phagocyte enzymes in bronchoalveolar lavage from patients with pulmonary sarcoidosis and collagen vascular disorders

Y. Sibille* ***, J.B. Martinot*, L.L. Polomski***, B. Wallaert**, M. Demusis***, J.A. Rankin***, C. Voisin**, J.B.L. Gee***

ate enzymes in bronchoalveolar lavage from patients with pulmonary is and collagen vascular disorders. Y. Sibille, J.B. Martinot, L.L. M. B. Wallaert, M. Demusis, J.A. Rankin, C. Voisin, J.B.L. Gee. RACT: The balance between proteases and antiproteases in the lower story tract is believed to play a role in the outcome of interstitial teases. In this cross-sectional study, we measure several phagocyte d enzymes, namely plasminogen activator, neutrophil elastase and defined protease active on the trialanine chromophore substrate d-alanine,-nitroanilide (SLAPN) in bronchoalveolar lavage (BAL) from 42 patients with pulmonary sarcoidosis and from 43 patients collagen vascular disease (CVD), 22 without lung disease (group I) associated with parenchymal lung disease (group II). The results a) that sarcoidosis is associated with increased plasminogen activautivity and with the presence of enzymatic activity against SLAPN onding at least in part to a metalloprotease; b) that CVD in the of radiographic lung disease is associated with an increase of ogen activator activity and increased levels of alpha, antiproteasehil elastase complexes; c) that the majority of untreated CVD (group stlents have detectable levels of neutrophil elastase activity. These show that patients with pulmonary sarcoldosis and CVD have tenzymatic profiles in their lower respiratory tract as assessed by Al. Thus, sarcoidosis (mostly lymphocytic) is associated with enhanced phage-derived proteolytic activity in BAL, while CVD patients both and without lung disease have increased neutrophil counts and airophil elastase complexed to alpha protease inhibitor and mandly inactive in BAL. Finally, only BAL from untreated CVD mb with interstitial lung disease contain neutrophil elastase activity. falter activity could contribute to the lung lesions frequently observed ese disorders.

Le Respir J., 1990, 3, 249-256.

* Pulmonary Section and Experimental Medicine Unit, International Institute of Cellular and Molecular Pathology, Catholic University of Louvain, Belgium.

** Dept of Pneumology, Hôpital A. Calmette, the Pasteur Institute, Lille, France.

*** Pulmonary Section, Dept of Medicine, Yale University School of Medicine, New Haven, and West Haven Veterans Hospital, West Haven, CT, USA.

Correspondence: Dr Y. Sibille, Pulmonary Section, Cliniques UCL Mont-Godinne, B-5180 Yvoir, Belgium.

Keywords: Bronchoalveolar lavage; interstitial lung disease; protease.

Received: December, 1988; accepted after revision October 25, 1989.

Work supported by Grant No. 1667A from the Council for Tobacco Research, by INSERM (Réseau de Recherche Clinique) and by Université de Lille II.

Impairment of the balance between proteases and improteases is believed to play a critical role in both and chronic lung injury [1, 2]. Proteolytic activity is the lung has been related to adult respiratory thress syndrome (ARDS), emphysema and diffuse almonary fibrosis [3-5].

hulmonary fibrosis [3–5].
hulmonary sarcoidosis and collagen vascular disorders (D) associated with interstitial lung disease represent diseases with very different cellular mechanisms and meal outcomes. Sarcoidosis is characterized by height-immunocellular activity affecting both alveolar cophages and lymphocytes (largely helper-inducer T but rarely leads to lung fibrosis [6]. In contrast, and involving the lung are frequently characterized by the neutrophilia, lymphocytosis, or a combination of [7–9]. Furthermore, CVD commonly leads to lung [18].

may contribute to architectural changes in the lungs

and given the often unpredictable clinical course of interstitial lung diseases (ILD), we decided to determine whether the proteolytic burden in the lower respiratory tract could vary among different types of ILD. To address this issue, we measured several polymorphonuclear neutrophil (PMN) and macrophage-derived proteases including plasminogen activator and PMN elastase in bronchoalveolar lavage (BAL) from patients with pulmonary sarcoidosis, CVD patients with ILD and CVD patients without ILD.

