



@ERSpublications

Delamanid shows low minimal inhibitory concentrations *in vitro* against *M. avium-intracellulare* strains <http://ow.ly/xDiM304ul7O>

David Krieger¹, Nicolas Schönfeld¹, Silvan Vesenbeckh¹, Gudrun Bettermann², Torsten Thomas Bauer^{1,3}, Holger Rüssmann² and Harald Mauch²

¹Klinik für Pneumologie, Lungenklinik Heckeshorn, HELIOS Klinikum Emil von Behring, Berlin, Germany. ²Institut für Mikrobiologie, Immunologie und Laboratoriumsmedizin, HELIOS Klinikum Emil von Behring, Berlin, Germany.

³Deutsches Zentralkomitee zur Bekämpfung der Tuberkulose (DZK), Berlin, Germany.

Correspondence: David Krieger, Klinik für Pneumologie, Lungenklinik Heckeshorn, HELIOS Klinikum Emil von Behring, Waltherhoeferstr. 11, 14165 Berlin, Germany. E-mail: david.krieger@helios-kliniken.de

Received: July 15 2016 | Accepted after revision: Sept 13 2016 | First published online: Nov 11 2016

Conflict of interest: None declared.

References

- 1 Matsumoto M, Hashizume H, Tomishige T, *et al.* OPC-67683, a nitro-dihydro-imidazooxazole derivative with promising action against tuberculosis *in vitro* and in mice. *PLoS Med* 2006; 3: e466.
- 2 Skripconoka V, Danilovits M, Pehme L, *et al.* Delamanid improves outcomes and reduces mortality in multidrug-resistant tuberculosis. *Eur Respir J* 2013; 41: 1393–1400.
- 3 Sotgiu G, Pontali E, Centis R, *et al.* Delamanid (OPC-67683) for treatment of multi-drug-resistant tuberculosis. *Expert Rev Anti Infect Ther* 2015; 13: 305–315.
- 4 Tadolini M, Garcia-Prats AJ, D'Ambrosio L, *et al.* Compassionate use of new drugs in children and adolescents with multidrug-resistant and extensively drug-resistant tuberculosis: early experiences and challenges. *Eur Respir J* 2016; 48: 938–943.
- 5 Tadolini M, Lingsang RD, Tiberi S, *et al.* First case of extensively drug-resistant tuberculosis treated with both delamanid and bedaquiline. *Eur Respir J* 2016; 48: 935–938.
- 6 Diacon AH, Dawson R, Hanekom M, *et al.* Early bactericidal activity of delamanid (OPC-67683) in smear-positive pulmonary tuberculosis patients. *Int J Tuberc Lung Dis* 2011; 15: 949–954.
- 7 Xu HB, Jiang RH, Li L. Treatment outcomes for *Mycobacterium avium* complex: a systematic review and meta-analysis. *Eur J Clin Microbiol Infect Dis* 2014; 33: 347–358.
- 8 Griffith DE, Aksamit T, Brown-Elliott BA, *et al.* An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. *Am J Respir Crit Care Med* 2007; 175: 367–416.
- 9 Schönfeld N, Haas W, Richter E, *et al.* Recommendations for diagnosis and treatment of nontuberculous mycobacterioses of the German Central Committee against tuberculosis and the German Respiratory Society. *Pneumologie* 2013; 67: 605–633.
- 10 Raju RM, Raju SM, Zhao Y, *et al.* Leveraging advances in tuberculosis diagnosis and treatment to address nontuberculous mycobacterial disease. *Emerg Infect Dis* 2016; 22: 365–369.
- 11 Pontali E, Sotgiu G, D'Ambrosio L, *et al.* Bedaquiline and multidrug-resistant tuberculosis: a systematic and critical analysis of the evidence. *Eur Respir J* 2016; 47: 394–402.
- 12 Radenbach KL. Diagnostische und therapeutische Fortschritte bei nichttuberkulösen Mykobakteriosen [Diagnostic and therapeutic progress in nontuberculous mycobacterioses]. *Prax Klin Pneumol* 1985; 39: 43–49.
- 13 Schönfeld N, Bergmann T, Vesenbeckh S, *et al.* Minimal inhibitory concentrations of first-line drugs of multidrug-resistant tuberculosis isolates. *Lung India* 2012; 29: 309–312.
- 14 Kent PT, Kubica GP. Public health mycobacteriology: a guide for the level III laboratory. Atlanta, USDHHS, Centers for Disease Control, 1985.
- 15 McClatchy JK. Antimycobacterial drugs: mechanism of action, drug resistance, susceptibiltiy testing and assays of activity in biological fluids. In: Lorian V, ed., *Antibiotics in Laboratory Medicine*, 2nd Edn. Baltimore, The Williams and Wilkins Co., 1986; pp. 181–222.

