: 583–593.
118. Molimard M, Buhl R, Niven R, Le Gros V, Thielen A, Thirlwell J, Maykut R, Peachey G. Omalizumab reduces oral corticosteroid use in patients with severe allergic asthma: real-life data. *Respir Med* 2010; 104: 1381-1385.
119. Pavord ID, Korn S, Howarth P, Bleecker ER, Buhl R, Keene ON, Ortega H, Chanez P. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. *Lancet* 2012; 380: 651–659.
120. Flood-Page P, Swenson C, Faiferman I, Matthews J, Williams M, Brannick L, Robinson D, Wenzel S, Busse W, Hansel TT, Barnes NC, International Mepolizumab Study G. A study to evaluate safety and efficacy of mepolizumab in patients with moderate persistent asthma. *Am J Respir Crit Care Med* 2007; 176: 1062-1071.
121. Bel EH, Wenzel SE, Thompson PJ, Prazma CM, Keene ON, Yancey SW, Ortega HG, Pavord ID, Investigators S. Oral glucocorticoid-sparing effect of mepolizumab in eosinophilic asthma. *N Engl J Med* 2014; 371: 1189–1197.
122. Castro M, Zangrilli J, Wechsler ME, Bateman ED, Brusselle GG, Bardin P, Murphy K, Maspero JF, O'Brien C, Korn S. Reslizumab for inadequately controlled asthma with elevated blood eosinophil counts: results from two multicentre, parallel, double-blind, randomised, placebo-controlled, phase 3 trials. *Lancet Respir Med* 2015; 3: 355–366.
123. Castro M, Mathur S, Hargreave F, Boulet LP, Xie F, Young J, Wilkins HJ, Henkel T, Nair P, Res-5- Study G. Reslizumab for poorly controlled, eosinophilic asthma: a randomized, placebo-controlled study. *Am J Respir Crit Care Med* 2011; 184: 1125–1132.
124. Ibrahim H, O'Sullivan R, Casey D, Murphy J, MacSharry J, Plant BJ, Murphy DM. The effectiveness of reslizumab in severe asthma treatment: a real-world experience. *Respir Res* 2019; 20: 289.
125. Laviolette M, Gossage DL, Gauvreau G, Leigh R, Olivenstein R, Katial R, Busse WW, Wenzel S, Wu Y, Datta V, Kolbeck R, Molfino NA. Effects of benralizumab on airway eosinophils in asthmatic patients with sputum eosinophilia. *J Allergy Clin Immunol* 2013; 132: 1086–1096.
126. Nair P, Wenzel S, Rabe KF, Bourdin A, Lugogo NL, Kuna P, Barker P, Sproule S, Ponnarambil S, Goldman M, Investigators ZT. Oral glucocorticoid-sparing effect of benralizumab in severe asthma. *N Engl J Med* 2017; 376: 2448–2458.
127. Lee HY, Rhee CK, Kang JY, Byun JH, Choi JY, Kim SJ, Kim YK, Kwon SS, Lee SY. Blockade of IL-33/ST2 ameliorates airway inflammation in a murine model of allergic asthma. *Exp Lung Res* 2014; 40: 66–76.
128. Ballantyne SJ, Barlow JL, Jolin HE, Nath P, Williams AS, Chung KF, Sturton G, Wong SH, McKenzie AN. Blocking IL-25 prevents airway hyperresponsiveness in allergic asthma. *J Allergy Clin Immunol* 2007; 120: 1324–1331.

**TABLE 1 Clinical evidence of the effects of TSLP blockade in humans\***

Reference	Setting	Outcomes
[85]	Patients with uncontrolled moderate-to-severe asthma (phase 2b study)	<ul style="list-style-type: none"><li>• Reduced exacerbation rate irrespective of baseline blood eosinophil count</li><li>• Increased FEV<sub>1</sub></li><li>• Reduced blood eosinophils, Fe<sub>NO</sub>, serum IgE throughout the 52-week treatment period</li></ul>
[110]	Patients with uncontrolled moderate-to-severe asthma (phase 2b study)	<ul style="list-style-type: none"><li>• Reduced blood eosinophils, Fe<sub>NO</sub>, serum IgE, IL-5, IL-13, periostin and TARC throughout the 52-week treatment period</li><li>• Reduced exacerbation rate irrespective of baseline blood eosinophils, Fe<sub>NO</sub>, serum IgE, IL-5, IL-13, periostin, TARC</li></ul>
[96]	Patients with mild allergic asthma (phase 1/2a study) after allergen challenge	<ul style="list-style-type: none"><li>• Reduced airway hyperresponsiveness</li><li>• Reduced bronchoconstriction</li><li>• Reduced blood and sputum eosinophils</li><li>• Reduced Fe<sub>NO</sub></li><li>• Reduced Th2:Th1 cell ratio in blood</li></ul>
[111]	Patients with mild allergic asthma (phase 1/2a study) after allergen challenge	<ul style="list-style-type: none"><li>• No change in T regulatory cell frequency</li></ul>

Fe<sub>NO</sub>: fractional exhaled nitric oxide; FEV<sub>1</sub>: forced expiratory volume in 1 second; IgE: immunoglobulin E; IL: interleukin; TARC: thymus and activation-regulated chemokine; Th: T helper; TSLP: thymic stromal lymphopoietin. \*All data are from studies of anti-TSLP human monoclonal antibody treatment. No clinical data evaluating anti-IL-33 or anti-IL-25 treatment have yet been published in peer-reviewed journals.