Methods

Study population

Nineteen normal volunteers, 42 patients with pulmonary sarcoidosis and 43 patients with collagen vascular disease were included in the present study. All were lifelong nonsmokers. Normal values were similar in the

three referral centres. The patients were referred to the University Hospital of Mont-Godinne, University of Louvain, Belgium, or to the University Hospital A. Calmette and Hôpital Régional, Lille, France or to the Yale University School of Medicine, New Haven, USA. The group of biopsy proven pulmonary sarcoidosis was further divided empirically into two subgroups based on BAL lymphocyte counts, namely lymphocytic sarcoidosis (LS) with BAL lymphocyte counts >15%, and nonlymphocytic sarcoidosis (NLS) with lymphocyte <15% in order to separate patients between respectively high and low intensity alveolitis as previously suggested [11]. In our hands, the 15% has been reported as a reasonable cut-off [12]. None of the patients had received steroids for at least one year. Three patients had previously been treated with steroids. Sarcoidosis groups LS and NLS, respectively, comprised 14 and 4 with hilar adenopathy alone, 7 and 7 with hilar adenopathy and also parenchymal infiltrates and 7 and 3 with infiltrates alone. All but one patients of those groups had normal lung function, i.e. no value of static or dynamic lung volumes or diffusing capacity of <80% predicted. The one abnormal LS sarcoid showed 60% predicted values for total lung capacity (TLC), forced vital capacity (FVC), forced expiratory volume in one second (FEV,) and pulmonary carbon monoxide diffusing capacity (DLco).

The CVD group totalled 43 persons and their features are shown in table 1. The diagnoses are based on previously described criteria [9].

Table 1. - Collagen vascular disorders

	Group I	Group II		Total
		Untreated	ed Treated	
Sjögren's syndrome	8	0	0	8
Rheumatoid arthritis	5	3	2	10
Scleroderma	6	7	4	17
Dermatomyositis	2	1	2	5
Lupus erythematosis	1	2	0	3

Group I: all had normal pulmonary function tests and normal chest X-rays. None had been treated with steroid. Group II showed parenchymal radiologic infiltrates. In group II, the untreated and treated groups, respectively, showed mean and ranges of % predicted values of TLC of 82% (50-105%) and 69% (47-90%), of FEV, of 76% (42-102%) and 71% (52-93%) and also of Dico of 66% (39-100%) and 66% (32-109%). There were no significant differences between these function tests in the two subgroups of collagen vascular disorders group II. Therapy comprised prednisone (30-40 mg daily) in all treated patients, one of whom also received cyclophosphamide.

Bronchoalveolar lavage

Bronchoalveolar lavage was performed in all normals and patients following the same standardized protocol using an Olympus BF-B3 Model fibreoptic bronchoscope [13, 14]. Briefly, after anaesthesia of the upper airways with lidocaine, the bronchoscope was wedged in a subsegmental bronchus and 5×50 ml aliquots of sterile 0.9% saline were instilled and then aspirated. Except for half of the normals, the first aliquot was discarded in no effect on protein and enzyme measurements analyses were performed on the pooled 4 or 5 Total cell and differential counts (on Cytospin tions) were performed prior to a 900 g centrifo separate the cell pellet from the supernatant in the normal volunteers, cell counts and cell dim were performed after the 900 g centrifugation on pellet. Part of the supernatant was concentrated in Amicon membrane (10,000 mw cut-off). Lavage ery volumes and volumes after concentration recorded and their ratio used to correct the mean of materials performed on concentrated lavage fluid those in the initial BAL fluid.

Albumin, alpha₁-protease inhibitor (Alpha₁P) alpha2-macroglobulin (Alpha2M) were measured unconcentrated BAL using an immunoradiometric (IRMA) with a sensitivity in the nanogram range and serum by immunonephelometry [14, 15]. Data we expressed per ml of unconcentrated lavage.

Enzyme assays

Neutrophil elastase (NE) in the concentrated BALL assayed by following the hydrolysis of a specific s strate, McOSuc-Ala,-Pro-Val-7-Amino-4-Methyl Coumarin (AMC) (Enzyme Systems Products, Livern CA) as described previously [16, 17]. Samples of 100; of concentrated BAL were added to 1 ml of buffer (0. M Tris at pH 7.5, with 0.5 M NaCl, 0.1 M CaCl, 10% DMSO) containing 0.1 M of the AMC suburn The fluorescence of the mixture was followed using a excitation wavelength of 370 nm and emission wave length of 460 nm on an Aminco-Bowns spectrofluorometer.

Plasminogen activator (PA) activity was also me ured on concentrated BAL according to SAKSEIA [18]. Agarose plates were prepared with a mixture of 34.4 m of 1.25% agarose in 0.1 M Tris HCl pH 8.0, 43 ml of 6% casein in 1% sodium azide and 4.3 ml of 0.1 M Tr HCl pH 8.0 with or without (for control) plasminoger $0.3~U\cdot ml^{-1}$. Sample wells (3.5 mm diameter) were pure into the gel. 10 μl of the concentrated BAL were locally the sample were locally than the sample were locally to the sam into the wells (each lavage sample being run in de cate) and the plates were incubated for 24 h at 37°C. diameters of the clear areas were measured and expressed in units of plasminogen activator according a standard curve. The lower detectable level of char was 0.01 U·ml-1. The specificity of the assay for PA conferred by the use of plasminogen.

The hydrolysis of succinyl-alanine,-nitrosnillar (SLAPN) was assayed at 410 nm using a spectropho ter as initially described by BIETH and WERMUTH [19]

detailed by Niederman et al. [20].