Eur Respir J 2016; 48: 1803–1804 | DOI: 10.1183/13993003.01420-2016 | Copyright ©ERS 2016

Specific airway resistance in preschool children: why not panting after all?



CrossMark

To the Editor:

Specific airway resistance (sR_{aw}) is measured with minimal cooperation in the preschool child during tidal breathing [1]. Methodological difficulties have been encountered in modern plethysmographs when the warming and humidification of the inspired gas [2] are replaced by numerical algorithms to eliminate the thermo hygrometric artefact [1, 3, 4]. Measuring sR_{aw} during panting [5] had been dismissed in preschool children based on the assumption that the ventilatory manoeuvre would be difficult to perform and

standardise. The feasibility of such an assumption, however, has not been verified. Therefore, the aim of this study was to assess the feasibility of measuring sR_{aw} during panting in preschool children with asthma and compare the outcome with the tidal breathing method.

Preschool children with a doctor diagnosis of asthma were recruited from the paediatric pulmonology clinics (Hôpital d'enfants, Vandoeuvre-lès-Nancy, France). Written informed consent was obtained from the children and their parents. The protocol was approved by the ethics committee (Comité de Protection des Personnes EST III, CHU de Nancy, Nancy, France). The plethysmography equipment, which has been described elsewhere [6], is operated by software that includes an algorithm which should eliminate the thermo hygrometric artefact from the plethysmographic volume signal (ΔV_{pleth}), which may be activated during tidal breathing and disabled during panting. Acquisition consisted of a series of four breaths selected automatically by the software and displayed on the computer screen. Three acquisitions were collected for each condition. Measurements were selected to retain those showing no artefact and where breathing frequency was lower than 50 breaths·min⁻¹ during tidal breathing or higher than 60 breaths per min⁻¹ during panting. sR_{aw} was computed between points of maximum ΔV_{pleth} (sR_{tot}) and in the flow range ± 0.5 L·s⁻¹ ($sR_{aw0.5}$). The overall flow interval within a given acquisition was measured graphically. Data were analysed with Pearson's Chi-squared test or analysis of variance as appropriate, and are expressed as mean \pm SD.

A total of 127 preschool children (73 boys) aged 3.5–6.5 years took part in the protocol. Measurements were achieved during tidal breathing in 83 children (34 ± 7 breaths·min⁻¹; flow amplitude 1.3 ± 0.3 L·s⁻¹) and during panting in 90 children (130 ± 36 breaths·min⁻¹; flow amplitude 2.0 ± 0.6 L·s⁻¹; $p < 0.0001$ for both breathing frequency and flow amplitude). Feasibility of measurement during tidal breathing and panting was not significantly different overall (65 and 71% respectively; $p = 0.35$), nor within age categories (figure 1a; $p > 0.05$). There was no difference between boys and girls during tidal breathing (64 versus 67%; $p = 0.9$) or panting (71 versus 70%; $p = 0.9$). In those with successful measurements, the average number of validated breaths was 8.9 ± 2.8 during tidal breathing and 9.6 ± 2.4 during panting ($p = 0.089$). The percentage of valid breaths obtained did not differ between either breathing regimens (figure 1b) or sexes.

$sR_{aw0.5}$ and sR_{tot} were significantly smaller during panting (0.71 ± 0.2 kPa·s·L⁻¹ and 0.96 ± 0.3 kPa·s·L⁻¹) than the respective value during tidal breathing (1.14 ± 0.2 kPa·s·L⁻¹ and 1.33 ± 0.2 kPa·s·L⁻¹; $p < 0.0001$ for both $sR_{aw0.5}$ and sR_{tot}). sR_{tot} was also significantly larger than $sR_{aw0.5}$ for both protocols ($p < 0.0001$). The intra-subject coefficient of variation was significantly larger during panting than during tidal breathing for both $sR_{aw0.5}$ ($13 \pm 7\%$ versus $8.5 \pm 6\%$; $p < 0.0001$) and sR_{tot} ($15 \pm 9\%$ versus $10 \pm 7\%$; $p = 0.0001$) and significantly larger for sR_{tot} than $sR_{aw0.5}$ during panting ($p = 0.02$).