**TABLE 2 Effects on T2 inflammatory biomarkers, and OCS-sparing potential, of anti-TSLP and approved biologic therapies for asthma**

Biologic	Effect on inflammatory biomarker				OCS-sparing data
	Fe <sub>NO</sub>	IgE	Blood eosinophils	Sputum eosinophils	
Tezepelumab (anti-TSLP)	↓ [85, 96, 110]	↓ [85, 96, 110]	↓ [85, 96, 110]	↓ [96]	Data pending (NCT03406078)
Dupilumab (anti-IL-4/IL-13)	↓ [100]	↓ [100]	↑ [100]	Insufficient data [112]	~70% reduction in daily OCS dosage (vs ~42% with placebo); 52% of patients achieved complete OCS weaning (placebo, 29%) [113]
Omalizumab (anti-IgE)	Minimal effects [114]	↓ [115]	Minimal effects [116]	↓ [117]	No RCT data; real-world data indicates reduction in OCS use [118]
Mepolizumab (anti-IL-5)	No effect [119]	No effect*	↓ [119, 120]	↓ [119, 120]	~50% reduction in daily OCS dosage (vs 0% with placebo); 14% of patients achieved complete OCS weaning (placebo, 8%) [121]
Reslizumab (anti-IL-5)	No effect <sup>†</sup>	No effect*	↓ [98, 122]	↓ [123]	No RCT data; real-world data indicates reduction in OCS use [124]
Benralizumab (anti-IL-5Rα)	No effect <sup>†</sup>	No effect*	↓ [99, 125]	↓ [125]	~75% reduction in daily OCS dosage (vs ~25% with placebo); 52–56% of patients achieved complete OCS weaning (placebo, 19%) [126]

Fe<sub>NO</sub>: fractional exhaled nitric oxide; IgE: immunoglobulin E; IL: interleukin; OCS, oral corticosteroid; RCT, randomized controlled trial; T2: type 2; TSLP: thymic stromal lymphopoietin. \*Observed in clinical practice. <sup>†</sup>Expected based on observations with other anti-IL-5 biologics.

**TABLE 3 Ongoing and recently completed clinical studies of anti-alarmin treatments in patients with asthma**

Drug	ClinicalTrials.gov identifier	Phase	Population	Primary endpoint
<b><i>Anti-TSLP</i></b>				
Tezepelumab	NCT02698501	2	Asthma, taking daily ICS	Decrease in airway hyperresponsiveness to mannitol
	NCT03688074	2	Uncontrolled asthma	Change from baseline in the number of airway submucosal inflammatory cells
	NCT03406078	3	Severe, uncontrolled asthma	Reduction in OCS dose while not losing asthma control
	NCT03347279	3	Severe, uncontrolled asthma	Annualised asthma exacerbation rate
CSJ117	NCT03138811	1	Mild, atopic asthma	Number of adverse events and serious adverse events
<b><i>Anti-IL-33</i></b>				
REGN3500 (SAR440340)	NCT02999711	1	Mild-to-moderate asthma	Number of treatment-emergent adverse events
	NCT03112577	1	Mild, allergic asthma	Sputum inflammatory biomarkers
AMG282 (RG6149/MSTT1041A)	NCT02918019	2b	Severe, uncontrolled asthma	Rate of exacerbations
Etokimab (ANB020)	NCT03469934	2a	Severe, eosinophilic asthma	Change in blood eosinophil count
GSK3772847 (CNT07160)	NCT03207243	2a	Moderately severe asthma	Loss of asthma control events at week 16

NCT03393806

2b

Moderate-to-severe asthma with allergic fungal airway disease    Blood eosinophil count and Fe<sub>NO</sub> levels at week 13

---

Fe<sub>NO</sub>: fractional exhaled nitric oxide; ICS: inhaled corticosteroids; IL: interleukin; OCS: oral corticosteroids; TSLP: thymic stromal lymphopoietin.

## **FIGURE LEGENDS**

### **FIGURE 1 Role of the alarmins in driving T2 inflammation, biomarkers and clinical outcomes in asthma.**

EOS: eosinophils; Fe<sub>NO</sub>: fractional exhaled nitric oxide; IgE: immunoglobulin E; IL: interleukin; ILC2: type 2 innate lymphoid cell; T2: type 2; Th2: type 2 T helper cell; TSLP: thymic stromal lymphopoietin. \*Released from Th2 cells and ILC2s.

### **FIGURE 2 Effects of blocking the alarmins on biomarkers and clinical outcomes in asthma.**

EOS: eosinophils; Fe<sub>NO</sub>: fractional exhaled nitric oxide; IgE: immunoglobulin E; IL: interleukin; T2: type 2; TSLP: thymic stromal lymphopoietin. Grey boxes represent findings from studies in mice, which are yet to be confirmed in humans [127, 128]. <sup>†</sup>Serum IL-4 was not measured in the tezepelumab phase 2b study. <sup>‡</sup>Top-line findings reported from a phase 2a study of anti-IL-33 [101], which are yet to be published in a peer-reviewed journal.

# Viruses, allergens, cigarette smoke and pollution