The effect of various inhibitors (table 2) on the against SLAPN present in BAL from 7 sarcold patients with high BAL lymphocytosis (mean lymphocytosis experience in high solutions in higher than the inhibitions in higher than the inhibition in high than the inhibition in higher than the inhibition in high than the inhibition in hig solutions in buffer at appropriate concentrations with BAL sample for 30 min at room temperature prior

Inhibition studies on SLAPN activity in

All and the same	EDTA 5 mM	PMN elastase° inhibitor 10 μΜ	Cathepsin L ⁺ inhibitor 10 μM	PMSF 5mM
100 miles	72.6*	10.3	12.6	12.7
	(n=7)	(n=4)	(n=5)	(n=4)

of the respective inhibitors, compared to the values in the absence of the inhibitor; n: number of BAL tested; PMN elatase inhibitor: (Mco-Suc-Ala₂-Pro-Ul); Cathepsin L inhibitor: (Z-Phe-Ala-Pro-Val-2005 compared to control by paired t-test.

control. All inhibitors were also tested alone in the sasay and showed no interference.

munoreactive neutrophil elastase-Alpha, PI bees were measured by ELISA purchased from (Darmstadt, Germany) [21]. This procedure toya antibodies specific to neutrophil elastase (NE) at a plastic tube. After BAL samples were incurvith this antibody, the tubes were washed, leaving lound to the tube walls. Enzyme-labelled (alkaline inplatase) antibody specific for the Alpha, PI was then at After these bound to the NE-Alpha, PI complex, excess labelled antibody was washed off and the tuning NE-Alpha, PI complex was assayed with 4-potential phosphate.

Statistics were performed by the NIH sponsored Clinfo system using the Wilcoxon Rank Sum Test (two-tailed) and Spearman Rank coefficients since much of the data showed different group variances and some were not normally distributed. Values of p≤0.05 were considered significant. Data are expressed as mean±sp.

Results

BAL cell composition

The cellular composition in the BAL fluids of normal subjects and patients with the two disorders are shown in table 3. BAL from sarcoidosis patients as a group were characterized by increased total cell and lymphocyte counts and by normal neutrophil (PMN) and eosinophil counts. CVD patients from group I and group II had elevated lymphocytes and PMN counts in BAL but total cell counts were only increased in group II (with lung disease). Eosinophils were only elevated in BAL from untreated group II CVD patients.

BAL enzyme constituents

The data on enzyme activities are expressed per ml of recovered BAL fluid. Since the initial and recovered lavage fluid volumes were no different in these groups, the data directly represents the quantity of these constituents recovered.

3 - Cellular components in BAL

	Age	Total cells*	% Mac	% Ly	% PMN	% Eos	
emils -19)***	33.7±7.8**	12.4±3.8	91.2±7.0	8.2±6.7	0.5±0.6	0.1±0.2	
sarcoid	36.6±13.5	43.1±57.3	70.5±10.6	28.0±10.2	0.6±0.8	0.4±0.7	
(4) sarcoid	44.2±15.0	22.5±12.5	91.7±3.8	7.6±3.8	0.6±0.8	0.1±0.2	
Nam vascular	49.0±15.0	25.0±26.2	72.7±17.0	18.2±16.0	7.4±8.8	1.6±3.4	
Sp1	46.6±15.9	18.6±27.6	75.9±14.5	19.3±13.8	4.4±3.7	0.4±0.8	
A Paraglia	48.7±16.0	32.1±19.3	71.5±16.5	17.8±18.2	8.6±6.4	2.1±2.4	
(a-5)	53.9±10.7	28.9±24.3	79.1±8.7	13.9±10.4	6.3±4.9	0.8±1.4	
Trailed	49.2±18.3	34.2±16.1	66.5±18.8	20.4±22.1	10.2±7.0	2.9±2.6	

10-ml⁻¹ recovered BAL. Instilled lavage volume and return was the same in all groups; **: mean±sp; ***: n= number measurements that were obtained on all subjects in each group; Mac: macrophages; Ly: lymphocytes; PMN: n= number neutrophils; Eos; eosinophils.

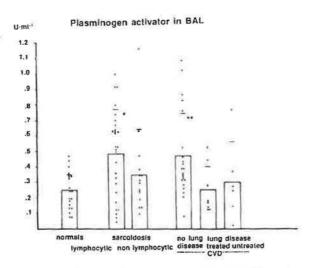


Fig. 1. – Plasminogen activator levels (U·ml·¹) in BAL. From left to right: normals (n=19); lymphocytic (BAL lymphocytosis >15%) (n=27) and non-lymphocytic (n=14) sarcoidosis patients and collagen vascular disease patients (CVD). The CVD group is subdivided into patients without lung disease (group I) (n=22) and patients with lung disease (group II). Group II includes treated (n=8) and untreated patients (n=6). Columns represent means, bars standard deviations. *: p<0.01; **: p<0.03 when compared to normals.