The potential of sR_{aw} in paediatric lung function testing was formerly demonstrated in studies where the thermal component in ΔV_{pleth} was properly eliminated by warming and humidifying the respired gas [2]. Recent studies eventually showed important equipment-dependent variability [1, 3] when numerical algorithms were used to correct for the thermo hygrometric artefact. The lack of agreement on sR_{aw} standardisation in lung function centres across the world was recently highlighted [7]. The finding that panting achieves a similar rate of successful measurements as tidal breathing suggests the interesting possibility of avoiding the methodological difficulties encountered with automated software that corrects for the thermo hygrometric component using algorithms only known to the manufacturers. With smaller tidal volumes and higher breathing frequencies, the thermo hygrometric component in ΔV_{pleth} is minimised [8] and the ΔV_{pleth} –flow relationship is studied within a time domain mostly unaffected by the dynamics of airways – gas thermo hygrometric exchanges [9].

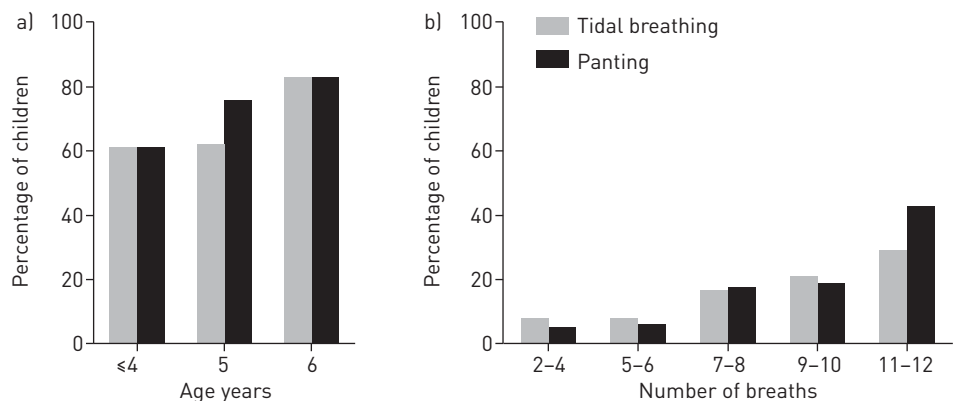


FIGURE 1 a) The success rate of specific airway resistance among age groups and b) the percentage of children achieving a given number of valid breaths are similar during tidal breathing and panting.

The current results raise the question of which estimate of sR_{aw} would be optimal as a routine paediatric lung function end-point. Panting opens the glottis [10], but turbulent airstream has the opposite influence on the measurement outcome. When $sR_{aw_{0.5}}$ is examined, however, the second mechanism is minimised, and the glottic opening should be more fully expressed to decrease sR_{aw} . In fact, similar to a previous report in older children [6], in the current study sR_{aw} was consistently lower during panting than during tidal breathing, and the highly significant difference was observed with both sR_{tot} and $sR_{aw_{0.5}}$. During tidal breathing, sR_{aw} was also reported to be smaller in preschool children when the respired gas was conditioned to body temperature, ambient pressure, saturated with water vapour (BTPS) than when it was numerically corrected for the thermo hygrometric effect [11]. Taken together, these observations in preschool children indicate that either BTPS conditioning or panting, by adequately correcting for the thermo hygrometric artefact, lead to an estimate of sR_{aw} being smaller than that achieved through software corrections applied during tidal breathing. However, the intra-subject variability in the current study was larger during panting than during tidal breathing, and whatever the mechanism, this might impede the ability of sR_{aw} to detect airway obstruction and reversibility. The fact that sR_{tot} was significantly larger than the corresponding $sR_{aw_{0.5}}$ was expected because of the enhanced contribution of nonlinearities and increased slope of airway pressure–flow relationship. Since flow is turbulent mostly in proximal airways, the diagnostic value of sR_{tot} may be lower than for $sR_{aw_{0.5}}$. Furthermore, the larger variability observed with sR_{tot} compared with $sR_{aw_{0.5}}$ was also expected, especially during panting, as the ventilatory effort is likely to vary from one breath to another, resulting in variable flow amplitude and contribution of nonlinearities throughout the acquisition. Hence, computation over a limited flow interval as reported in adults should be recommended [12], with the recent indication of better separation of children with asthma from controls based on $sR_{aw_{0.5}}$ [13]. However, it should be kept in mind that, in preschool children where ΔV_{pleth} and flow signals are small, computation of sR_{aw} within a limited flow range may compromise the signal-to-noise ratio. The fixed flow interval $\pm 0.5 \text{ L}\cdot\text{s}^{-1}$ is also likely to provide age-dependent estimates of sR_{aw} with more significant nonlinearities in shorter than taller children. An important step towards standardisation would be to define the optimal flow range to estimate sR_{aw} and the optimal breathing frequency interval for measuring with minimal variability.