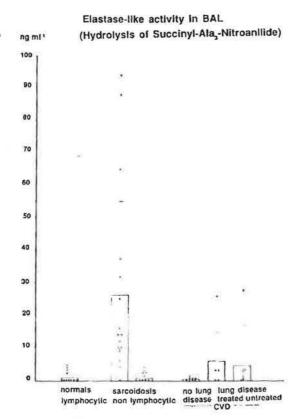


Fig. 2. – Elastase-like activity in BAL measured by hydrolysis of SLAPN substrate in the same groups as in figure 1. Normals (n=11), lymphocytic sarcoidosis (n=97), non-lymphocytic sarcoidosis (n=99), CVD group I (n=8) and treated (n=5) and untreated (n=8) CVD group II patients. Columns are means and bars standard deviations. *: p<0.001 compared to all other groups.

Plasminogen activator (PA) activity in BAL significantly increased in the sarcoidosis patients group (mean±sp: 0.44±0.28 U·ml·¹) compared to normal group (0.24±0.12), (p<0.01). However, and PA activity. When the sarcoidosis patients subdivided into two groups according to the BAL lymphocytosis, only the group with high laymphocyte counts (over 15%) had significantly high PA values (0.49±0.28) (p<0.01) as shown in figure 1 addition, PA activity was increased in BAL from Cygroup I (0.46±0.27) (p<0.01).

The activity against SLAPN was significantly increase in BAL from sarcoidosis patients as one group (mestable 17.5±26.1) compared to normals (1.3±1.8) (p-0.05). This increase was solely due to the patients with BAL lymphocytosis >15% (26.1±29.0) as illustrated in figure 2. In CVD, none of the groups show statistically significant differences from normal subjects, and only two patients exhibited values above the normal range Inhibition studies performed on BAL from sarcoidose patients containing proteolytic activity against SLAPN demonstrated significant inhibition (72.6%) with the metal chelator EDTA and minimal inhibition with inhibitors of serine proteases (table 2).

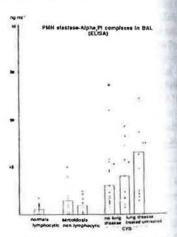


Fig. 3. – Neutrophil elastase activity against Meo-Suc-Ala, Provided Substrate in BAL. Groups similar to figures 1 and 2: Normal (n=19), lymphocytic sarcoidosis (n=25), non-lymphocytic sarcoidosis (n=24), CVD group I (n=22) and treated (n=7) and untreated (n=12 CVD group II patients. Values are significantly elevated in the uttreated CVD group II patients compared to the other groups (p-0.01)

Neutrophil elastase activities against AMC show a very different pattern to those observed with SLAPN. There was essentially no AMC activity in the normal subjects and the sarcoid groups. However, in the CVD groups detectable levels were present in lavages from 8-12 untreated group II patients, yielding an average activity of 1.5±1.3 ng·ml⁻¹ (fig. 3). Among treated group II cvD subjects, only 1 of 8 yielded such activity. This distribution is significantly different by Chi-square, p<0.05 in group I patients, activity was only detected in 1 of 2 lavages with a value of 6.8 ng·ml⁻¹ for inapparent reasons. Thus, untreated patients in CVD group II alone are associated with NE activity.

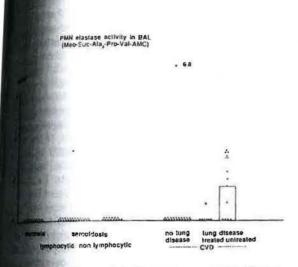
(fig. 4). By contrast, the CVD disorders show statistically significant higher levels. The combined groups I and II of CVD showed eight-fold increases (p<0.001) over the normal subjects. Even those CVD without radiologic or functional lung disease (group I) showed a four-fold increase (p<0.05), and in group II the elevated levels are more apparent in the untreated patients, as a group, (p<0.01 for 3 comparisons).

Albumin levels

The well described [14] increase in lavage albumin concentrations in LS sarcoid (p<0.001) but not NLS sarcoid are apparent (table 4): more interestingly, this is also apparent in all CVD disorders and in both groups I and II considered separately. The albumin concentrations in steroid treated group II CVD patients were significantly lower when directly compared with the untreated group II patients (p<0.01).

Antiproteases

These data are presented in table 4 as concentrations in lavage fluid rather than in coefficient of excretions relative to albumin (RCE), the form used previously [13] to attempt to correct for blood to lavage protein leak. This is appropriate since we are considering the protease antiprotease system in absolute terms. Thus, expressed in µg·ml⁻¹, both Alpha₁PI and Alpha₂M are significantly increased in LS sarcoid (p<0.001) but not NLS sarcoid.



Neutrophil clastase-alpha, PI complex levels in BAL (ELISA).

Interpretare similar to those in figures 1-3: normals (n=11), lympho-accoldesis (n=13), non-lymphocytic sarcoidesis (n=7), CVD group [15-14] and treated (n=8) and untreated (n=8) CVD group II patients [1605, **: p<0.01 when compared to normals.

Table 4. - Albumin and anti-protease levels in BAL

Albumin	Alpha ₂ M	Alpha, PI
26.0±19.9	0.04±0.03	1.55±1.25
90.0±94.0	1.20±1.55	6.60±5.43
31.0±16.7	0.08±0.08	1.25±1.23
46.2±35.8	0.80±1.50	3.70±4.10
37.9±21.2	0.30±0.85	2.57±3.44
54.9±44.8	1.44±1.84	5.02±4.54
36.4±25.6	0.63±0.47	2.51±3.52
67.2±51.4	1.78±2.11	6.07±4.62
	26.0±19.9 90.0±94.0 31.0±16.7 46.2±35.8 37.9±21.2 54.9±44.8 36.4±25.6	26.0±19.9 0.04±0.03 90.0±94.0 1.20±1.55 31.0±16.7 0.08±0.08 46.2±35.8 0.80±1.50 37.9±21.2 0.30±0.85 54.9±44.8 1.44±1.84 36.4±25.6 0.63±0.47

Data expressed as µg·ml-1 unconcentrated lavage (mean±sD)

strophil Elastase-Alpha, PI Complex measurement introduced at a later stage in this study; therefore, at the BAL materials were examined. However, at tubjects and both groups of sarcoidosis (despite states with increased levels) showed similar low of these complexes in the 1–2.5 ng·ml·l range

In group I CVD there is a marginal and nonsignificant elevation in Alpha, PI but a significant increase occurred in Alpha, M (p<0.05). However, group II CVD show three-fold rises in both antiprotease concentrations compared to group I (p<0.05). Moreover, in the group II CVD, both antiprotease BAL levels in the steroid treated

patients are at least half the levels found in the untreated patients (p<0.05 for both antiproteases).

Discussion

This study evaluates the presence of different phagocyte-derived enzymes in BAL from patients with interstitial lung disease and compares enzymatic measurements with the increases of the antiprotease levels reported in these disorders [12, 22].

Firstly, we measured the activity of plasminogen activator (PA) in BAL. This enzyme has been implicated in matrix degradation by human alveolar macrophages in vitro [23]. We have shown that PA was increased in BAL from sarcoidosis patients (mostly the ones with high BAL lymphocytosis) and group I CVD. Others [24] have reported diminished BAL PA levels in sarcoidosis and related this decrease in PA activity to an excess of PA inhibitor in sarcoidosis patient BAL fluid [25]. However, we generally studied recently diagnosed cases of pulmonary sarcoidosis, only one of which showed functional defects, while half the patients in the other study [24] had a disease duration of two years or more and 6 out of 14 had decreased FVC. Therefore, the apparent conflict in results may be related to patient selection, and reflects an increase in BAL PA activity early in disease with an association of increased levels of PA inhibitor(s) in chronic disease. This is supported by a recent study using an asbestos sheep model where acute inflammation was associated with high PA levels while chronic and fibrotic disease was associated with diminished PA levels in the lungs [26].

The assays employing SLAPN, AMC and NE-Alpha, PI complexes should be considered together and also in relation to the anti-protease measurements. While NE is active against both SLAPN and AMC, there has been considerable debate concerning the significance of SLAPN activity as a measure of free elastase activity [21, 27]. Our sarcoidosis data show major SLAPN activity totally unmatched by either AMC activity or NE-Alpha, PI.

This, along with the virtual absence of neutrophils in both sarcoid groups and normal nonsmoking subjects, strengthens the argument that SLAPN activity is not neutrophil-, but rather macrophage-derived, a view supported by previous studies [20, 27, 28]. The inhibition profile of SLAPN activity (inhibited by EDTA but not by Alpha, PI) suggests the presence of a metalloenzyme. Furthermore, there is an indication that the activity against SLAPN is not due to either free or complex forms of NE. A recent study suggests the activity against SLAPN reflects an endopeptidase capable of elastinolytic activity in co-operation with a metalloenzyme [29]. Thus, the nature and potential role of these enzymes is obscure in sarcoidosis. However, there is no compelling evidence that they alone attack native insoluble elastin in vivo.

A recent report [29] described enhanced activity against SLAPN in BAL cells of non-fibrotic sarcoid patient's. Since SLAPN activity has also been reported to be increased in the lavage fluid from smokers [20], it is likely

that enhanced activity against SLAPN heightened macrophage activity by either immunot mechanisms in sarcoidosis or by smoke-relative mechanisms.