In summary, valid measurements of sR_{aw} may be obtained during panting in preschool children. Since ΔV_{pleth} requires no correction, the test should be more readily applicable to any equipment, which is particularly helpful in the context of international collaborative studies. Compared with tidal breathing, however, the technique achieves lower sR_{aw} and larger intra-subject variability. Further case–control studies, including other lung function outcomes such as spirometry, are required to establish normative data and assess which estimate of sR_{aw} is the most useful for routine paediatric lung function studies.



@ERSpublications

Specific airway resistance may be measured in preschool children during routine lung function

<http://ow.ly/pvrx3034Bo8>

Iulia Ioan^{1,2}, Silvia Demoulin-Alexikova^{1,2}, Laurianne Coutier^{1,2}, Claude Bonabel^{1,2}, Jane Kirkby³, Kim G. Nielsen⁴, Waldemar Tomalak⁵, Bruce Thompson⁶, Cyril Schweitzer^{1,2}, Paul D. Robinson^{3,7,8} and François Marchal^{1,2}

¹Dept of Pediatric Lung Function Testing, Children Hospital, Vandoeuvre, France. ²Dept of Physiology, EA 3450, Faculty of Medicine, University of Lorraine, Vandoeuvre, France. ³University College London, Institute of Child Health, London, UK.

⁴Danish PCD & chILD Centre, CF Centre Copenhagen, Paediatric Pulmonary Service, Dept of Paediatrics and Adolescent Medicine, Copenhagen University Hospital, Rigshospitalet, Copenhagen, Denmark. ⁵National Research Institute for Tuberculosis and Lung Diseases, Rabka Branch, Rabka, Poland. ⁶Physiology Service Allergy, Immunology and Respiratory Medicine, The Alfred Hospital, Melbourne, Australia. ⁷Dept of Respiratory Medicine, The Children's Hospital at Westmead, Sydney, Australia. ⁸Dept of Paediatrics and Child Health, University of Sydney, Sydney, Australia.

Correspondence: François Marchal, Service d'Explorations Fonctionnelles Pédiatriques, Hôpital d'enfants, rue du Morvan, 54511 Vandoeuvre, France. E-mail: f.marchal@chru-nancy.fr

Received: April 11 2016 | Accepted after revision: Aug 04 2016 | First published online: Oct 20 2016

Conflict of interest: None declared.

Acknowledgements: Waldemar Tomalak passed away while the study was progressing. The members of the working group miss a good friend and an esteemed collaborator. They address their sincere condolences to his family.

References

- 1 Kirkby J, Stanojevic S, Welsh L, *et al*. Reference equations for specific airway resistance in children: the Asthma UK initiative. *Eur Respir J* 2010; 36: 622–629.
- 2 Dab I, Alexander F. A simplified approach to the measurement of specific airway resistance. *Pediatr Res* 1976; 10: 998–999.
- 3 Poorisrisak P, Vrang C, Henriksen JM, *et al*. Accuracy of whole-body plethysmography requires biological calibration. *Chest* 2009; 135: 1476–1480.