Activity against AMC, generally accepted relatively specific substrate for NE, is absent in the But sarcoidosis groups, confirming previous data in nonsmoking subjects [28]. In CVD, the NE activities essentially present in only 8 of 12 untreated patients lung disease as opposed to only two of the remaining patients in groups I and II. This presumed neuroe elastase activity is paradoxically associated with increase of immunoreactive Alpha₂M levels, Assumithis Alpha₂M can still complex to NE, such complex in the lungs may remain active against low molecules weight substrates, as previously reported in animals whether such complexes are active in vivo against man components is, however, debatable [31].

When an immunologic assay was used to describe NE-Alpha, PI complexes, most subjects, including normal nonsmokers, had detectable levels. Except for two individuals, sarcoidosis patients had complex levels similar to normals. In contrast, all groups of CVD patients exhibited an increase in NE-Alpha, PI complexes with no difference among the subgroups, suggesting most of these patients, with the possible exception of a untreated group II patients, had NE complexed to Alpha, PI and, therefore, present in an inactive form in vivo. This also suggests there is heightened activity of elastase and anticlastase (Alpha, PI and Alpha, MI systems in the lung of CVD patients.

The balance between matrix degradation and collages deposition is believed to be critical in the pathogen of fibrotic processes. Lung fibrosis is often asso with neutrophil infiltration of the interstitium and always oli, and thus a persistent neutrophil derived protectives activity could contribute to tissue injury and indirect fibrosis [2, 32, 33]. On this basis, others found increase collagenase activity in BAL from idiopathic pulmon fibrosis (IPF) and rheumatoid arthritis patients [34,35 Our data show that NE activity is increased in group II CVD with lung disease characteristically fibrotic in natural In contrast, the heightened anti-protease levels (bod Alpha, PI and Alpha, M) and the undetectable NE activate against AMC in sarcoidosis, particularly the LS group shows the dominance of anti-protease activities in this "active sarcoid" group. This feature may be pathological cally significant for the much lower risk of fibrosis in sarcoidosis.

Steroid treatment has been reported to have little effection BAL PMN counts in IPF patients [36–38]. In a longitudinal study of CVD patients with ILD, we observe a decrease of the BAL PMN percentage without improvement in lung function tests [22]. In the present sectional study of nonsmoking patients, steroid treatment compared to their percentage of PMN in BAL compared to untreated patients with ILD. We also observed that PMN clastase activity in BAL was also ished in patients under corticosteroid treatment. This

space and a decrease in BAL eosinophilia, reduced inflammation in the lower respiratory unough we did not observe significant differences real, radiological or functional data between the treated and untreated CVD patients, a longitudiate is necessary to demonstrate that the observed are due to steroid treatment alone. However, in these changes, steroid treatment generally does duence clinically the fibrotic processes in the lungs. In the detect lung functional differences in these approups of group II CVD.

have shown that: a) sarcoidosis in the absence of throsis is characterized by the enhanced activity of macrophage derived enzymes, PA and an enterized metalloprotease active against a trialandromophore substrate, in addition to enhanced morase levels; b) CVDs are associated with modest mobilia and minor eosinophilia in BAL along with seed levels of the elastase-antiprotease system, relarly where CVD is associated with restrictive lung esteroid only untreated CVD with lung involvement is realed with detectable PMN elastase activity in BAL, suggests an inappropriate antielastase response in tower respiratory tract of these untreated patients with restrictive lung discase and could contribute to the lung esteroid in these disorders.

Acknowledgements: The authors wish to thank Dr K. Willard-Gallo for reviewing this manuscript, and Mrs M.P. Heylens for the excellent editorial work.

References

Junoff A. - Proteases and lung injury. A state-of-the-art

Crystal RG, Bitterman PB, Rennard SI, Hance AJ, Keogh Interstitial lung diseases of unknown cause. Disorders accerized by chronic inflammation of the lower respiratory N Engl J Med. 1984, 310, 154-166; 235-244.

McGuire WW, Spragg RG, Cohen AB, Cochrane CG. - dies on the pathogenesis of the adult respiratory distress rations. J Clin Invest, 1982, 69, 543-553.

Jeoff A. - Elastases and emphysema. Current assessment protease-antiprotease hypothesis. Am Rev Respir Dis, 132, 417-433.

MM, Crystal RG. – Analysis of cellular and protein of broncho-alveolar lavage fluid from patients with pulmonary fibrosis and chronic hypersensitivity monitis. J Clin Invest, 1977, 59, 165–175.

Crystal RG, Roberts WC, Hunninghake GW, Gadek JE, ID, Line BR. – Pulmonary sarcoidosis: a disease characted and perpetuated by activated lung T-lymphocytes. Ann. Med. 1981, 94, 73–94.

Silver RM, Metcalf JF, Stanley JH, Leroy EC. – Interstiling disease in scleroderma. Analysis by bronchoalveolar Arthritis Rheum, 1984, 27, 1254–1258.

Garcia JGN, Parhami N, Killam D, Garcia PL, Keogh BA.

Choalveolar lavage fluid evaluation in rheumatoid arthritis.

Respir Dis, 1986, 133, 450–454.

Wallacri B, Hatron PY, Grosbois JM, Tonnel AB, Devulder Voisin C. - Subclinical involvement in collagen-vascular

diseases assessed by bronchoalveolar lavage. Relationship between alveolitis and subsequent changes in lung function. Am Rev Respir Dis. 1986, 133, 574-580.

Huang CT, Chin T, Lyons HA. - Comparison of pulmonary function in patients with systemic lupus erythematosus, scleroderma and rheumatoid arthritis. Am Rev Respir Dis, 1966, 93, 865-875.

 Keogh BA, Hunninghake GW, Line BR, Crystal RG, – The alveolitis of pulmonary sarcoidosis. Evaluation of natural history and alveolitis-dependent changes in lung function. Am Rev Respir Dis, 1983, 128, 256–265.

 Sibille Y, Martinot JB, Staquet P, Delaunois L, Chatelain B, Delacroix DL. - Antiproteases are increased in bronchoalveolar lavage in interstitial lung disease. Eur Respir J, 1988, 1 498-505

 Reynolds HY, Newball HH. – Analysis of proteins and respiratory cells obtained from human lungs by bronchial lavage. J Lab Clin Med, 1974, 84, 559-573.

14. Delacroix DL, Marchandise FX, Francis C, Sibille Y. – Alpha₂-Macroglobulin, monomeric and polymeric immunoglobulin A, and immunoglobulin M in bronchoalveolar lavage. *Am Rev Respir Dis*, 1985, 132, 829–835.

15. Delacroix DL, Hodgson HJF, McPherson A, Dive C, Vaerman JP. – Selective transport of polymeric immunoglobulin A in bile: quantitative relationships of monomeric and polymeric immunoglobulin A, immunoglobulin M on other proteins in serum, bile and saliva. *J Clin Invest*, 1982, 70, 230-241.

 Castillo JC, Nakajima K, Zimmerman M, Powers JC. – Sensitive substrates for human leukocytes and porcine pancreatic elastase. A study of the merits of various chromophobic and fluorogenic leaving groups in assays for serine proteases. Analyt Biochem, 1979, 99, 53-64.

17. Sibille Y, Lwebuga-Mukasa JS, Polomski L, Merrill WW, Ingbar DH, Gee JBL. – An in vitro model for polymorphonuclear-leukocyte-induced injury to an extracellular matrix. Relative contribution of oxidants and elastase to fibronectin release from amnionic membranes. Am Rev Respir Dis, 1986, 134, 134–140.

 Saksela O. – Radial caseinolysis in agarose: a simple method for detection for plasminogen activator in the presence of inhibitory substances and serum. Analys Biochem, 1981, 111, 276-282.

 Bieth J, Wermuth CG. – The action of elastase on p-nitroanilide substrates. Biochem Biophys Res Commun, 1973, 53, 383-390.

Niederman MS, Fritts LL, Merrill WW, Fick RB, Matthay RA, Reynolds HY, Gee JBL. – Demonstration of a free elastolytic metalloenzyme in human lung lavage fluid and its relationship to alpha₁-antiprotease. Am Rev Respir Dis, 1984, 129, 943-947.

21. Neumann S, Henrich H, Gunzer G, Lang H. – Enzyme-linked immunoassay for human granulocyte elastase in complex with alpha, proteinase inhibitor. In: Proteases: Potential role in health and diseases. W. Horl, A. Heidland eds, Plenum Press, New York, London, 1984, 379–390.

22. Martinot JB, Wallaert B, Hatron PY, Francis C, Voisin C, Sibille Y. – Clinical and subclinical alveolitis in patients with collagen vascular disorders. Contribution of alpha-macroglobulin levels in BAL fluid. Eur Respir J, 1989, 2, 437-443

23. Chapman HA, Reilly JJ Jr, Kobzik L. - Role of plasminogen activator in degradation of extracellular matrix protein by live human alveolar macrophages. Am Rev Respir Dis, 1988, 137, 412-419.

24. Chapman HA, Allen CL, Stone OL. - Abnormalities in

pathways of alveolar fibrin turnover among patients with interstitial lung disease. Am Rev Respir Dis, 1986, 133, 437-443.

25. Chapman H, Bertozzi P, Singhal A, Astedt B. — Mechanism of reduced alveolar urokinase activity in patients with sarcoidosis. Am Rev Respir Dis, 1988, 137, 210 (abstract).

26. Cantin A, Allard C, Begin R. — Increased alveolar plasminogen activator in early asbestosis. Am Rev Respir Dis, 1989, 139, 604-609.

 Hinman L, Stevens CA, Matthay RA, Gee JBL. – Elastase and lysozyme activities in human alveolar macrophages. Am

Rev Respir Dis, 1980, 121, 263-271.

28. Janoff A, Raju L, Dearing R. - Levels of elastase activity in bronchoalveolar lavage fluids of healthy smokers and nonsmokers. Am Rev Respir Dis, 1983, 127, 540-544.