- 4 Paton J, Beardsmore C, Lavery A, *et al.* Discrepancies between pediatric laboratories in pulmonary function results from healthy children. *Pediatr Pulmonol* 2012; 47: 588–596.
- 5 Dubois AB. Airway resistance. *Am J Respir Crit Care Med* 2000; 162: 345–346.
- 6 Coutier L, Varechova S, Demoulin B, *et al.* Specific airway resistance in children: panting or tidal breathing? *Pediatr Pulmonol* 2014; 49: 245–251.
- 7 Robinson PD, Stocks J, Marchal F, *et al.* Poor standardisation of plethysmographic specific airways resistance measurement despite widespread use. *Eur Respir J* 2015; 46: 1811–1814.
- 8 Dubois AB, Botelho SY, Comroe JH Jr. A new method for measuring airway resistance in man using a body plethysmograph: values in normal subjects and in patients with respiratory disease. *J Clin Invest* 1956; 35: 327–335.
- 9 Peslin R, Duvivier C, Vassiliou M, *et al.* Thermal artifacts in plethysmographic airway resistance measurements. *J Appl Physiol* 1995; 79: 1958–1965.
- 10 Stănescu DC, Clément J, Pattijn J, *et al.* Glottis opening and airway resistance. *J Appl Physiol* 1972; 32: 460–466.
- 11 Klug B, Bisgaard H. Measurement of the specific airway resistance by plethysmography in young children accompanied by an adult. *Eur Respir J* 1997; 10: 1599–1605.
- 12 Briscoe WA, Dubois AB. The relationship between airway resistance, airway conductance and lung volume in subjects of different age and body size. *J Clin Invest* 1958; 37: 1279–1285.
- 13 Coutier L, Ioan I, Sadegh-Eghbali A, *et al.* Flow dependence of specific airway resistance and diagnostic of asthma in children. *Pediatr Pulmonol* 2015; 50: 1107–1112.

Eur Respir J 2016; 48: 1804–1807 | DOI: 10.1183/13993003.01302-2016 | Copyright ©ERS 2016



Dichotomy in pulmonary graft-versus-host disease evident among allogeneic stem-cell transplant recipients undergoing lung transplantation

To the Editor:

Allogeneic haematopoietic stem-cell transplantation (HSCT) has become a life-saving treatment option for numerous benign and malignant diseases, with more than 14 500 procedures being performed annually in Europe alone [1]. Late-onset noninfectious pulmonary complications (NIPCs) have emerged as the main hurdle to long-term survival, affecting up to 26% of HSCT recipients and conferring 2- and 5-year survival rates of 44% and 13%, respectively [2, 3].

Current diagnostic criteria remain based on those proposed by the National Institutes of Health (NIH) in 2005 for pulmonary graft-versus-host disease (GvHD), which considered bronchiolitis obliterans exclusively accountable [4, 5]. Whilst histological confirmation was encouraged, invasive diagnostics have remained contentious due to reported complication rates from open lung biopsies and limited interpretability of trans-bronchial biopsies [6, 7].

Inevitably, surrogate parameters such as new-onset airway obstruction on spirometry and “air-trapping” on computed tomography (CT) have gained precedence in establishing a diagnosis. NIH guidance for monitoring pulmonary GvHD currently incorporates the lung function score that assesses symptoms, forced expiratory volume in 1 s (FEV₁) and diffusing capacity of the lung for carbon monoxide. Currently, diagnosis of pulmonary GvHD requires [4]: 1) FEV₁/forced vital capacity (FVC) <0.7, FEV₁ <75% predicted or residual volume >120% predicted; 2) high-resolution CT demonstrating air trapping, small airway thickening or bronchiectasis; 3) exclusion of infection; and 4) chronic GvHD in at least one extrapulmonary site.

Recently, evidence has emerged suggesting the presence of an additional restrictive form, due to interstitial disease rather than skin GvHD of the chest wall, as has been previously suggested [8, 9]. Through novel assessment of explanted native lungs from patients undergoing lung transplantation (LTx) for end-stage lung disease following allogeneic stem cell transplantation, we compared clinical, radiographic and histological data in this sub-group in an attempt to improve understanding of the pathological processes involved.

Patients who underwent cadaveric LTx between May 1, 2003 and May 1, 2014 at 12 European centres were included. Clinical, imaging and pathological data were collected using standardised reporting forms to facilitate central retrospective analysis. Spirometry was performed in accordance with American Thoracic