 Mordelet-Dambrine M, Lafuma C, Stanislas-Leguern G, Robert L, Chrétien J, Hornebeck W. – Elastase activity of bronchoalveolar cells in advanced pulmonary sarcoidosis. Eur

Respir J, 1988, 1, 748-757.

30. Stone PJ, Calore JD, Snider GL, Franzblau C. – Role of alpha-macroglobulin-elastase complexes in the pathogenesis of elastase-induced emphysema in hamsters. *J Clin Invest*, 1982, 69, 920–931.

31. Travis J, Salvesen GS. - Human plasma proteinase inhibitors. Ann Rev Biochem, 1983, 52, 655-709.

- 32. Senior RM, Campbell EJ. Neutral proteinases from human inflammatory cells. A critical review of their role in extracellular matrix degradation. Clin Lab Med, 1983, 3, 645-666.
- Haslam PL, Turton CW, Lukoszek HA, Salsbury A, Dewar JVC, Turner-Warwick M. Bronchoalveolar lavage fluids counts in cryptogenic fibrosing alveolitis and their relation to therapy. Thorax, 1980, 35, 328-339.

34. Gadek JE, Kelman JA, Weinberger SE, Horwitz AL, Reynolds HY, Fulmer JD, Crystal RG. – Collagenase in the lower respiratory tract of patients with idiopathic pulmonary fibrosis. N Engl J Med, 1979, 301, 737-742.

35. Weiland JE, Garcia JGN, Davis WB, Gadek JE. - Neutrophil collagenage in rheumatoid arthritis interstitial lung

disease. J Appl Physiol, 1987, 62, 628-633.

 Turner-Warwick M, Haslam PL. – The value of serial bronchoalveolar lavages in assessing the clinical progress of patients with cryptogenic fibrosing alveolitis. Am Rev Respir Dis, 1987, 135, 26-34.

 O'Donnell K, Keogh B, Cantin A, Crystal RG. –
 Pharmacologic suppression of the neutrophil component of the alveolitis in idiopathic pulmonary fibrosis. Am Rev Respir Dis, 1987, 136, 288-292.

38. Watters LC, Schwarz MI, Cherniack RM, Waldron JA,

Dunn TL, Standford RE, King TE. – Idiopathic pufibrosis. Pretreatment bronchoalveolar lavage cellular of ents and their relationships with lung histopatholoclinical response to therapy. Am Rev Respir Dis. 198 696-704.

Les enzymes phagocytaires dans les lavages broncho-alve de patients atteints de sarcoïdose pulmonaire et de colla Y. Sibille, J.B. Martinot, L. Polomski, B. Wallaeri, M. D.

J. Rankin, C. Voisin, B. Gee. RÉSUMÉ: L'équilibre entre les protéases et les anti-re du tractus respiratoire inférieur se voit attribuer un rôle développement des maladies interstitielles du poumon. cette étude transversale, nous avons mesuré divers e d'origine phagocytaire, notamment l'activateur du plasmi l'élastase neutrophilique, et une protéase mal définie ac un substrat de trialamine chromophore (SLAPN), d liquide de lavage broncho-alvéolaire chez 42 patients de sarcoïdose pulmonaire, et chez 43 patients ane collagénose (CVD), chez 22 sujets sans atte pulmonaire (groupe I) et chez 21 sujets atteints de l parenchymateuse pulmonaire (group II). Les observi montrent: a) que la sarcoïdose est associée à une activité de l'activateur du plasminogène et à la présence d'une ac enzymatique contre SLAPN, correspondant au moins en p à une métallo-protéase; b) que CVD, en l'absence d'are radiographique du poumon, est associée à une augmentation l'activité de l'activateur du plasminogène et de niveaux des complexes élastasiques alpha, antiprotéase-neutrophiles que la majorité des CVD non traitées (groupe II) ont des nive détectables d'activité élastasique des neutrophiles. C observations montrent que les patients atteints de sarcoi pulmonaire et de CVD ont des profils enzymatiques différe au niveau du tractus respiratoire inférieur lorsqu'on les son au lavage broncho-alvéolaire. Donc, la sarcoïdose, particulier sa forme lymphocytaire, est associée à une acti protéolytique d'origine macrophagique accrue dans le lave alvéolaire, alors que les patients CVD, avec ou sans mali pulmonaire, ont des décomptes de neutrophiles augmentés de l'elastase neutrophilique complexée à l'inhibiteur l'alpha, anti-protéase, et probablement inactifs dans le lava broncho-alvéolaire. Finalement, seul le lavage des patients à traités, atteints de CVD avec maladie pulmonaire interstiti contient une activité élastasique neutrophilique. Cette demi activité pourrait contribuer au développement des lésion pulmonaires fréquemment observées dans ces affections, Eur Respir J., 1990, 3, 249-256.